

Addition of Chiral and Achiral Allyltrichlorostannanes to Chiral α -Alkoxy Aldehydes

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Aliltricloroestananas quirais e aquirais reagem com α -alcóxi aldeídos quirais para fornecer alcoóis homoalílicos com moderados a bons níveis de diastereosseletividade 1,4-syn.

Achiral and chiral allyltrichlorostannanes reacted with chiral α -alkoxy aldehydes to give the corresponding homoallylic alcohols with moderate to good levels of 1,4-*syn*-diastereoselection.

Keywords: allyltrichlorostannanes, allylsilanes, homoallylic alcohols

Introduction

Allylsilanes and allylstannanes are among the most important groups of organometallic-type reagents available for the control of acyclic stereochemistry and their reaction with aldehydes in the presence of Lewis acids is an important procedure for the preparation of homoallylic alcohols.^{1,2} The addition of allylstannanes bearing a stereogenic center to chiral aldehydes is particularly interesting in organic synthesis. We recently communicated that *in situ* prepared chiral allyltrichlorostannanes react with chiral aldehydes to give 1,4-*syn* homoallylic alcohols with high levels of diastereoselectivity.³⁻¹¹

We wish to describe here a stereocontrolled reaction between achiral and chiral allyltrichlorostannanes with chiral lactate-derived aldehydes to give fragments which can be found in a large variety of naturally-occurring products with promising biological activities.¹² This study details our efforts to understand the double stereodifferentiating stereocontrol elements involved in chiral allyltrichlorostannane additions to chiral aldehydes.¹³

Results and Discussion

Achiral and chiral allylsilanes **1-4** were prepared from the corresponding easily available methyl esters, as described in previous papers from this laboratory (Scheme 1).^{3-11,14} According to previously established experimental procedures, the allylsilanes were mixed with $SnCl_4$ (1.0 equiv. in CH_2Cl_2) before the addition of a solution of the aldehyde in order to promote the $SiMe_3/SnCl_3$ exchange leading to the corresponding allyltrichlorostannanes **5-8** (Scheme 1).⁵

To the best of our knowledge, the first spectroscopic information available on exchange reactions involving allylsilanes and SnCl₄ was reported by Denmark and co-workers in 1988.¹³ In 1999, we described the first direct evidence for interaction between SnCl₄ and chiral allylic silane **3** bearing an ether functionality that generated a new species by means of NMR spectroscopy.⁵ In a continuation of these initial studies we have done a spectroscopic study (¹H and ¹¹⁹Sn NMR) of the reactions of allylsilanes **1-4** (0.15 molL⁻¹ solution in CDCl₃) with SnCl₄ leading to the corresponding allyltrichlorostannanes **5-8**, respectively (Scheme 1).

For allyltrimethylsilane 1 the SiMe₃/SnCl₃ exchange producing allyltrichlorostannane 5 and Me₂SiCl is complete after 2 h at room temperature (Scheme 1).⁵ For allylsilane 2 the SiMe₂/SnCl₂ exchange to give 6 and Me₂SiCl is faster, as expected for a 1,1-disubstituted electron-rich olefin, being complete after 10 minutes at room temperature.¹⁴ Upon addition of SnCl₄ to a solution of allylsilanes (R)-3 and (S)-4 in CDCl₂, at -60 °C, slightly yellow homogeneous solutions were obtained. The resulting NMR spectrum at -60 °C showed formation of Me₂SiCl and complete consumption of both allylsilanes within less than 1 minute to give allyltrichlorostannanes (R)-7 and (S)-8, respectively. It appears that the oxygen functionality is responsible for the rapid SiMe₂/SnCl₂ exchange reaction observed even at low temperatures for these particular allylsilanes and SnCl₄. The SiMe₃/SnCl₃ exchange is probably facilitated by coordination of tin to this oxygen followed by cleavage of the carbon-silicon bond by a free chloride ion.

Analysis of the corresponding ¹H NMR spectrum showed a deshielding for hydrogens H_1 to H_4 in allylstannane

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Scheme 1. SiMe₃/SnCl₃ exchange reaction of allylsilanes 1-4

5 when compared to the same signals for allylsilane **1** (Table 1).⁵

Table 1. ¹H NMR chemical shifts (δ /ppm) for 1 and 5

	$\begin{array}{c} H_1 \\ H_2 \\ H_3 \\ H_4 \\ H_4 \\ H_4 \\ H_4 \\ 1 \end{array}$		$\begin{array}{c} H_1 \\ H_2 \\ H_3 \\ H_4 \\ H_4 \\ H_4 \\ 5 \end{array} \\ \textbf{SnCl}_3 \\ \textbf{5} \end{array}$		
Compound	H_1	H_2	H ₃	H_4	
1	4.85	4.80	5.75	1.75	
5	5.45	5.35	5.95	3.25	

The same trend is observed for allylstannane **6** when compared to allylsilane **2** (Table 2).¹⁴

Table 2. ¹H NMR chemical shifts (δ /ppm) for 2 and 6^{14}



In the case of (*R*)-7, the deshielding of the hydrogens H_6 to H_9 in the ¹H NMR spectrum provides the best diagnostics (Table 3). The methylenic hydrogens H_6 and H_7 as well as the benzylic hydrogens H_8 and H_9 are too far away from the trichlorotin group to suffer from inductive effects. We believe that the deshielding observed for these hydrogens

in (*R*)-7 is due to the internal coordination of this oxygen to tin, as proposed in Table 3.⁵

Table 3. ¹H NMR chemical shifts (δ /ppm) for (*R*)-3 and (*R*)-7⁵



A similar behavior is observed for allylstannane (S)-8 when compared to allylsilane (S)-4 (Table 4).

Table 4. ¹H NMR chemical shifts (δ /ppm) for (S)-4 and (S)-8

H ₅ , Me Ph H ₆ H		H ₂ < H ₃ H ₄ SiMe ₃ S)- 4			H_{e} H_{e} $Ph \rightarrow H_{6}$	H ₁ 5.,. 0 H ₇	H₂ H₃ H₃ H₄ SnCl₃ (S)-8		
Compound	H_1	H_2	Н3	H_4	H_5	Η ₆	Н,	Me	
(S)- 4	4.80	5.00	1.26	1.32	3.84	4.52	4.59	1.30	
(S)- 8	5.01	5.05	2.91	3.05	4.17	4.65	5.10	1.28	

In addition, we have observed ¹¹⁹Sn resonance signals at -28 ppm for allylstannane **5** (Figure 1).⁵ The tin chemical shift for allylstannane (*R*)-**7** appeared at -187

ppm and for allylstannane (*S*)-**8** appeared at -169 ppm. The tin chemical shift for complexes **9** and **10** are -301 ppm and -599 ppm, respectively, while for free SnCl₄ it is -156 ppm. We believe that tin chemical shifts are highly sensitive to oxygen bonding, as observed for **9** and **10**, and the tin chemical shifts observed for (*R*)-**7** and (*S*)-**8** are strong evidence in favor of the proposed complexed intermediates.

The corresponding chiral aldehydes **11** and **12** were prepared in excellent yields from methyl lactate (Figure 2).^{15,16} These substrates have been selected to be representative of the complex fragments that might be coupled in polyacetate and polypropionate-derived aldol-type reactions. For aldehydes **11**, internal chelation is presumably prevented since, with few exceptions, silyl ethers are generally recognized for their poor coordinating and chelating abilities.¹⁷

In order to check the facial selectivities of aldehydes **11** and **12**, we reacted them with achiral allyltrichlorostannanes **5** and **6**. Achiral allyltrichlorostannane **5** reacted with chiral α -alkoxy aldehyde (*S*)-**11** in CH₂Cl₂ at -78 °C to give the corresponding 1,2-*syn* product **13** (*anti*-Felkin isomer) as the major isomer in 45% yield for the two-step sequence (preparation of the aldehyde from the ester and coupling reaction), with 60:40 diastereoselectivity (Scheme 2).^{18,19} Achiral allyltrichlorostannane **6** addition to the same aldehyde gave the corresponding 1,2-*syn* product **15** as the major isomer in 40% yield for the two-step sequence, again with 60:40 diastereoselectivity (Scheme 2). The stereoinduction observed in these reactions indicates

that the intrinsic facial bias imposed by the resident α -OTBS substituent results in preferential formation of the 1,2-*syn* diastereomer, with a small preference for the *anti*-Felkin type approach.¹⁹ One might project that the transition states of these reactions exhibit less charge separation than the aldol processes and are, accordingly, less subject to the electrostatic influence of the α -OTBS function.

The relative stereochemistry for the major product 13 was confirmed by comparison with data described in the literature.²⁰ In addition, we have also confirmed the relative stereochemistry for both 13 and 15 by analysis of the ¹H and ¹³C NMR chemical shifts for both syn and anti isomers, as described by Heathcock²¹ and Hoffmann²² for similar structures and applied to more complex substrates in this work. ¹H NMR and ¹³C NMR spectroscopy are very useful tools to study substituent effects on the electronic environment of a given carbon, as well as to determine the relative stereochemistry in acyclic molecules, especially by analysis of the coupling constants (J) in the corresponding ¹H NMR spectra. In the case of homoallylic alcohols 13-16, it is possible to assign the relative stereochemistry by ¹H and ¹³C NMR analysis, as these compounds, by adopting an internal hydrogen-bonded conformation, exhibit magnetically distinct NMR environments.

