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In vitro activity of phospholipase A₂ and of peptides from *Crotalus durissus terrificus* venom against amastigote and promastigote forms of *Leishmania (L.) infantum chagasi*

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Abstract

Background: American visceral leishmaniasis is caused by the intracellular parasite *Leishmania (L.) infantum chagasi*, and transmitted by the sand fly *Lutzomyia longipalpis*. Since treatment is based on classical chemotherapeutics with significant side effects, the search for new drugs remains the greatest global challenge. Thus, this in vitro study aimed to evaluate the leishmanicidal effect of *Crotalus durissus terrificus* venom fractions on promastigote and amastigote forms of *Leishmania (L.) infantum chagasi*.

Methods: Phospholipase A₂ (PLA₂) and a pool of peptide fraction (<3 kDa) were purified from *Crotalus* venom. Furthermore, promastigotes and peritoneal macrophages of mice infected by amastigotes were exposed to serial dilutions of the PLA₂ and peptides at intervals varying between 1.5625 µg/mL and 200 µg/mL. Both showed activity against promastigotes that varied according to the tested concentration and the time of incubation (24, 48 and 72 h).

Results: MTT assay for promastigotes showed IC₅₀ of 52.07 µg/mL for PLA₂ and 16.98 µg/mL for the peptide fraction of the venom. The cytotoxicity assessment in peritoneal macrophages showed IC₅₀ of 98 µg/mL and 16.98 µg/mL for PLA₂ and peptide by MTT assay, respectively. In peritoneal macrophages infected by *Leishmania (L.) infantum chagasi* amastigotes, the PLA₂ stimulated growth of parasites, and at higher doses reduced growth by 23 %. The peptide fraction prevented 43 % of the intracellular parasite growth at a dose of 16.98 µg/mL, demonstrating the toxicity of this dose to macrophages. Both fractions stimulated H₂O₂ production by macrophages but only PLA₂ was able to stimulate NO production.

Conclusion: We have demonstrated the in vitro leishmanicidal activity of the PLA₂ and peptide fraction of *Crotalus* venom. The results encourage further studies to describe the metabolic pathways involved in cell death, as well as the prospecting of molecules with antiparasitic activity present in the peptide fraction of *Crotalus durissus terrificus* venom.

Keywords: PLA₂, Peptides, *Crotalus durissus terrificus*, Venom, Leishmanicidal activity

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Background

The epidemiological relevance of American visceral leishmaniasis, caused by the parasite *Leishmania (L.) infantum chagasi*, as a neglected disease has increased significantly in Latin America and especially in Brazil, where 90 % of the cases occur [1, 2]. Transmitted by the bite of *Lutzomyia longipalpis*, infected with promastigote forms, dogs are considered the principal reservoir for humans [1, 3–5].

Its treatment is carried out via chemotherapeutics such as amphotericin B and pentamidine as well as meglumine antimonate and sodium stibogluconate, all of which present high indices of serious side effects [6–8]. The infection triggers a cluster of clinical manifestations that can lead to death, if the patient is not treated. The toxicity of drugs, the difficulties of administration and the duration of treatment, allied with their low efficacy for humans and practical inefficacy for other animals, has stimulated the research of new natural leishmanicidal compounds, such as those derived from animal venoms [9].

Snake venoms are composed of a complex mixture of distinct proteins and present different pharmacological activities that can generate different active molecules [10]. The venom of *Bothrops mojeeni* can inhibit the growth of *Leishmania* spp. due to the hydrogen peroxide generated by the activity of the enzyme L-amino acid oxidase [11]. Venoms from *Cerastes*, *Vipera*, and *Naja* also exert an inhibitory effect on growth of *Leishmania infantum donovani* [12]. The venom of *Crotalus durissus cascavella* has three main fractions that present antileishmanial activity. The South American rattlesnake *Crotalus durissus terrificus* (Cdt) possesses several biologically active proteins in its venom, such as giroxin, crotamine, convulxin, crotoxin and peptides.

Crotoxin, the major toxic component of Cdt venom, is constituted by two subunits, an acid one without enzymatic action called crotoptin, and a basic one that presents the function of phospholipase A₂ (PLA₂), which possesses neurotoxic and myotoxic action [13, 14]. Evaluations of the in vitro action of PLA₂ from snakes of the genus *Bothrops* against promastigote forms of *Leishmania* sp. have obtained encouraging results against these parasites [15–17]. Another important component of Cdt venom is its peptide fraction, which is composed of low-molecular-weight molecules. Despite previous descriptions of this peptide fraction having presented anesthetic action and natriuretic peptides, neither of these two aspects has been evaluated against *Leishmania*.

Considering that snake venoms constitute a great source of bioactive protein substances that can act on diverse physiological systems, the present work aimed to evaluate the action of PLA₂ and of the peptide fraction from *Crotalus durissus terrificus* snake venom against

Leishmania (L.) infantum chagasi, in in vitro infection of murine macrophages and in a culture of promastigotes.

