

Checklist of helminths from lizards and amphisbaenians (Reptilia, Squamata) of South America

Ávila RW (1), Silva RJ (1)

(1) Department of Parasitology, Botucatu Biosciences Institute, São Paulo State University (UNESP – Univ Estadual Paulista), Botucatu, São Paulo State, Brazil.

Abstract: A comprehensive and up to date summary of the literature on the helminth parasites of lizards and amphisbaenians from South America is herein presented. One-hundred eighteen lizard species from twelve countries were reported in the literature harboring a total of 155 helminth species, being none acanthocephalans, 15 cestodes, 20 trematodes and 111 nematodes. Of these, one record was from Chile and French Guiana, three from Colombia, three from Uruguay, eight from Bolivia, nine from Surinam, 13 from Paraguay, 12 from Venezuela, 27 from Ecuador, 17 from Argentina, 39 from Peru and 103 from Brazil. The present list provides host, geographical distribution (with the respective biome, when possible), site of infection and references from the parasites. A systematic parasite-host list is also provided.

Key words: Cestoda, Nematoda, Trematoda, Squamata, neotropical.

INTRODUCTION

Parasitological studies on helminths that infect squamates (particularly lizards) in South America had recent increased in the past few years, with many new records of hosts and/or localities and description of several new species (1-3). Such studies, however, date back to 1920, when Dr. Lauro Travassos and his colleagues presented important contributions to the field of helminth systematics (4-6).

The first comprehensive efforts to summarize the knowledge about helminths of squamates are those from Baker (7), who listed all available published data on the occurrence of nematodes infecting reptiles and amphibians. Other studies have a regional scope and treat each helminth group separately, for example, nematodes collected from reptiles of Brazil, digenetic trematodes of amphibians and reptiles from Argentina and acanthocephalans in amphibians and reptiles from Brazil and Paraguay (8-10).

The present checklist summarizes the diversity of helminths from lizards and amphisbaenians of South America, providing a host-parasite list with localities and biomes.

STUDIED REGIONS

This checklist was prepared based on records of nematodes, acanthocephalans, cestodes, and trematodes published from 1914 to 2010, excluding material from the so called “grey literature”, that is, records of unpublished theses or scientific meetings. The taxonomy of helminths follows: nematodes, cestodes, trematodes and acanthocephalans (6, 8, 11-15). Whenever possible, taxonomy of hosts is reviewed with recent publications.

Each record is constituted by the class, order, superfamily, family and species of the helminth; as well as the name of the authority who described it and year; besides its host(s), geographical record with country and state/province (when available) and information on biome of host. Comments

include taxonomy, synonyms and also changes in the host taxonomy.

Abbreviations for all political units cited in this paper are:

- Brazil: Acre (AC), Alagoas (AL), Amapá (AP), Amazonas (AM), Bahia (BA), Ceará (CE), Distrito Federal (DF), Goiás (GO), Espírito Santo (ES), Maranhão (MA), Mato Grosso (MT), Mato Grosso do Sul (MS), Minas Gerais (MG), Pará (PA), Paraíba (PB), Paraná (PR), Pernambuco (PE), Piauí (PI), Rio de Janeiro (RJ), Rio Grande do Norte (RN), Rio Grande do Sul (RS), Rondônia (RO), Roraima (RR), São Paulo (SP), Santa Catarina (SC), Sergipe (SE), and Tocantins (TO).
- Argentina: Buenos Aires (BA), Catamarca (CA), Córdoba (CR), Corrientes (CO), Chaco (CH), Chubut (CB), Entre Ríos (ER), Formosa (FO), Jujuy (JU), La Pampa (LP), La Rioja (LR), Mendoza (ME), Misiones (MI), Neuquén (NE), Rio Negro (RN), Salta (SA), San Juan (SJ), San Luis (SL), Santa Cruz (SC), Santa Fe (SF), Santiago del Estero (SE), Tierra del Fuego (TF), and Tucumán (TU).
- Paraguay: Alto Paraguay (AP), Alto Paraná (AA), Amambay (AM), Distrito Capital (DC), Boquerón (BO), Caaguazú (CA), Caazapá (CZ), Canindeyú (CN), Central (CE), Concepción (CO), Cordillera (CR), Guairá (GU), Itapúa (IT), Misiones (MI), Ñeembucú (NE), Paraguairí (PA), Presidente Hayes (PH), and San Pedro (SP).
- Bolivia: Chuquisaca (CH), Cochabamba (CO), Beni (BE), La Paz (LP), Oruro (OR), Pando (PA), Potosí (PO), Santa Cruz (SC), and Tarija (TA).
- Venezuela: Amazonas (AM), Anzoátegui (AN), Apure (AP), Aragua (AR), Barinas (BA), Bolívar (BO), Carabobo (CA), Cojedes (CO), Delta Amacuro (DA), Falcón (FA), Guárico (GU), Lara (LA), Mérida (ME), Miranda (MI), Monagas (MO), Nueva Esparta (NE), Portuguesa (PO), Sucre (SU), Táchira (TA), Trujillo (TR), Yaracuy (YA), Vargas (VA), and Zulia (ZU).
- Ecuador: Azuay (AZ), Bolívar (BO), Cañar (CA), Carchi (CR), Cotopaxi (CO), Chimborazo (CH), Galápagos (GA), Imbabura (IM), Loja (LO), Pichincha (PI), Tungurahua (TU), El Oro (EO), Esmeraldas (ES), Guayas (GU), Los Ríos (LR), Manabí (MA), Morona-Santiago (MS), Napo (NA), Pastaza (PA), Zamora-Chinchipe (ZC), Sucumbíos (SU), and Orellana (OR).
- Peru: Amazonas (AM), Ancash (AN), Apurímac (AP), Arequipa (AR), Ayacucho (AY), Cajamarca (CA), Callao (CL), Cusco (CU), Huancavelica (HU), Huánuco (HA), Ica (IC), Junín (JU), La Libertad (LL), Lambayeque (LA), Lima (LI), Loreto (LO), Madre de Dios (MD), Moquegua (MO), Pasco (PA), Piura (PI), Puno (PU), San Martín (SM), Tacna (TA), Tumbes (TU), and Ucayali (UC).
- Colombia: Amazonas (AM), Antioquia (AN), Arauca (AR), Atlántico (AT), Bolívar (BO), Boyacá (BY), Caldas (CA), Caquetá (CQ), Casanare (CS), Cauca (CU), Cesar (CE), Chocó (CH), Córdoba (CO), Cundinamarca (CN), Guainía (GU), Guaviare (GA), Huila (HU), La Guajira (LG), Magdalena (MA), Meta (ME), Nariño (NA), Norte de Santander (NS), Putumayo (PU), Quindío (QU), Risaralda (RI), San Andrés (SA), Santander (SN), Sucre (SU), Tolima (TO), Valle del Cauca (VC), Vaupés (VA), and Vichada (VI).
- Chile: Aisén (AI), Antofagasta (AN), Araucanía (AU), Arica (AR), Atacama (AT), Biobío (BI), Coquimbo (CO), Los Ríos (LR), Los Lagos (LL), Magallanes (MG), Maule (MA), O'Higgins (OH), Santiago (AS), Tarapacá (TA) and Valparaíso (VA).
- Uruguay: Artigas (AR), Canelones (CA), Cerro Largo (CL), Colonia (CO), Durazno (DU), Flores (FL), Florida (FO), Lavalleja (LA), Maldonado (MA), Montevideo (MO), Paysandú (PA), Río Negro (RN), Rivera (RI), Rocha (RO), Salto (SA), San José (SJ), Soriano (SO), Tacuarembó (TA), and Treinta y Tres (TT).
- Surinam: Brokopondo (BR), Commewijne (CO), Coronie (CR), Marowijne (MA), Nickerie (NI), Para (PA), Paramaribo (PR), Saramacca (SA), Sipaliwini (SI), and Wanica (WA).
- French Guiana: Awala-Yalimapo (AY), Apatou (AP), Camopi (CA), Cayenne (CY), Grand-Santi (GS), Iracoubo (IR), Kourou (KO), Macouria (MC), Mana (MA), Maripasoula (MR), Matoury (MT), Montsinéry-Tonnegrande (MO), Ouanary (OU), Papaïchton (PA), Régina (RE), Remire-Montjoly (RM), Roura (RO), Saint-Élie (SE), Saint-Georges (SG), Saint-Laurent-du-Maroni (SL), Saül (SA), and Sinnamary (SI).