The intramolecular hydrogen bond leads to a 5-member ring in which the substituents are *trans* (13 and 15) or *cis* (14 and 16) and the predominance of hydrogen-bonded conformations should be reflected in different ¹H and ¹³C



Figure 1. ¹¹⁹Sn chemical shifts (Me₄Sn (0.0 ppm) is used as an internal reference)





Scheme 2. Addition of achiral allyltrichlorostannanes 5 and 6 to aldehyde (S)-11

chemical shifts (Table 5). In fact, very strong experimental evidence for the existence of intramolecular hydrogen bonds in alcohols 13-16 comes from the observed chemical shifts in the ¹H NMR and ¹³C NMR spectra measured in CDCl₂ (Table 5). We have shown previously that the intrinsic low basicity of silvl ethers does not affect the capacity of the oxygen attached to the silicon atom to form intramolecular hydrogen bonds.23 The 1H NMR spectra for compounds 13-16 are first order and the coupling constants (J) and chemical shifts (δ) are directly measured from the spectra. The ¹H NMR chemical shifts of H_a and H_b for both 1,2-syn isomers 13 and 15 are more shielded than the corresponding signals for H₂ and H₂ in 1,2-anti homoallylic alcohols 14 and 16. For alcohol 13 (R = TBS), the ¹H chemical shifts are 3.38 (Ha) and 3.70 (Hb), showing a trans orientation between these two hydrogens. For alcohol 14, the ¹H chemical shifts are 3.56 (Ha) and 3.78 (Hb), showing a cis orientation between these two hydrogens. The same trend is observed for syn and anti homoallylic alcohols 15 and 16.

In addition, the 13 C chemical shifts for the methyl group in *syn* compounds **13** and **15** are more deshielded when compared to the 13 C chemical shifts in **14** and **16**.

We next examined the stereochemical impact of a benzyl-protecting group at the oxygen in position α to the carbonyl aldehyde. Before starting the study described in Scheme 3, we expected that under conditions favouring internal chelation, the carbonyl facial bias of aldehyde (*S*)-**12** should be highly predictable. In fact, that proved to be the case. The facial bias of aldehyde (*S*)-**12** was determined after reaction with allyltrichlorostannane **5** in CH₂Cl₂ at -78 °C to give a 97:3 mixture of diastereoisomers **17** and **18**, in 45% yield over the two step sequence (Scheme 3).

This benzyl-protecting group imposes an intrinsic facial bias on the carbonyl that results in the formation of the

Table 5. ¹H NMR chemical shifts for homoallylic alcohols 13-16



1,2-*syn*-dioxygen relationship. This leads to higher levels of diastereoselection when compared to the use of a TBS protecting group.

The 1,2-*syn* relative stereochemistry for adduct **17** was confirmed by comparison of ¹H- and ¹³C NMR data as well as its optical rotation with literature values.²⁴

Previous work from our laboratory showed that allyltrichlorostannane (*R*)-7 reacted with achiral aldehydes leading to the formation of 1,4-*syn* products as the major isomers (up to > 95:5 diastereoselectivity).³⁻¹¹

At this point we initiated the double stereodifferentiating studies involving allyltrichlorostannane (*R*)-7 and chiral aldehydes **11** and **12**. Addition of allyltrichlorostannane (*R*)-7 to aldehyde (*S*)-**11** in CH_2Cl_2 at -78 °C gave an 85:15 mixture of diastereoisomers **19** and **20**, respectively, in 70% yield for the two-step sequence (Scheme 4).

Allyltrichlorostannane (*R*)-7 reacted with aldehyde (*R*)-11 to give 1,4-*syn*-1,2-*syn* product 21 as the major product in 55% yield (2 steps), although with only 70:30 diastereoselectivity (Scheme 4).

The facial bias of this chiral allyltrichlorostannane is dominated by the α -methyl stereocenter and tends to give the 1,4-*syn* isomer with *Si*-face attack, but the facial bias of this particular aldehyde is to give the 1,2-*syn* product. We were surprised with the result with aldehyde (*R*)-**11** as we were expecting a higher level of diastereoselection in favor of the product **21**.

The relative stereochemistry for the major products was determined after conversion to the corresponding dimethylacetonides (Scheme 5). Treatment of a mixture of **19** and **20** with TBAF in THF at room temperature gave diols **23** and **24** (67% yield), which was followed by reaction with 2,2-dimethoxypropane and catalytic amounts of camphorsulphonic acid (CSA) to give acetonides **25** and **26** in 40% yield after purification by flash column chromatography (Scheme 5).

The *cis*- acetonide **25** comes from the 1,2-*anti* isomer **19** and the *trans*- acetonide **26** originates from the corresponding 1,2-*syn* isomer **20**. The dimethyl groups (Me_a and Me_b) in both *trans* and *cis* dimethylacetonides are in different (average) chemical environments, giving



Scheme 3. Addition of achiral allyltrichlorostannane 5 to aldehyde (S)-12



Scheme 4. Addition of allyltrichlorostannane (R)-7 to aldehydes (S)-11 and (R)-11

rise to characteristic signals. As observed by Lombardo and coworkers²⁵ the difference in chemical shifts of the methyl groups (Me_a and Me_b) in the five member ring of the dimethylacetonides is larger for the *cis* isomer (0.12-0.14 ppm) when compared to the *trans* isomer (0.01-0.04 ppm).²⁵ In Figure 3 we can observe the partial ¹H NMR for *cis* and *trans* acetonides **25** and **26**.

There is a larger difference in chemical shifts for the methyl groups (Me_a and Me_b) in the *cis* isomer ($\Delta\delta = 0.12$ ppm) when compared to the chemical shifts for the same methyl groups of the *trans* isomer ($\Delta\delta = 0.05$ ppm). Based on this result we conclude that the 1,2-*anti* isomer **19** is the major product.²⁶

The relative stereochemistry for compounds **21** and **22** was determined based on the same strategy (Scheme 6).

As before, we observed that the most intense signals come from the *trans*- acetonide **30**, which in this case originates from the 1,2-*syn* adduct **21** (Figure 4).

We next moved to investigate the addition of allyltrichlorostannane (*R*)-7 to enantiomeric aldehydes 12 (Scheme 7). Allyltrichlorostannane (*R*)-7 reacted with chiral α -benzyloxy-aldehyde (*S*)-12 in CH₂Cl, at -78 °C to give the





Figure 3. Partial ¹H NMR for acetonides 25 and 26



Scheme 5. Preparation of acetonides 25-(cis) and 26-(trans)



Scheme 6. Preparation of acetonides 29-(trans) and 30-(cis)



1.47 1.46 1.45 1.44 1.43 1.42 1.41 1.40 1.39 1.38 1.37 1.36 1.35 1.34ppm

Figure 4. Partial ¹H NMR for acetonides 29 and 30

The coupling reaction between allyltrichlorostannane (*R*)-7 and aldehyde (*R*)-12 in CH_2Cl_2 at -78 °C gave a 63:37 mixture of diastereoisomers 33 and 34, in 60% yield for the two-step sequence.

It is interesting to point out that as the facial bias of this aldehyde is to give the 1,2-*syn* products, we expected a *matched* case and much higher levels of diastereoselectivity in the reaction of (R)-7 with (R)-12. Again, we were surprised to see that this was not the case.

The relative stereochemistries for both **31** and **33** were determined by applying the same methodology based on the ¹³C NMR chemical shifts described before for **13-16** (Table 5). The 1,2-*anti* isomer is the major product as we



Figure 5. 13 C NMR chemical shifts for Me_c in homoallylic alcohols 31-34

can observe from the ¹³C NMR chemical shifts for the more shielded Me_c (Figure 5). In the case of **31** and **32**, the ¹³C chemical shifts for Me_c in **31** appears more shielded (14.8 ppm) when compared to **32** (15.5 ppm).

In the case of **33** and **34**, we were able to confirm that the 1,2-*syn* is the major product, based on the ¹³C chemical shifts for Me_a in **33** (15.5 ppm) and **34** (14.5 ppm).

At this point, we turned our attention to the coupling reactions involving allyltrichlorostannane (*S*)-**8**. Addition of allyltrichlorostannane (*S*)-**8** to aldehyde (*S*)-**11** gave a 65:35 mixture of diastereoisomers **35** and **36** in 60% yield (2 steps) (Scheme 8).

We next examined the addition of the same allylstannane to the enantiomeric aldehyde (R)-11 affording a 75:25 mixture of isomers 37 and 38 in 47% yield for the two-step sequence.



Scheme 7. Addition of chiral allyltrichlorostannane (R)-7 to chiral aldehydes 12



Scheme 8. Addition of chiral allyltrichlorostannane (S)-8 to chiral aldehydes 11

Again, these reactions with α -OTBS aldehydes are characterized by poor levels of diastereoselectivity.

The relative stereochemistry for the major products was again determined based on the analysis of the ¹³C NMR chemical shifts of the corresponding 5-membered dimethylacetonides (Scheme 9). Treatment of **35** and **36** with TBAF at rt followed by treatment of the corresponding diols under acidic conditions with 2,2-dimethoxypropane gave acetonides **41** and **42**, respectively.

The ¹H NMR methyl resonances observed at 1.34 and 1.46 for **41** are characteristic of a *cis*-acetonide and ¹H NMR methyl resonances at 1.38 and 1.40 for **42** are consistent with a *trans*-acetonide (Figure 6).²⁵

The same strategy was applied to **37** and **38**, providing acetonides **45** and **46** (Scheme 10).