Methods

Crotalus durissus terrificus (Cdt) venom was kindly provided by the Center for the Study of Venoms and Venomous Animals, UNESP, Botucatu, Brazil. The entire procedure followed the ethical principles for animal experimentation adopted by the Brazilian College of Animal Experimentation (COBEA), and was approved by the Ethics Committee for Animal Experimentation at the Botucatu Medical School, UNESP, CEEA 890-2011.

Protein quantification was determined by the Bradford method [18] using bovine serum albumin (BSA) as the molecular standard (Sigma, USA). Electrophoresis was carried out under denatured conditions on 13 % (m/v) polyacrylamide, in order to evaluate the purity of the samples and the molecular masses of the proteins isolated [19].

The purification of PLA₂ was performed utilizing the Äkta Explorer 100 (GE HealthCare®) by three chromatographic steps. In the first two, the chromatographic strategy adopted was molecular exclusion, utilizing the column Hiprep 26/60/Sephacryl S-100 HR (2.6 × 60 cm, GE HealthCare®). The chromatographic reading was done with a flow of 1 mL/min under absorbance of 280 nm. One gram of lyophilized Cdt venom was eluted in 50 mM of ammonium formate and 150 mM of NaCl, pH 3.5. The fraction eluted between the volumes 40 and 60 mL was collected and lyophilized, being rechromatographed under the same described conditions. Next, the fraction eluted between the volumes 42 and 52 mL was collected and lyophilized again. The third step, namely purification, employed an ion-exchange strategy by means of the Hitrap DEAE column (0.7 × 2.5 cm, 1 mL, GE HealthCare®) previously equilibrated in a 6 M buffer of urea and 50 mM of Tris, pH 7.3. A 100 µL solution with a concentration of 2.7 µg/mL of purified protein fraction diluted in equilibrium buffer was injected at a flow rate of 0.5 mL/min and absorbance of 280 nm, under a gradient from 0 to 100 % of buffer of 6 M urea, 50 mM Tris and 1 M NaCl. After 27 min, the fraction of interest was collected [20].

PLA₂ was identified by phospholipase activity [16] and by sequencing of amino acids from their N-terminal portion by the Edman Degradation Chemistry technique [21] using automated equipment, model PPSQ-21 (Shimadzu®), following the manufacturer's standard instructions. SDS-PAGE was performed under reducing or non-reducing conditions using a 13.5 % polyacrylamide gel [16]. These procedures were employed to evaluate whether the protein isolated was of interest for the present study.

To obtain the pool of peptide fraction (<3 kDa), 300 mg of lyophilized Cdt venom was diluted in 2 mL of 0.1 % TFA (v/v) and homogenized. The supernatant was placed in a spin concentrator, model Viva Spin 2-3000 MW (GE HealthCare®) and centrifuged at 3000 rpm for 30 min. This process was repeated three times until the entire peptide fraction had passed through the concentrator [22].

The *L. (L.) infantum chagasi* promastigotes (MHOM/BR/1972/LD) were maintained in agar-blood biphasic medium, Novy-MacNeal-Nicolle (NNN), associated with liver infusion tryptose (LIT) medium supplemented with 10 % (v/v) bovine fetal serum (BFS), 10 mg/mL of gentamicin in a humid incubator (Biologic Oxygen Demand TE-381) at 25 °C, containing 5 % CO₂ [23]. Subculturing was carried out every 15 days. The macrophages were obtained from the peritoneal cavity of BALB/c mice by washing with 3 mL of culture medium RPMI-1640 (Roswell Park Memorial Institute) supplemented with 10 % (v/v) BFS and were maintained at 37 °C in an atmosphere of 5 % CO₂ [24].

To determine the 50 % inhibitory concentration (IC₅₀) against promastigotes of *L. (L.) infantum chagasi* in the latest logarithmic phase of growth, the cultures were washed three times in phosphate buffer (PBS) and their concentration was adjusted to 10⁶/mL, starting from the counting performed in a Neubauer chamber. Subsequently, culture aliquots containing 5 × 10⁵ parasites were applied in 96-well plates with LIT medium supplemented with 10 % BFS. The PLA₂ and the peptide fraction of the Cdt venom were dissolved in PBS, and serially diluted starting from an initial concentration of 200 µg/mL, with dilution factor 2, that were next added individually to cultures. Amphotericin B was employed as positive control (0.890 µg/mL) and PBS as negative control.