- Guyana: Barima-Waini (BW), Pomeroon-Supenaam (PS), Essequibo Islands-West Demerara (EI), Demerara-Mahaica (DM), Mahaica-Berbice (MB), East Berbice-Corentyne (EC), Cuyuni-Mazaruni (CM), Potaro-Siparuni (PS), Upper Takutu-Upper Essequibo (TE), and Upper Demerara-Berbice (DB).

LIST OF HELMINTHS

Acanthocephala

Hosts: *Tropidurus torquatus* Wied-Neuwied, 1820

Site of infection: Stomach wall

Distribution: Argentina – CO (Chaco).

Comments: No indication of family and/or genus, but probably belong to Centrorhynchidae, since several cystacanths of this family have been found in South American lizards (16).

Archiacanthocephala

Oligacanthorhynchida

Oligacanthorhynchidae

Oligacanthorhynchus Travassos, 1915

Oligacanthorhynchus sp.

Hosts: *Ameiva ameiva* Linnaeus, 1758

Distribution: Paraguay – Rio Negro (Chaco), PH (10).

Palaeacanthocephala

Echinorhynchida

Echinorhynchidae

Hosts: *Mabuya macrorhyncha* Hoge, 1946

Site of infection: Stomach wall

Distribution: Brazil – Trancoso (Restinga), BA.

Comments: No indication of genus (17).

Acanthocephalus Koelreuther, 1771

Acanthocephalus saurius Bursey & Goldberg, 2003

Hosts: *Cercosaura oshaugnessyi* Boulenger, 1885

Site of infection: Small intestine

Distribution: Brazil – AC (Amazon) (18).

Acanthocephalus sp.

Hosts: *Enyalius perditus* Jackson, 1978

Site of infection: Stomach

Distribution: Brazil – São Sebastião (Atlantic Forest), SP (3).

Echinorhynchus Zoega in Müller, 1776

Echinorhynchus sp.

Hosts: *Tropidurus torquatus*

Site of infection: Small intestine

Distribution: Brazil (19).

Polymorphida

Centrorhynchidae

Hosts: *Mabuya macrorhyncha*; *M. agilis* Raddi, 1823, *Hemidactylus mabouia* Moreau de Jonnés, 1818, *Enyalius bilineatus* Duméril & Bibron, 1837

Site of infection: Peritonium, liver; mesentery

Distribution: Brazil – Queimada Grande island (Atlantic forest) and Valinhos (Cerrado), SP; Grussaí (Restinga) and Nova Iguaçu (Atlantic Forest), RJ; Praia das Neves (Restinga) and Marechal Floriano (Atlantic Forest), ES.

Comments: All citations above consist of cystacanth occurrence and are, thus, difficult to identify to species level, because reproductive structures are required (20-24).

Centrorhynchus Lühe, 1911

Centrorhynchus tumidulus Rudolphi, 1919

Hosts: *Ameiva ameiva*; *Tropidurus torquatus*; *Tupinambis teguixin* Linnaeus, 1758

Site of infection: Body cavity

Comments: *Tupinambis teguixin* found in Rio de Janeiro and in open habitats in South America may actually correspond to *T. merianae*.

Distribution: Brazil – Rio de Janeiro (Atlantic Forest), RJ (25, 26).

Centrorhynchus sp.

Hosts: *Tupinambis teguixin*

Site of infection: Not informed

Distribution: Paraguay – AP (Chaco) (10).

Nematoda

Adenophorea

Enoplida

Capilariidae

Capillaria (Zeder, 1800) Bell & Beverley Burton 1981

Capillaria (= *Amphibiocapillaria*) *freitaslenti* Araujo & Gandra, 1941

Hosts: *Tropidurus torquatus*; *Ameiva ameiva*

Site of infection: Intestine

Distribution: Brazil – GO (Cerrado); Peru – CU (Amazon) (1, 8).

Ichthyocapillaria Moravec, 1982

Ichthyocapillaria (= *Pseudocapillaria*) *maricaensis*

Rodrigues, 1992

Hosts: *Liolaemus lutzae* Mertens, 1938

Site of infection: Small intestine

Distribution: Brazil – Maricá (Restinga), RJ (27).