As can be seen from Figure 7, the *trans*-acetonide, which comes from the 1,2-*syn* adduct is the major isomer observed in this reaction.

Addition of allyltrichlorostannane (*S*)-**8** to aldehyde (*S*)-**12** at -78 °C in CH₂Cl₂, gave an 80:20 diastereoisomeric mixture in favor of the 1,2-*anti* isomer **47** in 70% yield for the 2 steps (Scheme 11). Addition of allyltrichlorostannane (*S*)-**8** to aldehyde (*R*)-**12** at -78 °C in CH₂Cl₂, led to a 60:40 diastereoisomeric mixture favoring the 1,2-*syn* isomer in 60% yield for the 2 steps (Scheme 11).

The selectivity in the latter case was somewhat disappointing, given the result observed in the reaction of aldehyde **12** with allyltrichlorostannane **5** (Scheme 3).

The relative stereochemistries for both **47** and **49** were determined by applying the same methodology described before for **13-16** (Table 5). The 1,2-*anti* isomer is the major product as can be observed from the ¹³C NMR chemical shifts for the more shielded Me_c (Figure 8). In the case of **49** and **50**, the ¹³C chemical shifts for Me_c in **50** appears more shielded (14.7 ppm) when compared to **49** (15.6 ppm).



Scheme 9. Preparation of acetonides 41-(cis) and 42-(trans)



Figure 6. Partial ¹H NMR for acetonides 41-(*cis*) and 42-(*trans*)



1.44 1.43 1.42 1.41 1.40 1.39 1.38

Figure 7. Partial ¹H NMR for acetonides 45-(trans) and 46-(cis)



Scheme 10. Preparation of acetonides 45-(trans) and 46-(cis)



Scheme 11. Addition of allyltrichlorostannane (S)-8 to chiral aldehydes 12



Figure 8. ¹H NMR chemical shifts for homoallylic alcohols 47-50

Conclusions

The examples presented in this work show that the levels of π -facial selection are dependent on the absolute stereochemistries of the aldehydes as well as of the allyltrichlorostannanes. The results from these experiments suggest that the stereochemical relationships between the allyltrichlorostannane and aldehyde substituents may confer either a reinforcing (matched) or opposing (mismatched) facial bias on the carbonyl moiety. One possible reason for this result could be attributed to the involvement of energetically similar chair and twist-boat transition states that lead to diastereomeric product formation. Another possibility to consider in these reactions is that nonbonded interactions between the allyltrichlorostannane and α substituents on the aldehyde may not be significant in pericyclic transition states leading to either Felkin or anti-Felkin addition products.¹³ We believe that this chemistry is significant in the context of acyclic diastereoselection and will prove to be useful in the synthesis of more complex molecules, like polyacetate and polypropionate-derived natural products.27,28

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Supplementary Information

Available free of charge at http://jbcs.org.br, as PDF file.

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- 26. Having confirmed the relative (*syn* or *anti*) relationship between allylstannane-derived stereogenic centers, the absolute stereochemistry of the newly formed hydroxyl substituent was determined by ascertaining its relationship to the stereocenter originating from the aldehydes, which are of known configuration.
- All new compounds were isolated as chromatographically pure materials and exhibited acceptable ¹H-NMR, ¹³C-NMR, IR, MS, and HRMS spectral data.
- 28. General procedure for allyltrichlorostannane coupling reactions: To a solution of 2.5 mmol of the corresponding allylsilane in 7 mL of dry CH_2Cl_2 at -78 °C was added 2.5 mmol of $SnCl_4$. The resulting solution was stirred at -78 °C for 30 min when 2.7 mmol of aldehyde in 2 mL of CH_2Cl_2 was added. This mixture was stirred at -78 °C for 1 h and quenched by the slow addition of 0.2 mL of Et_3N , followed by 10 mL of saturated NH_4Cl solution. The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (2x5 mL). The combined organic layer was dried (MgSO₄), filtered, and concentrated *in vacuo*. Purification by flash chromatography on silica gel (30% EtOAc/ hexanes) gave the corresponding homoallylic alcohols.

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Supplementary

Addition of Chiral and Achiral Allyltrichlorostannanes to Chiral α-Alkoxy Aldehydes

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General Informations: All reactions were carried out under an atmosphere of argon or nitrogen in flamedried glassware with magnetic stirring. Dichloromethane, triethylamine, 2,6-lutidine, diisopropylamine, dimethylformamide and N-methylpyrrolidone were distilled from CaH₂. Dimethyl sulfoxide was distilled under reduced pressure from calcium hydride and stored over molecular sieves. THF and toluene were distilled from sodium/benzophenone ketyl. Petrol refers to the fraction boiling between 40-60 °C. Purification of reaction products was carried out by flash chromatography using silica-gel (230-400 mesh). Analytical thin layer chromatography was performed on silica gel 60 and GF (5-40 µm thickness) plates. Visualization was accomplished with UV light and anisaldehyde, ceric ammonium nitrate stain or phosphomolybdic acid followed by heating or I₂ staining. ¹H-NMR spectra were taken in CDCl₃ at 300 MHz or at 500 MHz spectrometer and are reported in ppm using solvent as an internal standard (CDCl, at 7.26 ppm) unless otherwise indicated. Data are reported as (ap = apparent, s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, sext = sextet, ap t = apparent triplet, m = multiplet, br = broad, td = triplet of doublets, quint d = quintet of doublets, coupling constant(s) in Hz; integration. Proton-decoupled ¹³C-NMR spectra were taken in CDCl, at 75 MHz spectrometer and are recorded in ppm using solvent as an internal standard (CDCl₂ at 77.0 ppm) unless otherwise indicated.

Allylsilanes 2-4 (General Procedure): In a 3-necked 500 mL round bottomed flask powdered $CeCl_3.7H_2O$ (15.44 g, 41.4 mmol) was heated under vacuum (1 Torr) at 160 °C for 12 h with vigorous stirring, resulting in the formation of a mobile white solid. The reaction flask was flushed with argon and allowed to cool to rt when anhydrous THF (65 mL) was added to the vigorously stirred anhydrous cerium(III) chloride forming a uniform white suspension, which was kept under stirring for 2 h. During this time, a separate three-necked 100 mL flask, fitted with a condenser and a pressure-equalizing dropping funnel, was charged

with Mg turnings (1 g, 41.4 mmol), and the whole apparatus was flame dried under a flow of argon. To this flask was added dropwise a solution of ClCH₂SiMe₂ (5.8 mL, 41.4 mmol) in anhydrous THF (27 mL). This mixture was stirred for 3 h until almost all of the Mg had dissolved. The anhyd CeCl, suspension was now cooled to -78 °C. To this suspension was added dropwise the previously prepared Grignard reagent, forming an off-white suspension, which was stirred at -78 °C for 2 h. At this time, a solution of the corresponding ester (13.8 mmol) in anhydrous THF (8 mL) was added to the Grignard-cerium chloride complex dropwise over 5 min, and the resulting mixture was warmed gradually to r.t. When consumption of the starting ester was complete, as determined by TLC (3 h), the resulting grey solution was cooled to 0 °C and guenched by the addition of a sat. aq solution of NH₄Cl (30 mL). The organic layer was separated, and the aqueous layer was extracted with Et₂O (2 x 50 mL). The combined organic layers were washed with brine $(2 \times 50 \text{ mL})$ and dried (MgSO₄). The solvent was removed under reduced pressure to give a slightly yellow liquid that was dissolved in CH₂Cl₂ (100 mL). To this flask was added Amberlyst 15 (1.0 g) and this mixture was stirred at rt until complete consumption of starting material. The resin was then removed by filtration and washed with CH₂Cl₂ (100 mL). The solvent was removed under reduced pressure to give allylsilanes 2-4.



Trimethyl(2-methylenepentadecyl)silane (2): yellow oil; Yield: 68%; TLC: R_f 0.75 (EtOAc/hexane 20%); IR (Film): v 3072, 2953, 2926, 2854, 1633, 1466, 1248, 1157 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ 0.04 (s, 9H), 0.91 (t, J = 7.0 Hz, 3H), 1.29 (brs, 20H), 1.39-1.49 (m, 2H), 1.57 (s, 2H), 1.97 (t, J = 7.0 Hz, 2H), 4.52 (brs, 1H), 4.60 (d, J = 1.0 Hz, 1H); ¹³C NMR (CDCl₃, 75 MHz): δ 2.1 (CH₃), 14.2 (CH₂), 22.5 (CH₃), 22.8 (CH₂), 27.8 (CH₂), 29.4 (CH₂), 29.6 (CH₂), 29.8 (CH₂), 32.0 (CH₂), 37.9 (CH₂), 109.4 (CH₃), 146.2 (C₉).