The in vitro susceptibility of *Leishmania (L.) infantum chagasi* promastigotes treated with PLA₂ and the peptide fraction of Cdt venom was determined by the colorimetric method and by studying the mitochondrial oxidative activity using MTT tetrazolium salt at a concentration of 5 mg/mL, diluted in PBS, sterilized in a 0.22 µm methylcellulose membrane (Millipore®). Then, 20 µL per well was added, which was incubated for 4 h at 24 °C. The isolated incubated promastigotes were utilized as a viability control. The reenzymatic action was analyzed, whereas the precipitate was solubilized by the addition of 80 mL of SDS 10 % (m/v) dissolved in Milli-Q water; the optical density (OD) was determined by utilizing a plate reader (Multiskan MS, Uniscience®) at 550 nm. The definition of 100 % viability was based on the OD of the control [24].

To determine the 50 % inhibitory concentration (IC₅₀) against intracellular amastigotes of *Leishmania (L.) infantum chagasi*, peritoneal macrophages were obtained

from BALB/c mice, and the cellular concentration was adjusted to 8 × 10⁵ cells/mL. Next, the cell suspension was incubated on 16-well Nunc® microculture plates, (System Deslize™ Lab-Tek Chamber, USA) coupled with a histological slide, at the proportion of 4 × 10⁵ cells/well and maintained at 37 °C in an atmosphere of 5 % CO₂ for a period of 24 h so that the macrophages would adhere to the plate. The supernatant was removed and added to RPMI-1640 medium supplemented with 10 % BFS. After this period, PLA₂ and the peptide fraction of the Cdt venom were dissolved in PBS, and serially diluted starting from an initial concentration of 100 µg/mL, with dilution factor 2, that were next added individually to cultures. Untreated infected macrophages were used as control. The assays were performed in triplicate, and evaluated after 24, 48 and 72 h by optical microscopy. The slides were fixed with methanol and stained with Giemsa. IC₅₀ was determined by counting 500 macrophages per well and evaluating the number of infected macrophages. Amphotericin B was employed as positive control (IC₅₀ 0.890 µg/mL) and PBS as negative control [25].

To evaluate the cytotoxicity in mammalian cells, the peritoneal macrophages (4 × 10⁵ cells/well) were incubated with 200 µg/mL of PLA₂ and with 100 µg/mL of peptide fraction from Cdt and serially diluted at 37 °C in an atmosphere of 5 % CO₂ for a period of 24 h and 48 h [26, 27]. Amphotericin B (0.890 µg/mL) was employed as positive control. Mammalian cellular viability was determined by the MTT assay using the microplate reader MS Multi-skan (Uniscience®) at 550 nm.

For the determination of nitric oxide (NO) production, the infected macrophages were incubated in microtubes containing different concentrations of PLA₂ and of the peptide fraction (200 µg/mL, 100 µg/mL, 50 µg/mL, 25 µg/mL, 12.5 µg/mL, 6.25 µg/mL, 3.125 µg/mL, 1.5625 µg/mL) for 24, 48 and 72 h. After each incubation, the recollected supernatant was applied into 96-well microplates for measurement of nitrite (NO₂) and nitrate (NO₃). The nitrite concentration was verified by the reaction of Griess. The results were obtained by means of MS Multiskan microplates (Uniscience®) at 540 nm, in comparison to a zero standard consisting of Griess reagent. All the measurements were performed in triplicate and the results were expressed in micromoles of NO/2 × 10⁵ cells, starting from the standard curve established by known concentrations of molarities from 0.39 to 200 µM NO₂. The control was constituted by cells treated with lipopolysaccharide (LPS), and by untreated cells [28, 29].

The production of H₂O₂ was evaluated by incubation of peritoneal macrophages with different concentrations of PLA₂ and of the peptide fraction (200 µg/mL, 100 µg/mL, 50 µg/mL, 25 µg/mL, 12.5 µg/mL, 6.25 µg/mL,

3.125 µg/mL, 1.5625 µg/mL), for 24, 48 and 72 h. Added to the macrophages after each incubation period was 100 µL of a buffer containing 140 mM NaCl, 10 mM phosphate, pH 7, 5.5 mM Dextrose, 0.56 mM phenol red and 0.01 mg/mL peroxidase Type II (Sigma Chemical Co, USA) in each well. The plates were incubated at 37 °C, in an atmosphere of 5 % CO₂ for 4 h, and the reaction was stopped for the addition of 0.01 mL of 1 N NaOH. The absorbance was determined in an automatic microplate reader (MD 5000[®], Dynatech Laboratories Inc., USA) with a 620 nm filter in relation to a “zero standard” consisting of phenol red and 1 N NaOH. All measurements were performed in triplicate whereas H₂O₂ production was expressed in nanomoles/2 × 10⁵ cells, according to the standard curve established for each test. Under the experimental conditions, the curve was generated with the H₂O₂ concentrations of 0.5; 1.0; 2.0; 4.0 and 8.0 mM. The control was composed of cells treated with phorbol myrestate acetate (PMA) and of untreated cells [30].