Ascaridida

Anisakidae

***Contracaecum* Railliet & Henry, 1912**

Contracaecum sp.

Hosts: *Tupinambis teguixin*

Site of infection: Not informed

Distribution: Surinam – PR (Amazon) (28).

Ascarididae

***Dujardinascaris* Baylis, 1927**

Dujardinascaris sp.

Hosts: *Kentropyx pelviceps* Cope, 1868,

Tupinambis teguixin

Site of infection: Stomach

Distribution: Peru – CU (Amazon) (1).

***Freitasascaris* Sprent, 1983**

Freitasascaris alata Baylis, 1947

Hosts: *Tupinambis teguixin*

Site of infection: Intestine

Distribution: Surinam – PR (Amazon); Brazil – AM (Amazon); Uruguay – MO

Comments: *Tupinambis teguixin* from Uruguay may actually correspond to *T. merianae*. (28-30).

***Hexametra* Travassos, 1919**

Hexametra boddaertii Baird, 1860

Hosts: *Cnemidophorus littoralis* Rocha, Araujo, Vrcibradic & Costa, 2000; *C. ocellifer* Spix, 1825; *Mabuya agilis*; *M. frenata* Cope, 1862; *M. macrorhyncha*; *Tropidurus torquatus*

Site of infection: Body cavity

Distribution: Brazil – Jurubatiba, Grussaí and Marica (Restinga), RJ; Salvador and Trancoso (Restinga), BA; Valinhos (Cerrado), SP (17, 23, 31-35).

Atractidae

***Atractis* Dujardin, 1945**

Atractis cruciata Linstow, 1902

Hosts: *Tupinambis teguixin*, *Stenocercus roseiventris*

Site of infection: Intestine

Distribution: Brazil – Belém (Amazon), PA; Ecuador – PA (Amazon).

Comments: *Atractis cruciata* is considered a junior synonym of *A. opeatura* (7, 36). However,

it is considered a valid species by some authors (8, 37-39).

***Cyrtosomum* Gedoelst, 1919**

Cyrtosomum longicaudatum Brenes and Bravo Hollis, 1960

Hosts: *Anolis punctatus* Daudin, 1802; *Anolis transversalis* Duméril, 1851

Site of infection: Large intestine

Distribution: Peru – CU (Amazon); Brazil – AM (Amazon) (1, 40).

Cyrtosomum sp.

Hosts: *Ophiodes striatus*

Site of infection: Large intestine

Distribution: Brazil – MS (Cerrado) (41).

***Maracaya* Díaz-Ungría, 1964**

Maracaya belemensis Adamson & Baccam, 1988

Hosts: *Amphisbaena alba* Linnaeus, 1758

Site of infection: Large intestine

Distribution: Brazil – Belém (Amazon), PA (42).

Maracaya graciai Díaz-Ungría, 1969

Hosts: *Amphisbaena alba*

Site of infection: Intestine

Distribution: Venezuela – Maracay, AR (7, 42).

Maracaya pusilla (Miranda, 1924) Adamson & Baccam, 1988

Hosts: *Amphisbaena* sp.

Site of infection: Intestine

Distribution: Brazil – BA.

Remarks: The former *Aplectana pusilla* was transferred to *Maracaya* (42), according to a revision of the genus *Maracaya* (43), but it is cited in the genus *Aplectana* by other authors (42-44).

Cosmocercidae

gen. sp.

Hosts: *Cercosaura argulus* Peters, 1863

Site of infection: Small intestine

Distribution: Peru – Rio Nanay (Amazon), LO (45).

***Aplectana* Railliet & Henry, 1916**

Aplectana albae Adamson & Baccam, 1988

Hosts: *Amphisbaena alba*; *Amphisbaena ridleyi* Boulenger, 1890

Site of infection: Intestine

Distribution: Brazil – Belém (Amazon), PA; Fernando de Noronha Island, PE (42, 46).

Aplectana railletii Travassos, 1925

Hosts: *Amphisbaena alba*
 Site of infection: Intestine
 Distribution: Brazil – Angra dos Reis (Insular Atlantic Forest), RJ.
 Comments: The host *Amphisbaena fusca* was originally cited as *Amphisbaena alba* (5, 7, 8, 47).

Aplectana tucumanensis Ramallo, Bursey & Goldberg, 2008

Hosts: *Amphisbaena bolivica* Mertens, 1929
 Site of infection: Large intestine
 Distribution: Argentina – San Miguel de Tucumán (Chaco), TU (44).

Aplectana unguiculata Rudolphi, 1819

Hosts: *Amphisbaena* sp.
 Site of infection: Not informed
 Distribution: Brazil.
 Comments: The species is actually considered as *species inquirenda* (7).

Aplectana vellardi Travassos, 1926

Hosts: *Enyalius perditus*
 Site of infection: Intestine
 Distribution: Brazil – Ibitipoca (Cerrado), MG (48).

***Cosmocerca* Diesing, 1861**

Cosmocerca rara Freitas & Vicente, 1966
 Hosts: *Leposternon microcephalum* Wagler, 1824
 Site of infection: Large intestine
 Distribution: Brazil – Rio de Janeiro (Atlantic forest), RJ (8, 49).

Cosmocerca vrcibradici Bursey & Goldberg, 2004

Hosts: *Alopoglossus angulatus* Linnaeus, 1758; *Alopoglossus atriventris* Duellman, 1973; *Anolis fuscoauratus* D'Orbigny, 1837; *Cercosaura eigenmanni* Griffin, 1917; *C. oshaughnessyi* Boulenger, 1885; *Uranoscodon superciliosus* Linnaeus, 1758
 Site of infection: Intestine
 Distribution: Brazil – AC, AM, PA, RO (Amazon); Ecuador – SU (Amazon) (18, 50-52).

Cosmocerca sp.

Hosts: *Enyalius perditus*
 Site of infection: Stomach
 Distribution: Brazil – São Sebastião (Insular Atlantic forest), SP (3).

***Paradollfusnema* Baker, 1982**

Paradollfusnema amphisbaenia Baker, 1981
 Hosts: *Leposternon microcephalum*
 Site of infection: Intestine
 Distribution: Brazil – Recife, PE (8, 53).