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(*R*)-(4-(benzyloxy)-3-methyl-2-methylenebutyl) trimethylsilane (3): Yield: 88%; Rf = 0.38 (EtOAc/hexanes 5%); $[\alpha]_{D}^{22}$: +12.6 (*c* 1.3, CHCl₃); IR (film) v (cm⁻¹): 3069, 3030, 2957, 2851, 1632, 1497, 1453, 1414, 1364, 1247, 1158, 1097, 1031, 952, 852, 735, 696, 634 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 0.03 (s, 9H), 1.04 (d, *J* = 6.8 Hz, 3H), 1.46 (d, *J* = 13.6 Hz, 1H), 1.52 (d, *J* = 13.6 Hz, 1H), 2.28 (m, 1H), 3.26 (dd, *J* = 9.3, 8.3 Hz, 1H), 3.53 (dd, *J* = 9.3, 5.4 Hz, 1H), 4.52 (d, *J* = 12.1 Hz, 1H), 4.53 (d, *J* = 12.1 Hz, 1H), 4.62 (s, 1H), 4.64 (s, 1H), 7.25-7.40 (m, 5H); ¹³C NMR (CDCl₃, 75 MHz), δ (ppm): -1.3, 17.1, 26.6, 41.0, 72.9, 75.0, 106.5, 127.4, 127.5, 128.3, 138.7, 149.7; Elemental analysis: calcd. for C₁₆H₂₆OSi: C, 73.22%; H, 9.98%; found: C, 73.15%; H, 10.02%.



(*S*)-(3-(benzyloxy)-2-methylenebutyl)trimethylsilane (4): Rf = 0.34 (EtOAc/hexanes 5%); $[\alpha]_{D}^{22}$: +12.6 (*c* 1.3, CHCl₃); IR (film) v (cm⁻¹): 3068, 3023, 2951, 1720, 1603, 1495, 1454, 1248, 1093; ¹H NMR (CDCl₃, 300 MHz), δ (ppm): 0.08 (s, 9H), 1.30 (d, *J* = 6.6 Hz, 3H), 1.48 (d, *J* = 14.5 Hz, 1H), 1.62 (d, *J* = 14.5 Hz, 1H), 3.84 (q, *J* = 6.6 Hz, 1H), 4.35 (d, *J* = 11.7 Hz, 1H), 4.57 (d, *J* = 11.7 Hz, 1H), 4.80 (s, 1H), 5.00 (s, 1H), 7.25-7.40 (m, 5H); ¹³C NMR (CDCl₃, 75 MHz), δ (ppm): -0.8, 20.6, 21.1, 70.0, 78.8, 108.7, 127.2, 127.5, 128.2, 138.8, 147.6.

Allyltrichlorostannane (5): ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 3.06 (d, J = 5,2 Hz, 1H), 5.33 (d, J = 5.6 Hz, 1H), 5.40 (d, J = 6.6 Hz, 1H), 5.97 (m, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 35.0, 121.0, 127.0. Obs. The signal at 0.45 ppm corresponds to TMSCl.



Trichloro(2-methylenepentadecyl)stannane (6): ¹H NMR (CDCl₃, 300 MHz): δ 0.89 (t, *J* = 6.0 Hz, 3H), 1.30 (brs, 20H), 1.51 (m, 2H), 2.14 (t, *J* = 8.0 Hz, 2H), 3.15 (s, 2H), 5.07 (brs, 1H), 5.10 (brs, 1H). Obs. The signal at 0.45 ppm corresponds to TMSCI.



(*R*)-(4-(benzyloxy)-3-methyl-2-methylenebutyl) trichlorostannane (7): ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 0.95 (d, *J* = 7.0 Hz, 3H), 2.48 (m, 1H), 3.19 (d, *J* = 11.2 Hz, 1H), 3.36 (d, *J* = 11.2 Hz, 1H), 3.53 (dd, *J* = 9.9, 8.4 Hz, 1H), 3.70 (dd, *J* = 9.9, 4.4 Hz, 1H), 4.71 (d, *J* = 13.2 Hz, 1H), 4.77 (d, *J* = 13.2 Hz, 1H), 5.04 (s, 1H), 5.18 (s, 1H), 7.30-7.50 (m, 5H). Obs. The signal at 0.45 ppm corresponds to TMSCl; ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 15.7, 39.9, 42.7, 73.0, 74.5, 114.6, 127.5, 128.3, 128.7, 138.7, 144.0. Obs. The signal at 3.6 ppm corresponds to TMSCl.



(S) - (3 - (benzyloxy) - 2 - methylenebutyl) trichlorostannane (8): ¹H NMR (CDCl₃, 300 MHz, -60 °C) δ (ppm): 1.28 (d, J = 6.6 Hz, 3H), 2.91 (d, J =14.5 Hz, 1H), 3.05 (d, J = 14.5 Hz, 1H), 4.17 (q, J = 6.6Hz, 1H), 4.68 (d, J = 11.7 Hz, 1H), 5.01 (s, 1H), 5.05 (s, 1H), 5.10 (d, J = 11.7 Hz, 1H), 7.20-7.40 (m, 5H). Obs. The signal at 0.45 ppm corresponds to TMSCI.

Homoallylic Alcohols (General Procedure): To a solution of the corresponding allylsilane (1.5 mmol) in CH_2Cl_2 (5 mL) at rt was added $SnCl_4$ (1.1 mmol). The resulting solution was stirred at rt for 2 h and then cooled to -78 °C when a solution of aldehyde (1.2 mmol) in CH_2Cl_2 (2 mL) was added. This mixture was stirred for 2 h at -78 °C and quenched by the slow addition of a sat. aq solution of NaHCO₃ (5 mL) followed by CH_2Cl_2 (5 mL). The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (2 x 5 mL). The combined organic layers were dried (MgSO₄), filtered, and concentrated in vacuo. Purification by flash chromatography on silica gel (30% EtOAc–hexane) gave the corresponding homoallylic alcohols.



(2*S*,3*S*)-2-(*tert*-butyldimethylsilyloxy)hex-5-en-3-ol (13) and (2*S*,3*R*)-2-(*tert*-butyldimethylsilyloxy)hex-5-en-3-ol (14): Yield: 45%; R*f* = 0.26 (EtOAc/hexanes 5%); IR (film) ν (cm⁻¹): 3568, 3468, 2956, 2957, 2932, 2858, 1641,

1473, 1389, 1074, 1005, 968, 912, 777; ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 0.08 (s, 3H), 0.09 (s, 3H), 0.91 (s, 9H), 1.17 (d, J = 6.2 Hz, 3H), 2.18-2.32 (m, 2H), 3.33-3.43 (m, 1H), 3.70-3.82 (m, 1H), 5.01-5.16 (m, 2H), 5.81-5.59 (m, 1H). Minor isomer: 0.07 (s, 3H), 1.12 (d, J = 6.2 Hz, 3H), 3.53-3.60 (m, 1H), 3.76-3.82 (m, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): -4.7, -4.0, 18.2, 20.2, 25.9, 38.1, 70.9, 75.2, 116.8, 135.1; Minor isomer (**14**): 17.5, 36.8, 74.5, 117.2.



(2S,3S)-2-(tert-butyldimethylsilyloxy)-5methyleneoctadecan-3-ol (15) and (2S,3R)-2-(tertbutyldimethylsilyloxy)-5-methyleneoctadecan-3-ol (16): Yield: 40%; Rf = 0.36 (EtOAc/hexane 5%); IR (film) v (cm⁻¹): 3465, 3067, 2957, 2930, 2857, 1645, 1371, 1255, 1092, 835, 775; ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 0.08 (s, 6H), 0.88 (m, 3H), 0.89 (s, 9H), 1.16 (d, *J* = 6.0 Hz, 3H), 1.25 (brs, 22H), 1.36-1.49 (m, 2H), 2.01-2.18 (m, 2H), 3.49 (dq, *J* = 4.4 Hz, 6.6 Hz, 1H), 3.66-3.75 (m,1H), 4.81 (d, *J* = 4.7 Hz, 2H); Minor isomer: 0.06 (s, 6H), 1.12 (d, *J* = 6.0 Hz, 3H), 3.56-3.66 (m,1H); ¹³C NMR (CDCl₃, 63 MHz) δ (ppm): -4.8, -4.1, 14.1, 18.0, 22.7, 25.8, 27.7, 29.4, 29.7, 31.9, 71.0, 73.4, 111.3. Minor isomer: 20.0, 29.7, 73.0.



(2*S*,3*S*)-2-(benzyloxy)hex-5-en-3-ol (17) and (2*S*,3*R*)-2-(benzyloxy)hex-5-en-3-ol (18): Yield: 40%; R*f* = 0.44 (EtOAc/hexane 10%); IR (film) v (cm⁻¹): 3566, 3453, 3062, 3030, 2969, 2871, 1603, 1645, 1454, 1072, 1028, 993, 914, 737, 698; ¹H NMR (CDCl₃, 500 MHz) δ (ppm): 1.21 (d, *J* = 6.0 Hz, 3H), 2.15-2.26 (m, 1H), 2.31-2.40 (m, 1H), 3.44 (apqt, *J* = 6.2 Hz, 1H), 3.52 (ddd, *J* = 4.4 Hz, 6.2 Hz, 7.7 Hz, 1H), 4.38 (d, *J* = 11.7 Hz, 1H), 4.66 (d, *J* = 11.7 Hz, 1H), 5.09 (d, *J* = 9.5 Hz, 1H), 5.12(d, *J* = 20.3 Hz, 1H), 5.87 (ddt, *J* = 7.3 Hz, 9.5 Hz, 20.5 Hz), 7.26-7.38 (m, 5H). Minor isomer: 1.15 (d, *J* = 6.2 Hz, 3H); ¹³C NMR (CDCl₃, 63 MHz) δ (ppm): 15.4, 37.5, 71.0, 74.2, 117.2, 127.8, 128.4, 134.7, 138.3. Minor isomer: 13.8, 36.9, 70.7.



butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-

3-ol (19) and (2S,3S,6R)-7-(benzyloxy)-2-(*tert*butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-**3-ol (20):** Yield: 55%; Rf = 0.28 (EtOAc/hexanes 10%); IR (film) v (cm⁻¹): 3465, 3067, 2957, 2930, 2857, 1645, 1371, 1092, 835, 775; ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 0.08 (s, 6H), 0.9 (s, 9H), 1.07 (d, J = 7.0 Hz, 3H), 1.15 (d, J = 6.2 Hz, 3H), 2.07 (dd, J = 9.2, 14.3 Hz, 1H), 2.31 (dd, J= 4.0, 14.3 Hz, 1H), 2.50 (m, 1H), 3.39 (dd, J = 6.6, 9.2 Hz, 2H), 3.42 (dd, J = 7.0, 9.2 Hz, 2H), 3.63 (m, 1H), 3.72 (m, 1H), 4.93 (s, 1H), 4.95 (s, 1H), 7.26-7.32 (m, 5H); ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): -4.6, -4.2, 17.5, 18.1, 18.7, 25.9, 38.7, 39.3, 71.4, 73.0, 73.5, 74.5, 111.6, 127.4, 128.2, 138.2, 149.0.