Data (mean ± SD, MPa) were analyzed by one-way ANOVA and Tukey test ($p < 0.05$). Nonparametric data were expressed as median and were analyzed using the Mann-Whitney test ($p < 0.05$). All analysis was performed with GraphPad PRISM[®] Version 1.5 software (GraphPad Software Inc., USA).

Results

Purification of PLA₂ and of peptide fraction from Cdt venom

Figure 1 – a shows the chromatographic profile resulting from the molecular exclusion of venom from Cdt. The eluted peaks between 40 and 60 mL (peak 3) were re-chromatographed by the same technique (Fig. 1 – b). Then, the peaks eluted between 43 and 53 mL (peak 3.1) were submitted to ion-exchange chromatography. Figure 1 – c

shows the elution of two distinct peaks. Peak 3.1.2 was identified as PLA₂ and confirmed by SDS-PAGE (Fig. 1 – d), phospholipase activity (Fig. 2) and Edman sequencing (data not shown).

Leishmanicidal activity

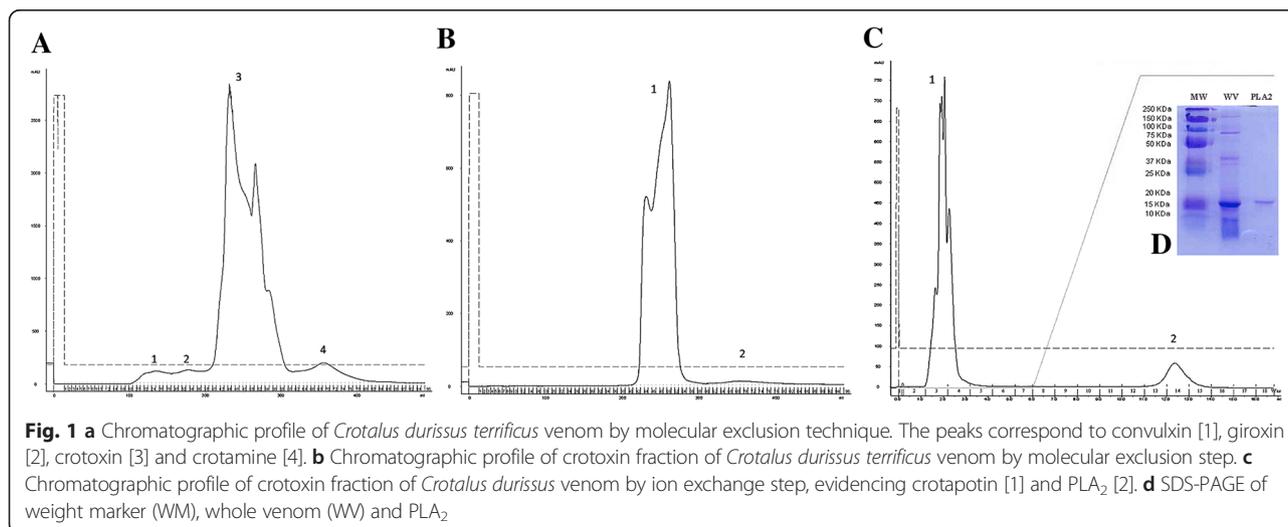
Figure 3 (a and b) displays leishmanicidal activities of PLA₂ toxin and of the Cdt peptide fraction, respectively. The MTT colorimetric method in microplates revealed that the IC₅₀ levels were 52.07 µg/mL for PLA₂ and 16.98 µg/mL for the peptide fraction. Amphotericin B (positive control) showed IC₅₀ 0.089 µg/mL.

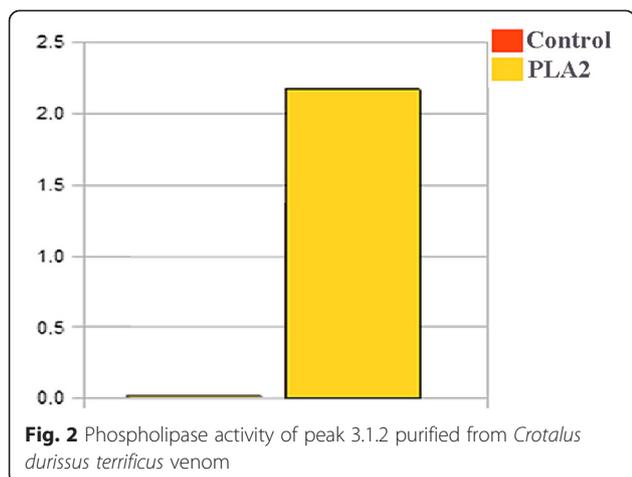
Cytotoxic activity in macrophages

Figure 4 (a and b) shows the cytotoxicity of PLA₂ and the peptide fraction of Cdt by utilizing peritoneal macrophages of BALB/c mice, as evaluated via MTT colorimetry. The IC₅₀ levels were, respectively, 52.07 µg/mL for PLA₂, and 16.98 µg/mL for the peptide fraction.