***Railletnema* Travassos, 1927**

Railletnema brachyspiculatum Bursey, Goldberg, Salgado-Maldonado, and Mendez-de-la-Cruz, 1998
 Hosts: *Morunasaurus annularis* (O'Shaughnessy, 1881)
 Site of infection: Large intestine
 Distribution: Ecuador – PA (Amazon) (39).

Heterakidae

***Africana* Travassos, 1920**

Africana chabaudi Baker, 1981
 Hosts: *Uranoscodon superciliosus*
 Site of infection: Intestine
 Distribution: Brazil – PA, RO and RR (Amazon) (50, 54).

Africana dardanelosi Ávila & Silva, 2009

Hosts: *Hoplocercus spinosus*
 Site of infection: Intestine
 Distribution: Brazil – MT (Amazon) (55).

***Moaciria* Freitas, 1956**

Moaciria alvarengai Freitas, 1956
 Hosts: *Trachylepis atlantica* Schmidt, 1945
 Site of infection: Large intestine
 Distribution: Brazil – Fernando de Noronha Island, PE (8, 46, 56).

***Spinicauda* Travassos, 1920**

Spinicauda spinicauda Olfers, 1819
 Hosts: *Ameiva ameiva*, *Tupinambis teguixin*, *Tupinambis merianae*, *Trachylepis atlantica*, *Enyalioides praestabilis* (O'Shaughnessy, 1881)
 Site of infection: Intestine
 Distribution: Brazil – MT (Pantanal), Rio de Janeiro, RJ, São Gonçalo, RN and Fortaleza, CE, Fernando de Noronha Islands, PE; Peru – CU (Amazon); Surinam – PR (Amazon); Venezuela – AR; Ecuador – PA (Amazon) (4, 7, 28, 39, 46, 50, 57-60).

***Strongyluris* Mueller, 1894**

Strongyluris oscari Travassos, 1923
 Hosts: *Ameiva ameiva*; *Anolis fuscoauratus*; *A.*

punctatus; *A. transversalis*; *Enyalius iheringii* Boulenger, 1885; *E. perditus*; *Eurolophosaurus nanuzae* Rodrigues, 1981; *Mabuya agilis*; *Plica plica* Linnaeus, 1758; *Plica umbra* Linnaeus, 1758; *Stenocercus caducus*; *S. roseiventris* D'Orbigny in Duméril & Bibron 1837; *Tropidurus* sp.; *T. guarani* Alvarez, Cei & Scolaro, 1994; *T. spinulosus* Cope, 1862; *T. torquatus*; *T. melanopleurus*
 Site of infection: Stomach, intestine
 Distribution: Brazil – AC, AM, BA, CE, DF, ES, GO, MG, MS, PA, PB, RJ, RO, SP (Pantanal, Restinga, Amazon, Caatinga, Atlantic Forest); Argentina (Chaco); Paraguay (Chaco); Ecuador – SU and PA (Amazon); Peru – CU (Amazon); Bolivia – Florida province, SC.
 Comments: Currently, 31 species are recognized within the genus, and based on morphological characteristics, only four occurs in neotropical regions (61). *Strongyluris oscar* seems to be extremely polymorphic, since the species *S. freitasi*, *S. travassosi* and *S. sai* have been synonymized (62). The records of *S. oscar* infecting *T. spinulosus* in the region of Salobra, MS, Brazil correspond to *T. guarani* (1, 3, 32, 35, 39-41, 48, 51, 60, 62-69).

Kathlaniidae

***Cruzia* Travassos, 1917**

Cruzia fulleborni Khalil & Vogelsang, 1930
 Hosts: *Tupinambis teguixin*
 Site of infection: Intestine
 Distribution: Argentina; Paraguay (7, 70-72).

Cruzia rudolphii Ruiz, 1947

Hosts: *Hoplocercus spinosus* Fitzinger, 1843
 Site of infection: Large intestine
 Distribution: Brazil – Dois Irmãos do Buriti (Cerrado), MS (71, 73).

Cruzia tentaculata Rudolphi, 1819

Hosts: *Tupinambis teguixin*
 Site of infection: Intestine
 Distribution: Uruguay – MO (29, 71).

Cruzia travassosi Kalil & Vogelsangi, 1932

Hosts: *Tupinambis merianae*
 Site of infection: Intestine
 Distribution: Brazil – Salobra (Pantanal) and Bodoquena (Cerrado) MS; Bolivia; Argentina (7, 41, 71).

***Falcaustra* Lane, 1915**

Falcaustra belemensis Baker & Bain, 1981
 Hosts: *Neusticurus bicarinatus* Linnaeus, 1758; *Potamites ecleopus* Cope, 1876; *P. strangulatus* Cope, 1868
 Site of infection: Rectum
 Distribution: Ecuador – PA (Amazon) (39).

Falcaustra sp.

Hosts: *Anolis punctatus*
 Site of infection: Large intestine
 Distribution: Brazil – AM (Amazon) (40).

Seuratidae

***Skrjabinelazia* Sypliaxov, 1930**

Skrjabinelazia galliardi Chabaud, 1973
 Hosts: *Gonatodes humeralis* Guichenot, 1855
 Site of infection: Stomach
 Distribution: Brazil – Belém (Amazon), PA (8, 74, 75).

Skrjabinellazia intermedia Freitas, 1940

Hosts: *Anolis punctatus*; *Cnemidophorus natio* Rocha, Bergallo & Peccinini-Seale, 1997; *Tropidurus guarani*; *T. torquatus*; *Stenocercus caducus* (Cope, 1862)
 Site of infection: Stomach, intestine
 Distribution: Brazil – Salobra (Pantanal) and Bodoquena (Cerrado), MS; Salvador and Guaratiba (Restinga), BA; Jurubatiba (Restinga), RJ; Paraguay – (Chaco); Peru – CU (Amazon).
 Comments: Originally described as *Salobrella intermedia* found in *Tropidurus spinulosus* from Salobra, MS. See comments on host taxonomy at *Strongyluris oscar* (1, 35, 41, 62, 65, 76, 77).

Subuluridae

***Subulura* Molin, 1860**

Subulura lacertilia Vicente, Van-Sluys, Fontes & Kiefer, 2000
 Hosts: *Eurolophosaurus nanuzae*, *Cnemidophorus natio*
 Site of infection: Intestine
 Distribution: Brazil – Serra do Cipó (Cerrado), MG; Guaratiba (Restinga), BA (66, 77, 78).