(2 S, 6 R) - 7 - (b e n z y l o x y) - 6 - m e t h y l - 5 - methyleneheptane-2,3-diols (23) and (24): Yield: 67%;*Rf* $= 0.26 (EtOAc/hexanes 50%); ¹H NMR (CDCl₃, 300 MHz) <math>\delta$ (ppm): 1.02 (d, *J* = 7.0 Hz, 3H), 1.17 (d, *J* = 6.2 Hz, 3H), 2.13-2.29 (m, 2H), 2.51-2.55 (m, 1H), 3.39-3.51 (m, 2H), 3.70-3.77 (m, 1H), 3.84-3.92 (m, 1H) 4.51 (s, 2H), 5.00 (s, 2H); 5.30 (s, 2H), 7.28-7.38 (m, 5H); ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 17.5, 36.8, 39.0, 69.9, 72.0, 73.1, 74.3, 112.8, 127.8, 128.2, 137.8, 148,8.



(4R,5S)-4-((R)-4-(benzyloxy)-3-methyl-2methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (25) and (4S,5S)-4-((R)-4-(benzyloxy)-3-methyl-2methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (26): Rf = 0.72 (EtOAc/hexanes 20%); IR (film) v (cm⁻¹): 3047, 2986, 2934, 2872, 1645, 1454, 1377, 1223, 1080; ¹H NMR(CDCl₂, 300 MHz) δ (ppm): 1.02 (d, J = 6.4 Hz, 3H), 1.12 (d, J = 6.7 Hz, 3H), 1.31 (s, 3H), 1.49 (s, 3H), 2.07 (dd, J = 4.9, 15.6 Hz, 1H), 2.34 (dd, J = 8.4, 15.6 Hz, 1H), 2.07 (st, J = 6.7 Hz, 1H), 3.25 (dd, J = 7.2, 8.9 Hz, 1H), 3.43 (dd, J = 5.8, 8.9 Hz, 1H), 4.09 (qt, J =6.4 Hz, 1H), 4.21 (dt, J = 5.3, 8.5 Hz, 1H), 4.33 (s, 2H), 4,93 (d, J = 13.7 Hz, 2H), 7.28-7.38 (m, 5H); ¹³C NMR (benzene-d6, 75 MHz) δ (ppm): 16.1, 17.4, 26.0, 28.9, 35.9, 40.4, 73.1, 74.0, 75.1, 76.9, 107.4, 110.7, 126.5, 127.6, 128.5, 139.3, 149.3.



(2R, 3R, 6R) - 7 - (benzyloxy) - 2 - (tert butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (21) and (2R,3S,6R)-7-(benzyloxy)-2-(tertbutyldimethylsilyloxy)-6-methyl-5-methyleneheptan-**3-ol (22):** Rf = 0.53 (hexanes: EtOAc, 95:05): IR (film) v (cm⁻¹): 3463, 3031, 2950, 2857, 1645, 1559, 1497, 1455, 1255, 1092, 895; ¹H NMR (CDCl., 300 MHz) δ (ppm): 0.09 (s, 3H), 0.09 (s, 3H), 0.91 (s, 9H), 1.09 (d, J = 7.0Hz, 3H), 1.16 (d, J = 6.2 Hz, 3H) 2.03-2.35 (m, 2H); 2.50(qt, J = 7.0 Hz, 1H), 3.32-3.39 (m, 1H), 3.45-3.64 (m, 1H)2H), 3.72-3.80 (m, 1H), 4.52 (s, 2H), 4.91 (s, 1H), 4.94 (s, 1H), 7.33-7.34 (m, 5H).Minor isomer: 0.07 (s, 3H), 0.08 (s, 3H), 0.90 (s, 9H), 1.11 (d, J = 2.2 Hz, 3H); ¹³C NMR (CDCl₂, 75 MHz) δ (ppm): -4.8, -4.2, 17.4, 18.0, 18.3, 19.7, 25.8, 38.9, 39.4, 71.0, 73.0, 79.7, 74.6, 111.4, 127.5, 128.3, 138.4, 149.2. Minor isomer: 17.2, 38.5, 39.6, 71.4, 74.8, 79.7, 74.6, 111.8.



(2R, 6R) - 7 - (benzyloxy) - 6 - methyle-5 - methyleneheptane-2,3-diol (27) and (28): <math>Rf = 0.65(EtOAc/hexanes 50%); ¹H NMR (CDCl₃, 500 MHz) δ (ppm): 1.01 (d, J = 7.0 Hz, 3H), 1.18 (d, J = 6.0 Hz, 3H); 2.02-2.57 (m, 3H); 3.87-3.34 (m, 4H) 4.49 (d, J = 2.3 Hz, 1H), 4.97 (d, J = 2.3 Hz, 1H), 7.28-736 (m, 5H). Minor isomer: 1.08 (d, J = 7.0 Hz, 3H), 1.15 (d, J = 6.0 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ (ppm): 17.8, 19.0, 38.5, 40.1, 70.7, 73.2, 74.8, 113.0, 127.8, 128.4, 138.0, 148.8. Minor isomer: 17.3, 37.3, 39.7, 69.9, 75.0, 112.7, 127.7, 128.4.



(4R,5R)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane(29) and (4S,5R)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (30):Yield: 50%;*Rf*= 0.38 (EtOAc/hexanes 5%); IR (film) v(cm⁻¹): 3055, 2986, 2936, 2874, 1645, 1454, 1379, 1090, $898, 842; ¹H NMR (C₆D₆, 500 MHz) <math>\delta$ (ppm): 1.10 (d,

J = 6.1 Hz, 3H), 1.13 (d, *J* = 7.0 Hz, 3H), 1,40 (s, 6H), 2.13 (dd, *J* = 4.0, 15.0 Hz, 1H), 2.32 (dd, *J* = 7.6, 15.0 Hz, 1H), 2.55 (apsex, *J* = 6.4 Hz, 1H), 3.23 (dd, *J* = 7.3, 8.8 Hz, 1H), 3.42 (dd, *J* = 5.8, 8.8 Hz, 1H), 3.62 (dq, *J* = 5.8, 8.2 Hz, 1H), 3.68-3.72 (m,1H), 4.33 (s, 2H), 4.91 (s, 2H), 4.91 (s, 1H), 5.02 (s, 1H), 7.15-7.31 (m, 5H). Minor isomer: 1.11 (d, *J* = 6.0 Hz, 3H), 1.31 (s, 3H), 1.49 (s, 3H), 2.06 (dd, J = 4.6, 15.4 Hz, 1H), 4.20 (m,1H), 4.25 (m,1H), 7.08-7.11 (m, 5H); ¹³C NMR (C₆D₆, 125 MHz) δ (ppm): 17.2, 17.8, 27.5, 27.6, 39.0, 40.0, 73.1, 75.1, 77.3, 81.6, 111.5, 128.0, 128.5, 139.3, 149.0. Minor isomer: 16.0, 17.4, 26.0, 28.9, 35.7, 40.2, 74.0, 75.0, 76.4, 108.0, 110.8.



(2S, 3R, 6R) - 2, 7-bis (benzyloxy)-6-methyl-5-methyleneheptan-3-ol (31) and (2S, 3S, 6R)-2, 7bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (32): Yield: 70%; *Rf* = 0.36 (EtOAc/hexanes 20%); IR (film) v (cm⁻¹): 3454, 3061, 3026, 2970, 2872, 1643, 1498, 1454, 1367, 1264, 1090; ¹H NMR (CDCl₃, 250 MHz) δ (ppm): 1.04 (d, *J* = 7.0 Hz, 3H), 1.22 (d, *J* = 6.3 Hz, 3H), 2.09-2.53 (m, 3H), 3.33- 3.54 (m, 3H), 3.80-3.88 (m, 1H), 4.50 (s, 2H), 4.52 (d, *J* = 12.0 Hz, 1H), 4.63 (d, *J* = 12.0 Hz, 1H), 4.93 (s, 1H), 4.97 (s, 1H), 7.29-7.35 (m, 10H). Minor isomer: 1.09 (d, *J* = 7.0 Hz, 3H), 1.17 (d, *J* = 6.0 Hz, 3H), 3.62-3.72 (m, 1H), 4.00-4.10 (m, 1H); ¹³C NMR (CDCl₃, 63 MHz) δ (ppm): 14.8, 17.5, 39.0, 70.9, 73.0, 74.5, 112.0, 127.5, 127.7, 128.3, 138.1, 149.0. Minor isomer: 15.5, 17.2, 71.0, 71.7, 74.9, 138.7.