Determination of production of Nitric Oxide (NO) and Hydrogen Peroxide (H₂O₂)

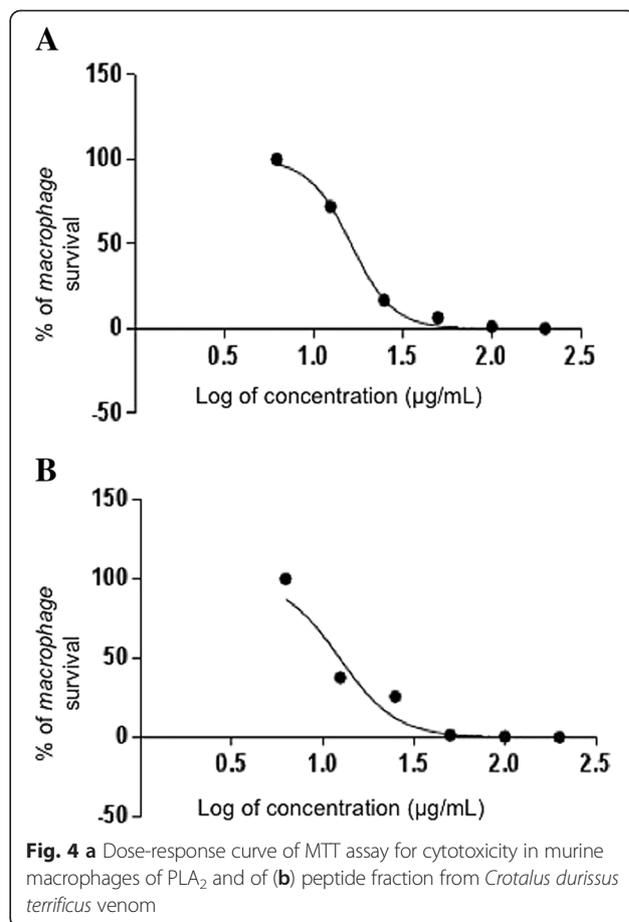
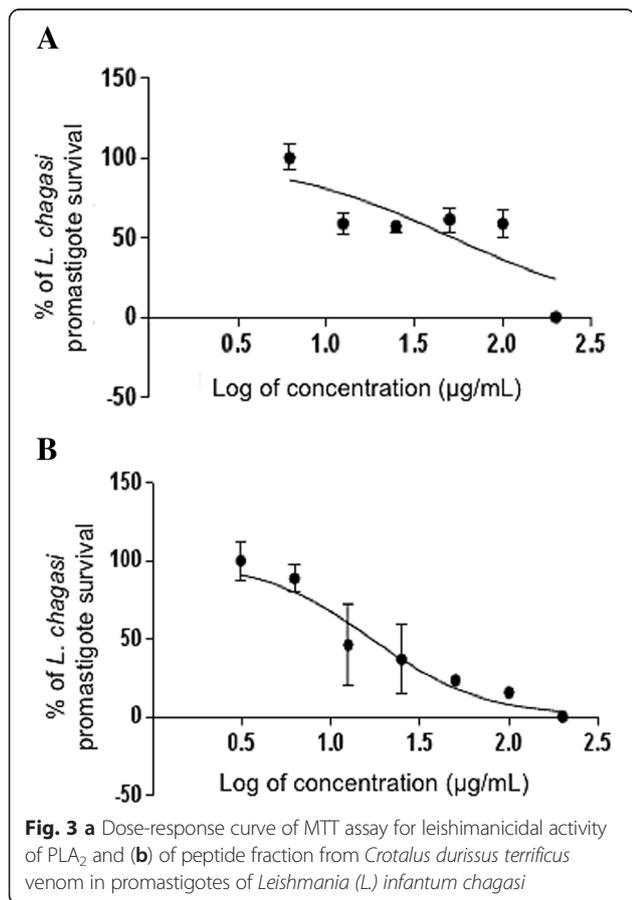
Figure 5 (a, b and c) displays the NO production of supernatant from murine macrophages infected by *L. (L.) chagasi* treated with PLA₂, measured at three different incubation moments (24, 48, and 72 h). At 24 h (Fig. 4 – a), NO production by the lipopolysaccharide-treated control (LPS) was greater than that of the control constituted by untreated macrophages (MO). In relation to the production of NO between the treatments with PLA₂ and the controls, the treatments T1, T3, T5, T6 and T8 did not differ from the MO control. As to the LPS control, these treatments produced less NO. The treatment T4 differed from the other treatments T1, T3, T5, T6 and T8 and the MO control, but did not present a





difference in relation to the LPS control. The treatment T7 differed significantly only from the MO control.