Subulura sp.

Hosts: *Tropidurus torquatus*
 Site of infection: Small intestine
 Distribution: Brazil – Jurubatiba (Restinga), RJ (35).

Oxyurida

Oxyuridae

***Gynaecometra* Araújo, 1976**

Gynaecometra bahiensis Araújo, 1976

Hosts: *Polychrus acutirostris* Spix, 1825

Site of infection: Intestine

Distribution: Brazil – Xique-xique (Caatinga), BA; Campo Grande (Cerrado), MS (41, 79).

***Typhlonema* Kreis, 1938**

Typhlonema sp.

Hosts: *Tropidurus guarani*

Site of infection: Not informed

Distribution: Brazil – Salobra (Pantanal), MS (8).

Pharyngodonidae

***Alaeuris* Thapar, 1925**

Alaeuris caudatus Lent & Freitas, 1948

Hosts: *Iguana iguana*

Site of infection: Intestine

Distribution: Brazil – Exu (Caatinga), PE; Venezuela – La Puerta, GU; Peru – TU (8, 29, 80).

Alaeuris conolophi Cuckler, 1938

Hosts: *Conolophus subcristatus* Gray, 1831

Site of infection: Intestine

Distribution: Ecuador – Galapagos Islands (7).

Alaeuris galapagensis Cuckler, 1938

Hosts: *Conolophus subcristatus* Gray, 1831

Site of infection: Intestine

Distribution: Ecuador – Galapagos Islands (7).

Alaeuris labicula Cuckler, 1938

Hosts: *Conolophus subcristatus*

Site of infection: Intestine

Distribution: Ecuador – Galapagos Islands (7).

Alaeuris longispicula Cuckler, 1938

Hosts: *Conolophus subcristatus*

Site of infection: Intestine

Distribution: Ecuador – Galapagos Islands (7).

Alaeuris vogelsangi Lent & Freitas, 1948

Hosts: *Iguana iguana*

Site of infection: Intestine

Distribution: Brazil – Exu (Caatinga), PE, CE; Venezuela – La Puerta, GU (8, 29, 81).

***Ozolaimus* Dujardin, 1845**

Ozolaimus cirratus Linstow, 1906

Hosts: *Iguana iguana*

Site of infection: Intestine

Distribution: Brazil – Santa Luzia (Caatinga), PB; Peru; Venezuela – La Puerta, GU, Blanquilla, Los Frailes, Los Testigos and Margarita; Colombia – LG; Surinam – PR and Marienburg, CO (8, 29, 82, 83).

Ozolaimus megatyphlon Rudolphi, 1819

Hosts: *Iguana iguana*

Site of infection: Intestine

Distribution: Brazil – Santa Luzia (Caatinga), PB; Peru; Venezuela – La Puerta, GU, Blanquilla, Los Frailes, Los Testigos and Margarita; Colombia – LG; Surinam – PR and Marienburg, CO (8, 29, 82, 83).

***Paralaeuris* Cuckler, 1938**

Paralaeuris dorochila Cuckler, 1938

Hosts: *Conolophus subcristatus*

Site of infection: Intestine

Distribution: Ecuador – Galapagos Islands (7).

***Parapharyngodon* Chatterji, 1933**

Parapharyngodon alvarengai Freitas, 1957

Hosts: *Trachylepis atlantica*, *Ameiva ameiva*, *Amphisbaena ridleyi*

Site of infection: Large intestine

Distribution: Brazil – Fernando de Noronha Island, PE and Itaguaí (Restinga), RJ.

Comments: after the synonym it was cited as *Thelandros alvarengai* (46, 84, 85).

Parapharyngodon arequipensis Calisaya & Córdova, 1997

Hosts: *Microlophus peruvianus* (Lesson, 1826)

Site of infection: Large intestine

Distribution: Peru – Omate, MO.

Comments: This species is apparently ignored by several authors (86-90).

Parapharyngodon largitor Alho & Rodrigues, 1963

Hosts: *Ameiva ameiva*, *Mabuya agilis*, *Hemidactylus mabouia*, *Dicrodon guttulatatum*

Site of infection: Intestine

Distribution: Brazil – Manguinhos and Grumari (Restinga), RJ and Valinhos (Cerrado), SP; PERU – LA (8, 22, 33, 91, 92).

Parapharyngodon moqueguensis Calisaya & Córdova, 1997

Hosts: *Microlophus peruvianus*, *Dicrodon guttulatatum*

Site of infection: Large intestine
 Distribution: Peru – Moquegua, MO, LI, TU, PI, LA, LL, AN.
 Comments: check *P. arequipensis* (89, 90, 92).

Parapharyngodon riojensis Ramallo, Bursley & Goldberg, 2002

Hosts: *Phymaturus punae* Cei, Etheridge & Videla, 1985; *P. palluma* Molina, 1782; *Liolaemus buergeri* Werner, 1907
 Site of infection: Large intestine
 Distribution: Argentina – LR (2, 86).

Parapharyngodon sceleratus Travassos, 1923

Hosts: *Ameiva ameiva*; *Cnemidophorus littoralis*; *Eurolophosaurus nanuzae*; *Hemidactylus mabouia*; *Kentropyx pelviceps*; *Liolaemus lutzae*; *Mabuya agilis*; *M. bistriata*; *M. caissara* Rebouças-Spieker, 1974; *M. frenata*; *M. macrorhyncha*; *Microlophus albermalensis* Baur, 1890; *P. johnwrighti*; *Phyllopezus pollicaris*; *T. guarani*; *T. hispidus*; *T. itambere*; *T. semitaeniatus* Spix, 1825; *T. torquatus*; *T. melanopleurus* and *Tropidurus* sp.
 Site of infection: Intestine

Distribution: Brazil – Abrolhos, Trancoso (Restinga) and Canudos, BA; CE; Linhares and Praia das Neves, ES; Serra do Cipó (Cerrado), MG; Salobra (Pantanal), MS; Xavantina, GO; Cachimbo, PA; Mogeiro, Lagoa Remígio, Umbuseiro and João Pessoa, PB; Garanhuns, PE; Grumari, Rio de Janeiro, Arraial do Cabo, Grussaí, Manguinhos and Maricá, RJ; Cruzeta, Currais Novos and Ceará Mirim (Caatinga), RN; Queimada Grande, Caraguatatuba and Valinhos, SP; Bolivia: Florida province, SC and El Carmen; Paraguay; Peru – CU; Ecuador – Galapagos Islands (1, 7, 8, 17, 20, 22, 27, 32-34, 62, 65, 66, 69, 81, 91, 93-97).