(2R, 3R, 6R) - 2, 7-bis(benzyloxy)-6-methyl-5methyleneheptan-3-ol (33) and (2R, 3S, 6R) - 2, 7bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (34): Yield: 60%; Rf = 0.18 (EtOAc/hexanes 10%); IR (film) v (cm⁻¹): 3695, 3055, 2976, 2930, 1715, 1452, 1072, 897; ¹H NMR (CDCl₃, 250 MHz) δ (ppm): 1.06 (d, J = 7.0 Hz, 3H), 1.21 (d, J = 6.0 Hz, 3H), 2.08-2.71 (m, 3H), 3.31-3.55 (m, 3H), 3.65-3.76 (m, 1H), 4.48 (d, J = 9.2 Hz, 1H), 4.51 (s, 2H), 4.66 (d, J = 11.3 Hz, 1H), 4.92 (s, 1H), 4.94 (s, 1H), 7.10-7.37 (m, 10H). Minor isomer: 1.20 (d, J = 6.3 Hz, 3H), 3.79-3.87 (m, 1H), 4.00-4.10 (m, 1H); ¹³C NMR (CDCl₃, 63 MHz) δ (ppm): 15.5, 17.4, 38.8, 39.1, 39.5, 71.1, 72.6, 74.5, 111.8, 127.7, 128.3, 138.3, 148.9. Minor isomer: 14.5, 17.2, 38.7, 70.8, 72.0, 74.8, 112.0, 126.0.



(2S, 3R, 6S) - 6 - (benzyloxy) - 2 - (tertbutyldimethylsilyloxy)-5-methyleneheptan-3-ol (35) and (2S,3S,6S)-6-(benzyloxy)-2-(tert-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (36): Yield: 35%; Rf = 0.17 (EtOAc/hexanes 5%); IR (film) v (cm⁻¹): 3564, 3445, 3052, 2955, 2931, 2858, 1651, 1454, 1372, 1092; ¹H NMR $(CDCl_{2}, 500 \text{ MHz}) \delta$ (ppm): 0.08 (s, 3H); 0.09 (s, 3H); 0.90 (s, 9 H); 1.16 (d, *J* = 6.2 Hz, 3H); 1.31 (d, *J* = 6.6 Hz, 3H); 2.03-2.34 (m, 2H); 3.55 (dq, J = 3.3, 1.5 Hz, 1H); 3.71-3.81 (m, 1H); 3.95-4.03 (m, 1H); 4.36 (d, J = 11.7 Hz, 1H); 4.56 (d, J = 11.7 Hz, 1H); 5.07 (s, 1H); 5.12 (s, 1H); 7.27-7.33(m, 5H). Minor isomer: 0.07 (s, 3H); 0.08 (s, 3H); 0.89 (s, 9 H); 1.13 (d, J = 5.9 Hz, 3H); 1.32 (d, J = 6.2 Hz, 3H);); 4.36 (d, J = 11.7 Hz, 1H); 4.53 (d, J = 11.7 Hz, 1H); 5.10(s, 1H); 5.14 (s, 1H). ¹³C NMR (C_6D_6 , 75 MHz) δ (ppm): -4.7, -4.2, 18.2, 19.2, 20.5, 26.0, 34.7, 70.2, 71.6, 74.9, 78.9, 113.7, 114.1, 127.8, 128.5, 139.2, 148.0. Minor isomer: -4.6, -4.3, 18.5, 21.2, 72.1, 74.6, 79.1.



(2R, 3R, 6S) - 6 - (benzyloxy) - 2 - (tertbutyldimethylsilyloxy)-5-methyleneheptan-3-ol (37) and (2R,3S,6S)-6-(benzyloxy)-2-(tert-butyldimethylsilyloxy)-**5-methyleneheptan-3-ol (38):** Yield: 47%; *Rf* = 0.25 (EtOAc/hexanes 10%); IR (film) v (cm⁻¹): 3564, 3435, 3052, 2960, 2931, 2862, 1647, 1454, 1371, 1090; ¹H NMR $(CDCl_2, 250 \text{ MHz}) \delta$ (ppm): 0.09 (s, 6H), 0.90 (s, 9 H), 1.16 (d, J = 6.3 Hz, 3H), 1.31 (d, J = 6.6 Hz, 3H), 2.03-2.49 (m, 2H), 3.57 (dq, J = 4.1, 1.3 Hz, 1H), 3.65-3.76 (m, 1H), 3.92-4.05 (m, 1H), 4.36 (d, J = 11.7 Hz, 1H), 4.52 (d, J = 11.7 Hz, 1H), 5.10 (d, J = 1.6 Hz, 1H), 5.14 (s, J = 1.6 Hz, 100 Hz)1H), 7.26-7.35 (m, 5H). Minor isomer: 0.08 (s, 3H), 0.89 (s, 9H), 1.15 (d, J = 6.3 Hz, 3H), 1.32 (d, J = 6.6 Hz, 3H),4.36 (d, J = 11.7 Hz, 1H), 4.53 (d, J = 11.7 Hz, 1H), 5.07 (d, J = 1.6 Hz, 1H); ¹³C (CDCl₃, 63 MHz) δ (ppm): -4.8, -4.2, 18.0, 20.0, 20.1, 25.8, 35.3, 70.0, 71.5, 74.3, 78.4, 113.2, 127.7, 128.4, 138.7, 147.2; Minor isomer: 18.6, 20.3, 34.3, 74.6, 114.1.



(2*S*,6*S*)-6-(benzyloxy)-5-methyleneheptane-2,3-diol (39) and (40): Yield: 67%; *Rf* = 0.36 (EtOAc/hexanes 50%); ¹H NMR (CDCl₃, 300 MHz) δ (ppm): 1.18 (d, *J* = 6.6 Hz, 3H), 1.32 (d, *J* = 6.6 Hz, 3H), 2.14-2.45 (m, 2H), 3.61 (dt, *J* = 3.7, 8.8 Hz, 1H), 3.70 (br, 2H), 3.79-3.87 (m, 1H), 4.00 (q, *J* = 6.6 Hz, 1H), 4.41 (d, *J* = 11.7, 1H), 4.56 (d, *J* = 11.7, 1H), 5.07 (s, 1H), 5.10 (s, 1H), 7.27-7.37 (m, 5H). Minor isomer: 1.19 (d, *J* = 6.0 Hz, 3H), 1.33 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 17.4, 19.9, 33.6, 70.1, 74.6, 78.7, 115.9, 127.7, 128.5, 137.8, 146.8. Minor isomer: 19.2, 19.4, 35.6, 70.3, 78.5.



(4R,5S)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (41) and (4S,5S)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3dioxolane (42): Yield: 50%; Rf = 0.55 (EtOAc/hexanes 20%); IR (film) v (cm⁻¹): 3055, 2986, 2934, 2872, 1647, 1454, 1371, 1086; ¹H NMR (C₆D₆, 300 MHz)δ (ppm): 1.01 (d, J = 6.2 Hz, 3H), 1.28 (d, J = 6.6 Hz, 3H), 1.31 (s, 3H),1.48 (s, 3H), 2.10-2.33 (m, 2H), 3.55-3.93 (m, 1H), 3.91 (q, J = 6.6 Hz, 1H), 4.06 (apquint, J = 6.2 Hz, 1H), 4.30(d, J = 12.0 Hz, 1H), 4.53 (d, J = 12.0 Hz, 1H), 5.10 (s, J = 12.0 Hz, 100 Hz)1H), 5.14 (s, 1H), 7.07-7.37 (m, 5H). Minor isomer: 1.08 (d, J = 5.9 Hz, 3H), 1.39 (s, 3H), 1.41 (s, 3H), 3.75 (ddd, 3H), 3H), 3H (s, 3H), 3H (s,J = 3.3, 8.4 Hz, 1H), 4.21-4.40 (m, 2H); ¹³C NMR (C₆D₆, 75 MHz) δ (ppm): 15.9, 20.6, 27.5, 27.6, 31.4, 70.1, 74.0, 76.9, 77.2, 81.1, 107.4, 112.9, 139.6, 147.1. Minor isomer: 17.6, 26.0, 28.9, 33.4, 78.9, 78.6, 108.0, 113.2.