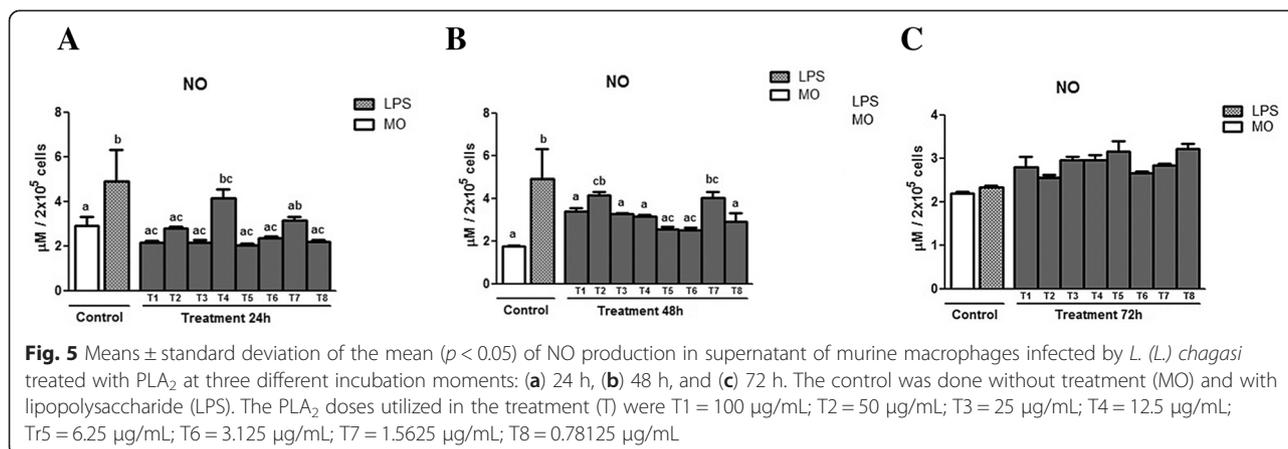
After 48 h of incubation (Fig. 5 – b), treatment T2 presented higher production than the untreated group, but did not differ from the LPS control. Treatment T7 presented greater production than the untreated control, but showed no difference in relation to the LPS-treated control. The treatments T5 and T6 produced less NO than the LPS-treated group. At 72 h (Fig. 5 – c), a



tendency toward increasing NO production was found only in the PLA₂-treated groups although there was no significant difference in relation to the controls. The peptide fraction was unable to stimulate the production of NO by macrophages.

Figure 6 (a, b and c) shows the production of H₂O₂ in the supernatant of macrophages treated with PLA₂ at 24, 48 and 72 h. At 24 h of incubation (Fig. 5 – a), a difference in relation to the two controls was observed only in treatment T1. At 48 and 72 h (Fig. 6 – b and c), an increase of H₂O₂ production was found, most prominently after 72 h, when all the treated groups differed ($p < 0.05$) from the controls. There was no difference among the treatments.

Figure 7 (a, b and c) displays the H₂O₂ production in the supernatant of macrophages treated with Cdt peptide fraction at 24, 48, and 72 h. At 24 h of incubation (Fig. 7 – a), the controls did not differ from the treatments ($p < 0.05$). At 48 h of incubation (Fig. 7 – b), a significantly greater production of H₂O₂ was observed in treatments T3, T6 and T7 ($p < 0.05$) in relation to the controls MO and PMA. After 72 h of incubation, in all the treatments, the H₂O₂ production was higher ($p < 0.05$) than those of PMA and the MO controls.



Discussion

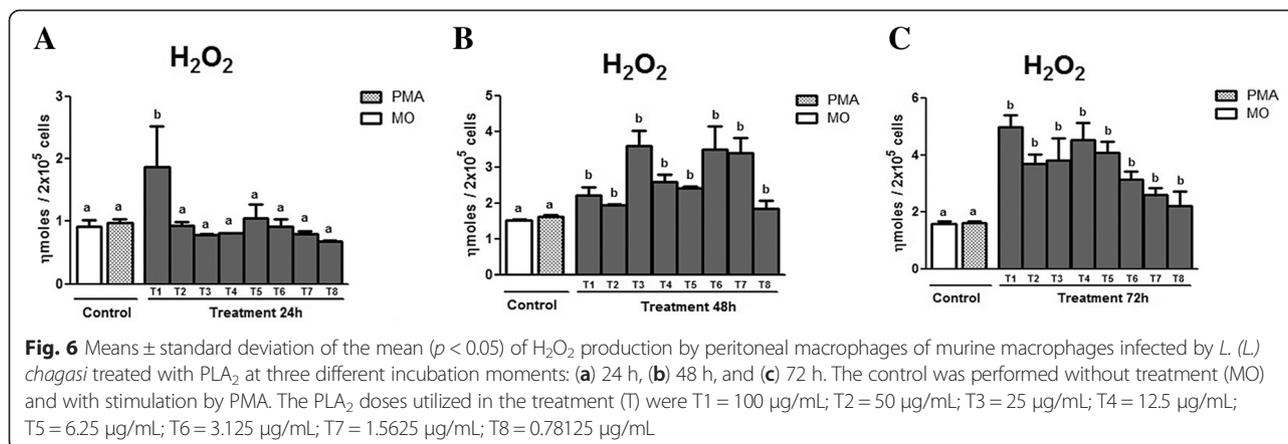
An antiparasitic effect of snake venom has been amply described with the aim of discovering target molecules. The ultrastructural alterations caused by these venoms in parasites involve mitochondrial edema and kinetoplast dysregulation or intense cytoplasmic vacuolization and enlargement of the flagellar pocket [17, 31]. The venoms can affect the respiratory chain through ATP leakage in promastigotes and, therefore, these organelles would be the first to suffer from the treatment [11, 31].