Comments: This species has been widely reported as *P. scleratus* (1, 86). However, according to the original description and the latter redescription, the proper spelling of the specific name is *sceleratus* (59, 93). *Cnemidophorus ocellifer*, cited by Ribas *et al.* (94), actually corresponds to *C. littoralis* (31).

Parapharyngodon senisfaciecaudus Freitas, 1957

Hosts: *Liolaemus signifer* Duméril & Bibron, 1837
 Site of infection: Large intestine
 Distribution: Bolivia – LP.

Comments: It was originally described infecting *Liolaemus lenzi*, which actually corresponds to *L. signifer* (98).

Parapharyngodon verrucosus Freitas & Dobbin, 1959

Hosts: *Diploglossus lessonae*
 Site of infection: Intestine
 Distribution: Brazil – João Alfredo (Caatinga), PE; CE (Caatinga) (8, 81, 99).

Parapharyngodon yurensis Calisaya & Córdova, 1997

Hosts: *Microlophus peruvianus*
 Site of infection: Large intestine
 Distribution: Peru – Yura, AR.
 Comments: check *P. arequipensis* (89, 90).

Parapharyngodon sp

Hosts: *Ameiva ameiva*, *Cnemidophorus nativo*, *Hemidactylus mabouia*, *Tropidurus torquatus*, *T. etheridgei*, *Dicrodon guttulatatum*
 Site of infection: Intestine
 Distribution: Brazil – Salobra, MS; Cabo Frio, Nova Iguaçu and Maricá, RJ, Salvador and Guaratiba, BA; Argentina – CO, SA; Peru – TU, LA, LI (8, 16, 21, 77, 92, 100-102).

***Pharyngodon* Diesing, 1861**

Pharyngodon cesarpinto Pereira, 1935
 Hosts: *Cnemidophorus* sp., *Ameiva ameiva*, *Liolaemus lutzae*, *Tropidurus itambere*

Site of infection: Large intestine
 Distribution: Brazil – Juazeiro and Mogeiro (Caatinga), PB, BA, RN; Maricá (Restinga), RJ; Bodoquena (Cerrado), MS.
 Comments: The type of host is apparently misidentified in the original description (60). However, *C. lemniscatus* does not occur in the Caatinga biome, but *C. ocellifer* and at least three undescribed species occur (41, 60, 103, 104).

Pharyngodon micrurus Freitas & Ibañez, 1963

Hosts: *Dicrodon heterolepis*, *D. guttulatatum*
 Site of infection: Intestine
 Distribution: Peru – Trujillo, LL; TU, PI, LA, AN, LI (7, 92, 105).

Pharyngodon travassosi Pereira, 1935

Hosts: *Ameiva ameiva*
 Site of infection: Large intestine
 Distribution: Brazil – Areia (Caatinga), PB (60).

Pharyngodon sp.

Hosts: *Tropidurus torquatus*, *Tropidurus hispidus* Spix, 1825

Site of infection: Intestine
Distribution: Brazil – Salvador, BA; CE (8, 81).

***Pseudostrongyluris* Guerrero, 1971**

Pseudostrongyluris polychrus Guerrero, 1971
Hosts: *Polychrus marmoratus*
Site of infection: Not informed
Distribution: Venezuela – Federal District (7).

***Skrjabinodon* Inglis, 1968**

Skrjabinodon dixoni Bursey & Goldberg, 2007
Hosts: *Uracentron flaviceps* (Guichenot, 1855)
Site of infection: Large intestine
Distribution: Peru – LO (Amazon); Ecuador – SU (106).

Skrjabinodon heliocostai Vicente, Vrcibradic, Muniz-Pereira & Pinto, 2000

Hosts: *Mabuya frenata*
Site of infection: Large intestine
Distribution: Brazil – Valinhos (Cerrado), SP (107).

Skrjabinodon spinulosus Vicente, Vrcibradic, Rocha & Pinto, 2002

Hosts: *Mabuya dorsivittata*
Site of infection: Intestine
Distribution: Brazil – Itatiaia (Atlantic Forest), RJ; Itirapina (Cerrado), SP (108,109).

***Spauligodon* Skrjabin, Schikhobalova & Lagodovsk., 1960**

Spauligodon lobo Ramallo, Bursey & Goldberg (2002)

Hosts: *Liolaemus capillitas* Hulse, 1979; *L. huacahuasicus* Laurent, 1985; *L. quilmes* Etheridge, 1993; *L. ornatus* Koslowsky, 1898; *L. ramirezae* Lobo, 1999

Site of infection: Large intestine
Distribution: Argentina – Tafídel Valle and Amaicha del Valle, TU; El Cerrillo, Andalgalá, Belén, and Santa María, CA; San Antonio de los Cobres, Cachi, and La Poma, SA; and Tilcara and Humahuaca, JU (110).

Spauligodon maytacapaci Vicente & Ibañez (1968)

Hosts: *Leiocephalus* sp.; *Liolaemus andinus* Koslowsky, 1895; *L. chilensis* Lesson, 1830; *L. elongatus* Koslowsky, 1896; *L. lemniscatus* Gravenhorst, 1838; *L. pictus* Duméril & Bibron, 1837; *L. tenuis* Duméril & Bibron, 1837
Site of infection: Intestine

Distribution: Peru – Pumarongo, CA; Argentina; Chile (2, 111, 112).

Spauligodon oxkutzcabiensis (Chitwood, 1938)

Hosts: *Thecadactylus solimoensis* Bergmann & Russell, 2007; *Phyllodactylus reissii* Peters, 1862; *P. inaequalis* Cope, 1876; *P. johnwrighti*; *P. microphyllus* Cope, 1876; *Tropidurus guarani*; *Bogertia lutzae* (Loveridge, 1941).

Site of infection: Intestine
Distribution: Peru – CU, (Amazon); Paraguay – Arroyo Corrientes (Chaco), PA; Brazil – AL (Atlantic Forest).