(2*R*,6*S*)-6-(benzyloxy)-5-methyleneheptane-2,3-diol (43) and (44): Yield: 90%; *Rf* = 0.40 (EtOAc/hexanes 50%); IR (film) v (cm⁻¹): 3416, 3069, 2976, 2930, 2867, 1722, 1647, 1454, 1371, 1275, 1070; ¹H NMR (CDCl₃,

300 MHz) δ (ppm): 1.19 (d, J = 6.6 Hz, 3H), 1.34 (d, J = 6.6 Hz, 3H), 2.14-2.27 (m, 2H), 2.40 (d, J = 3.3 Hz, OH), 3.49-3.55 (m, 1H), 3.61 (apqt, J = 6.2 Hz, 1H), 4.01 (q, J = 6.6 Hz, 1H), 4.44 (d, J = 12.0 Hz, 1H), 4.52 (d, J = 12.0 Hz, 1H), 5.05 (s, 1H), 5.14 (s, 1H), 7.26-7.37 (m, 5H). Minor isomer: 1.18 (d, J = 6.6 Hz, 3H), 1.32 (d, J = 6.6 Hz, 3H), 2.44 (d, J = 3.3 Hz, OH), 3.80-3.88 (m, 1H), 5.11 (s, 1H); ¹³C NMR (CDCl₃, 75 MHz) δ (ppm): 19.3, 19.5, 35.7, 70.3, 74.3, 78.5, 115.4, 127.7, 128.4, 137.8, 145.9. Minor isomer: 17.5, 20.0, 74.6, 78.7.



(4R,5R)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (45) and (4S,5R)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3dioxolane (46): Yield: 90%; Rf = 0.27 (EtOAc/hexanes 5%); IR (film) v (cm⁻¹): 3053, 2963,2936, 2874, 1724, 1649, 1454, 1379, 1265, 1089, 912, 842; ¹H NMR (CDCl., 300 MHz) δ (ppm): 1.26 (d, J = 6.0 Hz, 3H), 1.31 (d, J = 6.6 Hz, 3H), 1.39 (s, 6H), 2.13-2.38 (m, 2H), 3.68-3.79 (m, 1H), 3.95-4.09 (m, 1H), 4.31-4.41 (m, 1H), 4.33 (d, J = 12.0 Hz, 1H), 4.50 (d, J = 12.0 Hz, 1H), 5.15 (s, 2H), 7.23-7.36 (m, 5H); Minor isomer: 0.99 (d, J = 6.6 Hz, 3H), 1.17 (d, J = 6.2 Hz, 3H), 1.35 (s, 3H), 1.46 (s, 3H), 5.06 (s, 2H); ¹³C NMR (CDCl₂, 75 MHz) δ (ppm): 17.9, 20.5, 27.6, 33.5, 70.2, 78.7, 81.2, 108.1, 113.7, 127.8, 128.5, 138.8, 146.6. Minor isomer: 16.1, 19.5, 26.1, 28.9, 30.8, 72.0, 74.0, 76.2, 112.9, 115.1.



(2*S*,3*R*,6*S*)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (47) and (2*S*,3*S*,6*S*)-2,6-bis(benzyloxy)-5methyleneheptan-3-ol (48): Yield: 70%; *Rf* = 0.55 (EtOAc/hexanes 20%); IR (film) v (cm⁻¹): 3695, 3055, 2986, 2930, 2685, 1715, 1603, 1452, 1265, 1072, 744; ¹H NMR (CDCl₃, 250 MHz) δ (ppm): 1.20 (d, *J* = 6.3 Hz, 3H), 1.32 (d, *J* = 6.6 Hz, 3H), 2.12-2.46 (m, 2H), 3.51 (ddd, *J* = 4.3, 6.3, 12.6 Hz, 1H), 3.87-4.13 (m, 2H), 4.37 (d, *J* = 12.0, 1H), 4.49 (d, *J* = 11.7 Hz, 1H), 4.51 (d, *J* = 11.7 Hz, 1H), 4.61 (d, *J* = 11.7 Hz, 1H), 5.08 (s, 1H), 5.14 (s, 1H), 7.26-7.37 (m, 10H). Minor isomer: 1.22 (d, *J* = 6.3 Hz, 1H); ¹³C NMR (CDCl₃, 63 MHz) δ (ppm): 14.1, 19.9, 34.1, 70.7, 72.0, 77.5, 78.5, 114.0, 127.6, 128.4, 138.6, 146.7.Minor isomer: 15.2, 20.3, 34.5, 70.0, 71.0, 73.3, 78.4, 79.4, 114.2, 127.5, 126.9, 138.5, 146.9.



(2*R*,3*R*,6*S*)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (49) and (2*R*,3*S*,6*S*)-2,6-bis(benzyloxy)-5methyleneheptan-3-ol (50): Yield: 88%; *Rf* = 0.55 (EtOAc/ hexanes 10%); ¹H NMR (CDCl₃, 250 MHz) δ (ppm): 1.22 (d, *J* = 6.3 Hz, 3H), 1.32 (d, *J* = 6.3 Hz, 3H), 2.11-2.42 (m, 1H), 3.42-3.54 (m,1H), 3.68-3.83 (m,1H), 3.93- 4.08 (m, 1 H), 4.34- 4.69 (m, 4H), 5.09 (d, *J* = 15 Hz, 1H), 5.14 (d, *J* = 5.4 Hz, 1H), 7.23-7.35 (m, 10H). Minor isomer: 1.23 (d, *J* = 6.3 Hz, 1H); ¹³C NMR (CDCl₃, 63 MHz) δ (ppm): 15.6, 20.2, 34.9, 70.0, 71.1, 73.4, 77.8, 78.4, 113.5, 127.7, 127.8, 128.3, 128.4, 138.6, 146.9. Minor isomer: 14.7, 34.7, 70.9, 73.0, 114.6, 127.4, 127.5, 138.3.



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 $\label{eq:Figure 1S. } Figure \ 1S. \ ^{1}H \ NMR \ (CDCl_{3}, \ 300 \ MHz) - (R) - (4 - (benzyloxy) - 3 - methyl - 2 - methylenebutyl) trichlorostannane \ (7).$

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Figure 2S. ¹³C NMR (CDCl₃, 75 MHz) - (*R*)-(4-(benzyloxy)-3-methyl-2-methylenebutyl)trichlorostannane (7).

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Figure 3S. ¹H NMR (CDCl₃, 300 MHz) - (S)-(3-(benzyloxy)-2-methylenebutyl)trichlorostannane (8).



Figure 4S. ¹H NMR (CDCl₃, 300 MHz) - Trichloro(2-methylenepentadecyl)stannane (6).



Figure 5S. ¹H NMR (CDCl₃, 300 MHz).



Figure 6S. ¹³C NMR (CDCl₃, 75 MHz).



Figure 7S. IR (film).



Figure 8S. ¹H NMR (CDCl₃, 300 MHz) - (*R*)-(4-(benzyloxy)-3-methyl-2-methylenebutyl)trimethylsilane (**3**).



Figure 9S. ¹³C NMR (CDCl₃, 75 MHz) - (S)-(3-(benzyloxy)-2-methylenebutyl)trimethylsilane (4).



Figure 10S. IR (film) - (S)-(3-(benzyloxy)-2-methylenebutyl)trimethylsilane (4).



Figure 11S. ¹H NMR (CDCl₃, 300 MHz).



Figure 12S. ¹³C NMR (CDCl₃, 75 MHz).



Figure 13S. IR (film).



Figure 14S. ¹H NMR (CDCl₃, 300 MHz) - (S)-(3-(benzyloxy)-2-methylenebutyl)trimethylsilane (4).



Figure 15S. ¹³C NMR (CDCl₃, 75 MHz) - (S)-(3-(benzyloxy)-2-methylenebutyl)trimethylsilane (4).



Figure 16S. IR (film) - (S)-(3-(benzyloxy)-2-methylenebutyl)trimethylsilane (4).



Figure 17S. ¹H NMR (CDCl₃, 300 MHz) - Trimethyl(2-methylenepentadecyl)silane (2).



Figure 185. ¹³C NMR (CDCl₃, 75 MHz) - Trimethyl(2-methylenepentadecyl)silane (2).



 $Figure \ 19S. \ IR \ (film) \ - \ Trimethyl(2-methylenepentadecyl) silane \ (2).$



Figure 20S. ¹H NMR (CDCl₃, 300 MHz).



Figure 21S. ¹³C NMR (CDCl₃, 63 MHz).



Figure 22S. IR (film).



Figure 23S. ¹H NMR ($CDCl_3$, 300 MHz) - (2S,3S)-2-(*tert*-butyldimethylsilyloxy)hex-5-en-3-ol (13) and (2S,3R)-2-(*tert*-butyldimethylsilyloxy)hex-5-en-3-ol (14).



Figure 24S. 13 C NMR (CDCl₃, 75 MHz) - (2S,3S)-2-(*tert*-butyldimethylsilyloxy)hex-5-en-3-ol (13) and (2S,3R)-2-(*tert*-butyldimethylsilyloxy)hex-5-en-3-ol (14).



Figure 25S. IR (film) - (2S,3S)-2-(tert-butyldimethylsilyloxy)hex-5-en-3-ol (13) and (2S,3R)-2-(tert-butyldimethylsilyloxy)hex-5-en-3-ol (14).



 $\label{eq:Figure 26S. 1} Figure 26S. \ ^{1}H \ NMR \ (CDCl_{3}, 300 \ MHz) - (2S, 3S) - 2 - (tert - butyldimethylsilyloxy) - 5 - methyleneoctadecan - 3 - ol \ (15) \ and \ (2S, 3R) - 2 - (tert - butyldimethylsilyloxy) - 5 - methyleneoctadecan - 3 - ol \ (16).$



Figure 278. ¹³C NMR (CDCl₃, 63 MHz) - (2S,3S)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneoctadecan-3-ol (15) and (2S,3R)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneoctadecan-3-ol (16).



Figure 285. IR (film) - (25,35)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneoctadecan-3-ol (15) and (25,3R)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneoctadecan-3-ol (16).