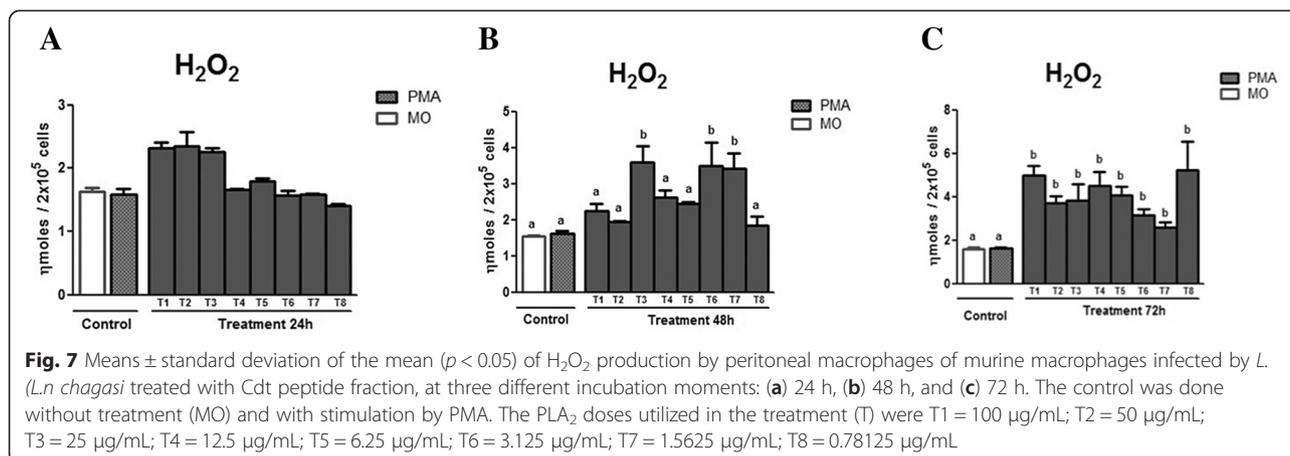
In our present work, we have noted that the PLA₂ isolated and purified from the venom of *Crotalus durissus terrificus* (Cdt) was able to inhibit the proliferation of *Leishmania chagasi* promastigotes at a rate from 50 % to 83 % at doses from 50 to 200 $\mu\text{g/mL}$, respectively. It was also observed that this inhibition rate was dose-dependent.

The action of PLA₂ from venom of snakes of the genus *Bothrops* in the inhibition of *Leishmania* promastigote growth was described against *Leishmania major*, *L. (L.) amazonensis*, *L. (V.) braziliensis*, and *L. chagasi* [15–17]. The results found for PLA₂ from Cdt corroborate these observations, despite small differences in the composition of PLA₂ and in the susceptibility of the

parasites. The PLA₂ of Cdt is an enzymatically active Asp-49 whose action is associated with its catalytic site while it interacts hydrophobically with the plasma membrane penetrating into the lipid bilayer, causing an electrostatic perturbation and thereby rupturing the membrane [18, 27, 32, 33]. This direct action may have caused the inhibition of promastigote growth observed in our study.

In vivo studies have shown that promastigotes of *Leishmania amazonensis* treated with PLA₂ purified from *Crotalus durissus colilineatus* venom augment the size of lesions in BALB/c mice. A prior study indicated that PLA₂ appears to be a factor in the progression of cutaneous leishmaniasis, since macrophages treated with PLA₂ presented elevated levels of prostaglandin E₂ (PGE₂) (an inflammatory lipid mediator) and inhibited levels of IL-2, a cytokine associated with Th-1 response [34]. The results of the present study corroborate these authors since weaker dilutions of the PLA₂ utilized to treat the peritoneal macrophages infected by *L. (L.) infantum chagasi* amastigotes induced the proliferation of parasites inside the macrophages, while at doses higher than 50 $\mu\text{g/mL}$ they were able to diminish the infection by up to 27.3 %.