Comments: *Thecadactylus* was recently subject to revision, and *T. solimoensis* was described by Bergmann & Russel in localities that encompasses the Cuzco, Peru (1, 65, 97, 113-115).

Spauligodon viracochai Freitas, Vicente & Ibañez, 1968

Hosts: *Phyllodactylus gerrhopygus* Wiegmann, 1834; *P. angustidigitus* Dixon & Huey, 1970
Site of infection: Intestine
Distribution: Peru – Trujillo, LL and IC (116, 117).

***Thelandros* Wedl, 1862**

Thelandros bulbosus Salas & Campos, 1974

Hosts: *Microlophus peruvianus*
Site of infection: Not informed
Distribution: Peru.

Comments: Despite the citation by Pérez *et al.* (117), this oxyurid was incorrectly described in “Libro de Resúmenes del IV Congreso Nacional de Biología”, a meeting abstract. Due to an inaccordance with articles 8.1 and 9.9 of the International Code for Zoological Nomenclature, it was therefore considered a *species inquirenda* (107, 118).

Thelandros capacitypanquii Freitas, Vicente & Ibañez, 1968

Hosts: *Dicrodon holmbergi* Schmidt, 1957; *Dicrodon guttulatum*
Site of infection: Intestine
Distribution: Peru – Trujillo, LL, IC, AN (92, 107, 118).

Thelandros sp.

Hosts: *Tropidurus etheridgei*
Site of infection: Intestine
Distribution: Argentina – Departamento Anta (Chaco), SA (102).

Rhabditida

Rhabdiasidae

***Chabirenia* Lhermitte Vallarino, Bain, Deharo, Bertani, Voza, Attout & Gaucher, 2005**

Chabirenia cayennensis Lhermitte Vallarino, Bain, Deharo, Bertani, Voza, Attout & Gaucher, 2005

Hosts: *Ameiva ameiva*

Site of infection: Mouth

Distribution: French Guiana – CY (119).

***Rhabdias* Stiles & Hassall, 1905**

Rhabdias anolis Bursey, Goldberg & Telford, 2003

Hosts: *Anolis punctatus*

Site of infection: Lungs

Distribution: Peru – CU (Amazon) (1).

Rhabdias elegans Gutierrez, 1945

Hosts: *Anolis punctatus*

Site of infection: Lungs

Distribution: Ecuador – PA (Amazon) (1).

Rhabdias sp.

Hosts: *Anisolepis undulatus* Wiegmann, 1834; *Anolis fuscoauratus*; *A. punctatus*; *Enyalius iheringii*; *E. bilineatus*; *E. perditus*; *Tropidurus hispidus*

Site of infection: Lungs, stomach

Distribution: Brazil – Marechal Floriano (Atlantic Forest), ES; PA, AM (Amazon); São Sebastião (Atlantic Forest), SP, Chapada do Araripe (Caatinga), CE; Ecuador – SU.

Comments: Of the neotropical lizards, only *Rhabdias anolis* has been described, despite several reports in the same region (3, 24, 40, 51, 120, 121).

Strongyloididae

***Strongyloides* Grassi, 1897**

Strongyloides cruzi Rodrigues, 1968

Hosts: *Hemidactylus mabouia*

Site of infection: Small intestine

Distribution: Brazil – Manguinhos, RJ (8, 122).

Spirurida

Acuariidae

Hosts: *Cercosaura argulus*; *Hemidactylus mabouia*; *Mabuya agilis*; *M. macrorhyncha*; *Tropidurus torquatus*; *Enyalius bilineatus*; *Phyllodactylus lepidopygus* Tschudi, 1845

Site of infection: Body cavity

Distribution: Brazil – Queimada Grande Island (Atlantic Forest) and Valinhos (Cerrado), SP;

Jurubatiba and Grussaí (Restinga), RJ; Marechal Floriano (Atlantic Forest) and Praia das Neves (Restinga), ES; Peru – Rio Nanay, LO.

Comments: all aforementioned citations consist of cysts and thus difficult the identification to species level since reproductive structures are required (20, 22-24, 35, 45).

Diplotriaeidae

***Hastospiculum* Skrjabin, 1923**

Hastospiculum sp.

Hosts: *Plica umbra*

Site of infection: Peritoneum

Distribution: Peru – CU (Amazon) (1).

Filariidae

***Filaria* Mueller, 1787**

Filaria multipapilla Molin,

Hosts: *Dracaena guianensis* Daudin, 1802

Site of infection: Body cavity

Distribution: Brazil – Belém (Amazon), PA.

Comments: The species is actually considered *species inquirenda*, because these first records were lost and no new information about the species has been presented since the description (123-125).

Onchocercidae

***Macdonaldius* Khanna, 1933**

Macdonaldius grassi (Caballero, 1954)

Hosts: *Phyllopezus pollicaris* (Spix, 1825)

Site of infection: Underneath the epidermis of the throat

Distribution: Paraguay – CO (Chaco) (126).

***Oswaldofilaria* Travassos, 1933**

Oswaldofilaria azevedoi Bain, 1974

Hosts: *Polychrus marmoratus* Linnaeus, 1758; *Stenocercus roseiventris*

Site of infection: Body cavity

Distribution: Brazil – Belém (Amazon), PA; Peru – CU (Amazon) (1, 127).

Oswaldofilaria belemensis Bain and Dulahian, 1974

Hosts: *Dracaena guianensis*

Site of infection: Heart, aorta e vena cava

Distribution: Brazil – Belém (Amazon), PA (8).

Oswaldofilaria brevicaudata Rodhain and Vuylsteke, 1937

Hosts: *Iguana iguana*, *Anolis punctatus*

Site of infection: Body cavity
 Distribution: Brazil – Marajó (Amazon), PA, AM (Amazon), Exu (Caatinga), PE; Venezuela – ZU (7, 40, 125, 127-129).

Oswaldofilaria petersi Bain and Sulahian, 1974
 Hosts: *Tupinambis teguixin*, *Tropidurus hispidus*
 Site of infection: Body cavity
 Distribution: Brazil – Belém (Amazon), PA, Ibiraba (Caatinga), BA (8, 130).

Oswaldofilaria spinosa Bain and Sulahian, 1974
 Hosts: *Mabuya bistrata*
 Site of infection: Body cavity
 Distribution: Brazil – Belém (Amazon) (8).