Figure 29S. ¹H NMR (CDCl₃, 500 MHz) (2S,3S)-2-(benzyloxy)hex-5-en-3-ol (17) and (2S,3R)-2-(benzyloxy)hex-5-en-3-ol (18).



Figure 30S. ¹³C NMR (CDCl₃, 63 MHz) - (2*S*, 3*S*)-2-(benzyloxy)hex-5-en-3-ol (17) and (2*S*, 3*R*)-2-(benzyloxy)hex-5-en-3-ol (18).



Figure 31S. IR (film) - (2S,3S)-2-(benzyloxy)hex-5-en-3-ol (17) and (2S,3R)-2-(benzyloxy)hex-5-en-3-ol (18).



 $\label{eq:Figure 325. } \ensuremath{^{1}\text{H}}\ \text{NMR}\ (\text{CDCl}_3, 300\ \text{MHz}) - (2S, 3R, 6R) - 7 - (benzyloxy) - 2 - (tert-butyldimethylsilyloxy) - 6 - methyl - 5 - methyleneheptan - 3 - ol\ (19)\ \text{and}\ (2S, 3S, 6R) - 7 - (benzyloxy) - 2 - (tert-butyldimethylsilyloxy) - 6 - methyl - 5 - methyleneheptan - 3 - ol\ (20).$



Figure 335. ¹³C NMR ($CDCl_3$, 75 MHz) - (2*S*, 3*R*, 6*R*)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (19) and (2*S*, 3*S*, 6*R*)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (20).



Figure 34S. IR (film) - (2S,3R,6R)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (19) and (2S,3S,6R)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (20).



Figure 35S. ¹H NMR (CDCl₃, 300 MHz) - (2S,6R)-7-(benzyloxy)-6-methyl-5-methyleneheptane-2,3-diols (23) and (24).



Figure 36S. ¹³C NMR (CDCl₃, 75 MHz) - (2*S*,6*R*)-7-(benzyloxy)-6-methyl-5-methyleneheptane-2,3-diols (23) and (24).



Figure 375. ¹H NMR (CDCl₃, 300 MHz) - (4R,55)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (25) and (4S,55)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (26).



 $\label{eq:Figure 385.13C} Figure 385. {}^{13}C NMR (CDCl_3, 75 MHz) - (4R,5S) - 4 - ((R) - 4 - (benzyloxy) - 3 - methyl - 2 - methylenebutyl) - 2, 2, 5 - trimethyl - 1, 3 - dioxolane (25) and (4S,5S) - 4 - ((R) - 4 - (benzyloxy) - 3 - methyl - 2 - methylenebutyl) - 2, 2, 5 - trimethyl - 1, 3 - dioxolane (26).$



Figure 395. IR (film) - (4R,55)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (25) and (45,55)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (26).



Figure 405. ¹H NMR (CDCl₃, 300 MHz) (2R,3R,6R)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (21) and (2R,3S,6R)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (22).



Figure 41S. ${}^{13}C$ NMR (CDCl₃, 75 MHz) - (2*R*,3*R*,6*R*)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (21) and (2*R*,3*S*,6*R*)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (22).



Figure 42S. IR (film) - (2R,3R,6R)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (21) and (2R,3S,6R)-7-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-6-methyl-5-methyleneheptan-3-ol (22).



Figure 438. ¹H NMR (C_6D_6 , 500 MHz) - (4*R*,5*R*)-4-((*R*)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (29) and (4*S*,5*R*)-4-((*R*)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (30).



Figure 44S. ${}^{13}C$ NMR (C_6D_6 , 125 MHz) - (4*R*,5*R*)-4-((*R*)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (29) and (4*S*,5*R*)-4-((*R*)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (30).



Figure 45S. IR (film) - (4R,5R)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (29) and (45,5R)-4-((R)-4-(benzyloxy)-3-methyl-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (30).



Figure 46S. ¹H NMR (CDCl₃, 250 MHz) - (2*S*,3*R*,6*R*)-2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (**31**) and (2*S*,3*S*,6*R*)-2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (**32**).



Figure 475. ${}^{13}C$ NMR (CDCl₃, 63 MHz) - (2*S*,3*R*,6*R*)-2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (31) and (2*S*,3*S*,6*R*)-2,7-bis(benzyloxy)-6-methyleneheptan-3-ol (32).



Figure 485. IR (film) - $(2S_3R_6R)$ -2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (31) and $(2S_3S_6R)$ -2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (32).



Figure 495. ¹H NMR ($CDCl_3$, 250 MHz) - (2R, 3R, 6R)-2, 7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (**33**) and (2R, 3S, 6R)-2, 7-bis(benzyloxy)-6-methyleneheptan-3-ol (**34**).



Figure 50S. 13 C NMR CDCl₃, 75 MHz) - (2R,3R,6R)-2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (33) and (2R,3S,6R)-2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (34).



Figure 51S. IR (film) - (2R,3R,6R)-2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (33) and (2R,3S,6R)-2,7-bis(benzyloxy)-6-methyl-5-methyleneheptan-3-ol (34).



Figure 528. ¹H NMR ($CDCl_3$, 300 MHz) - (2*S*,3*R*,6*S*)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (**35**) and (2*S*,3*S*,6*S*)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (**36**).



Figure 53S. ¹³C NMR (C_6D_6 , 75 MHz) - (2*S*,3*R*,6*S*)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (**35**) and (2*S*,3*S*,6*S*)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (**36**).



Figure 54S. IR (film) - $(2S_3R_6S)$ -6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (35) and $(2S_3S_6S)$ -6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (36).



Figure 558. ¹H NMR (CDCl₃, 250 MHz) - (2R,3R,6S)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (**37**) and (2R,3S,6S)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (**38**).



Figure 56S. ¹³C NMR (CDCl₃, 63 MHz) - (2R, 3R, 6S)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (37) and (2R, 3S, 6S)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (38).



Figure 575. IR (film) - (2R,3R,6S)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (37) and (2R,3S,6S)-6-(benzyloxy)-2-(*tert*-butyldimethylsilyloxy)-5-methyleneheptan-3-ol (38).



Figure 58S. ¹H NMR (CDCl₃, 300 MHz) - (2*S*,6*S*)-6-(benzyloxy)-5-methyleneheptane-2,3-diol (39) and (40).



Figure 59S. ¹³C NMR (CDCl₃, 75 MHz) - (2*S*,6*S*)-6-(benzyloxy)-5-methyleneheptane-2,3-diol (39) and (40).



Figure 60S. ¹H NMR (C_6D_6 , 300 MHz) - (4*R*,5*S*)-4-((*S*)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (41) and (4*S*,5*S*)-4-((*S*)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (42).



Figure 61S. ¹³C NMR (C_6D_6 , 300 MHz) - (4*R*,5*S*)-4-((*S*)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (41) and (4*S*,5*S*)-4-((*S*)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (42).



 $\label{eq:Figure 62S. IR (film) - (4R,5S) - 4 - ((S) - 3 - (benzyloxy) - 2 - methylenebutyl) - 2, 2, 5 - trimethyl - 1, 3 - dioxolane (41) and (4S,5S) - 4 - ((S) - 3 - (benzyloxy) - 2 - methylenebutyl) - 2, 2, 5 - trimethyl - 1, 3 - dioxolane (42). }$



Figure 63S. ¹H NMR (CDCl₃, 300 MHz) - (2*R*,6*S*)-6-(benzyloxy)-5-methyleneheptane-2,3-diol (43) and (44).



Figure 64S. ¹³C NMR (CDCl₃, 75 MHz) - (2*R*,6*S*)-6-(benzyloxy)-5-methyleneheptane-2,3-diol (43) and (44).



Figure 65S. IR (film) - (2*R*,6*S*)-6-(benzyloxy)-5-methyleneheptane-2,3-diol (43) and (44).



Figure 66S. ¹H NMR (CDCl₃, 250 MHz) - (4R,5R)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (45) and (45,5R)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (46).



Figure 675. ${}^{13}C$ NMR (CDCl₃, 63 MHz) - (4*R*,5*R*)-4-((*S*)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (45) and (4*S*,5*R*)-4-((*S*)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (46).



Figure 685. IR (film) - (4R,5R)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (45) and (45,5R)-4-((S)-3-(benzyloxy)-2-methylenebutyl)-2,2,5-trimethyl-1,3-dioxolane (46).



Figure 695. ¹H NMR ($CDCl_3$, 250 MHz) - (2*S*,3*R*,6*S*)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (47) and (2*S*,3*S*,6*S*)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (48).

Figure 705. ${}^{13}C$ NMR (CDCl₃, 63 MHz) - (2S,3R,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (47) and (2S,3S,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (48).

Figure 71S. IR (film) - (2S,3R,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (47) and (2S,3S,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (48).

Figure 72S. ¹H NMR (CDCl₃, 250 MHz) - (2R,3R,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (49) and (2R,3S,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (50).

Figure 735. 13 C NMR (CDCl₃, 63 MHz) (2*R*, 3*R*, 6*S*)-2, 6-bis(benzyloxy)-5-methyleneheptan-3-ol (49) and (2*R*, 3*S*, 6*S*)-2, 6-bis(benzyloxy)-5-methyleneheptan-3-ol (49).

Figure 74S. IR (film) - (2R,3R,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (49) and (2R,3S,6S)-2,6-bis(benzyloxy)-5-methyleneheptan-3-ol (50).