The low-molecular weight compounds found in snake venoms have aroused the interest of the pharmaceutical industry due to their great variety of pharmacological effects. The peptides that have already been isolated from snake venoms include: type-C natriuretic peptides, bradykinin-potentiating peptides (BPPs), sarafotoxins, waglerins and peptides that present inhibitory activity against metalloproteinases, and antimicrobial peptides [25, 35].

The antimicrobial peptides, which represent an extremely diverse group, participate primarily in the innate immune system by acting as an initial barrier of immune defense in many organisms, including plants, insects, bacteria and vertebrates [26, 36, 37]. The peptide Pep5Bj, from the venom of *Bothrops jararaca*, presents a high level of inhibitory activity against the growth of the fungi *Fusarium oxysporum* and *Colletotrichum lindemuthianum* and the yeasts *Candida albicans* and *Saccharomyces cerevisiae* [38].

The peptide fraction of Cdt venom presented an IC_{50} of 16.98 $\mu\text{g/mL}$ for antileishmanial activity against the promastigotes, equivalent to the cytotoxic activity IC_{50} of 16.98 $\mu\text{g/mL}$, and is able to inhibit the growth of not only promastigotes, but also 43 % of amastigotes at the same dose.

An important factor in leishmaniasis is the action of macrophages, since they are the main host cells of the parasite, and also play an important role in the control of parasites, by means of producing cytokines and metabolic oxygen compounds such as nitric oxide (NO) [37, 39].

The action mechanism of NO, as well as its specific targets within the metabolic pathways in *Leishmania* sp., were previously described as was their susceptibility to NO production by the host cell [36, 37]. In our study, the quantity of NO produced during the infection was measured at 24, 48, and 72 h, showing different results among these moments. At 24 h, the controls differed from treatment T7 (1.5625 $\mu\text{g/mL}$). After 48 h of

treatment, there was a difference between the control groups, as well as between some of the treated groups. Yet at 72 h, greater NO production was shown, even in the controls.

The production of NO, one of the main microbicidal agents from murine macrophages, can be stimulated by the action of PLA_2 from Cdt venom [40]. However, some species of *Leishmania*, such as *L. Amazonensis* and *L. chagasi*, are able to inhibit NO production [41]. In this manner, we may infer that in the first 24 h of treatment, the parasites had supported the cellular defenses through some mechanism of escape. But after 72 h the defenses produced by the macrophages would become sufficient to suppress the modulation of the parasite in the macrophage defenses. In this sense, the action of PLA_2 impeded the parasite from maintaining its escape mechanism, a fact also observed in a prior study that measured the production of $TNF-\alpha$, NO, and IL-10 in treatment with PLA_2 from *B. pauloensis* in experimental infection of murine macrophages with *L. amazonensis* [17].

It is important to emphasize that in relation to NO production by a pool of peptides, there was no difference between the treatments and the controls, in any of the readings.

H_2O_2 is an antiparasitic compound that is produced by defense cells such as macrophages, when challenged by such intracellular parasites as *Leishmania* sp. A difference in H_2O_2 production was observed in our study only at 48 h of treatment.

Previous studies demonstrate the resistance of *Leishmania* spp. to H_2O_2 , where it could be noted that the parasites produce catalase and superoxide dismutase, enzymes that confer an efficient escape mechanism from the action of H_2O_2 [15, 17, 35]. This resistance of the parasite to H_2O_2 production may account for the low production of this metabolite in the first 24 h, after which the activity of the peptide fraction perturbed the metabolism of the parasites, by which the parasite

ceases or diminishes the production of enzymes that degrade H₂O₂.

Conclusion

Due to the high toxicity presented by drugs utilized for the treatment of leishmaniasis and their important side effects, it has become necessary to seek new molecules with therapeutic potential against these diseases. In this context, the present work has demonstrated in vitro leishmanicidal activity of the PLA₂ and peptide fraction of *Crotalus dutissus terrificus* venom. New studies must be performed to describe the metabolic pathways involved in the process of cellular death. Furthermore, the prospecting of molecules whose peptide fraction presents antiparasitic activity, as well as the immunomodulation of these new molecules, must be investigated.

Ethics committee approval

All experiments followed the ethical principles for animal experimentation adopted by the Brazilian College of Animal Experimentation (COBEA) and were approved by the Ethics Committee for Animal Experimentation at the Botucatu Medical School, UNESP (CEEA 890-2011).

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

GACB and AVP carried out the experimental protocols. LCB and AU performed the purification procedures. SAC and LDS analyzed data. BB and RSFJ designed experiments and wrote and revised the manuscript. RSFJ supervised the research work. All authors read and approved the final manuscript.

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