Oswaldofilaria sp.
 Hosts: *Tupinambis teguixin*, *Mabuya frenata*
 Site of infection: Body cavity
 Distribution: Surinam – PR; Brazil – Valinhos (Cerrado), SP (28, 34).

Piratuba Freitas & Lent, 1947
Piratuba digiticauda Lent & Freitas, 1941
 Hosts: *Tropidurus torquatus*, *Tropidurus guarani*, *Plica umbra*, *Kentropyx calcarata*
 Site of infection: Body cavity, intestine
 Distribution: Brazil – Salvador, BA; Piratuba (Amazon), PA; Salobra (Pantanal), MS; Juara (Amazon), MT; Peru – CU; Paraguay – Chaco.
 Comments: *Tropidurus spinulosus* found in Salobra actually correspond to *T. guarani* (1, 62, 63, 65, 125, 131).

Piratuba lainsoni Bain, 1974
 Hosts: *Anolis punctatus*, *Polychrus marmoratus*
 Site of infection: Body cavity, large intestine
 Distribution: Brazil – Belém (Amazon), PA; Peru – CU (Amazon) (1, 127).

Piratuba scaffii Bain, 1974
 Hosts: *Ameiva ameiva*
 Site of infection: Body cavity
 Distribution: Brazil – Belém (Amazon), PA (127).

Piratuba shawi Bain, 1974
 Hosts: *Kentropyx calcarata* Spix, 1825
 Site of infection: Body cavity
 Distribution: Brazil – Belém (Amazon), PA (127).

Piratuboides Bain & Sulahian, 1974
Piratuboides zae (Bain, 1974) Bain and Sulahian,

1974
 Hosts: *Mabuya bistrata* Spix, 1825
 Site of infection: Body cavity, large intestine
 Distribution: Brazil – Belém (Amazon), PA; Peru – CU (Amazon) (1, 127).

Physalopteridae

Abbreviata Travassos, 1920
Abbreviata spiralis (Schneider, 1866) Chabaud, 1956
 Hosts: *Amphisbaena* sp.
 Site of infection: Not informed
 Distribution: Brazil
 Comments: This species is considered a *species inquirenda* (7).

Physaloptera Rudolphi, 1819

Physaloptera lutzi Cristofaro, Guimarães & Rodrigues, 1976
 Hosts: *Ameiva ameiva*; *Cnemidophorus abaetensis* Reis, Dias, Rocha & Vrcibradic, 2002; *C. littoralis*; *Enyalius bilineatus*; *Eurolophosaurus nanuzae*; *Liolaemus alticolor* Barbour 1909; *L. ornatus*; *L. quilmes*; *Tropidurus guarani*; *T. itambere*; *T. torquatus*
 Site of infection: Stomach
 Distribution: Argentina – SA, TU; Bolivia – Roboré (Pantanal); Brazil – Salvador, Serrinha and Canudos (Caatinga), BA; Linhares and Conceição da Barra (Restinga), Marechal Floriano (Atlantic Forest), ES; Serra do Cipó (Cerrado), MG; Porto Esperança and Salobra (Pantanal), MS; Xavantina, MT; Cachimbo (Amazon), PA; Maricá and Jurubatiba (Restinga), RJ; Valinhos (Cerrado), SP; Paraguay – Chaco (8, 24, 31, 32, 35, 62, 65, 66, 94-96, 132, 133).
 Comments: *Cnemidophorus ocellifer*, cited by Ribas *et al.* (94), actually corresponds to *C. littoralis* (31).

Physaloptera retusa Rudolphi, 1819

Hosts: *Ameiva ameiva*; *Ameiva festiva* Lichtenstein, 1856; *Amphisbaena alba*; *Anolis fuscoauratus*; *Anolis punctatus*; *Cnemidophorus abaetensis*; *C. lemniscatus*; *C. littoralis*; *C. native*; *C. ocellifer*; *Enyalius bilineatus*; *Iguana iguana*; *Kentropyx altamazonica*; *K. calcarata*; *K. pelviceps*; *Leiosaurus bellii* Duméril & Bibron, 1837; *L. catamarcensis* Koslowsky, 1898; *Liolaemus lutzae*; *L. neuquensis* Cei & Videla, 2003; *Mabuya agilis*; *M. bistrata*; *M. dorsivittata* Cope, 1862; *Ophiodes striatus* Spix, 1824; *Plica plica*; *P. umbra*; *Potamites*

eupleopus; *Stenocercus roseiventris*; *Thecadactylus solimoensis*; *Tropidurus guarani*; *T. hispidus*; *T. torquatus*; *Tupinambis rufescens* Günther, 1871; *T. longilineus* Ávila-Pires, 1995; *T. teguixin*; *Uracentron flaviceps*; *Tropidurus melanopleurus* Boulenger, 1902

Site of infection: Stomach, intestine

Distribution: Brazil – Salvador and Guaratiba (Restinga), BA; Conceição da Barra (Restinga) and Marechal Floraino (Atlantic Forest), ES; Salobra (Pantanal), MS; Juara (Amazon), MT; Altamira, Belém, Cachimbo, Novo Progresso and Santarém (Amazon), PA; Itatiaia (Atlantic Forest), Maricá and Jurubatiba (Restinga), RJ; Ilha Seca (Atlantic Forest) and Itirapina (Cerrado), SP; Paraguay – Chaco; Bolivia – El Carmen and Florida, SC; Argentina; Peru – CU (Amazon); Surinam – PR (Amazon); Uruguay – Montevideo; Venezuela – Colón, ZU; Atures, AM; Salamanca, NE; Cumuná (Arid zone) and La Orchila, Mar Caribe; Colombia – CH.

Comments: Baker (7) cited several species of lizards that does not occur in Brazil, such as *Tupinambis rufescens*, *Cnemidophorus lateristrigus*, *Euprepis spixii*, *Podinema graphica* and *P. scripta*. According to the same author, *Pygopus gronovii* actually corresponds to *Ophiodes striatus* and *Podinema* corresponds to *Tupinambis*, but no species are currently recognized as synonyms of *P. scripta* and *P. graphica* (7). *Cnemidophorus ocellifer*, cited by Ribas *et al.* (94), actually corresponds to *C. littoralis* (1, 8, 24, 28, 29, 31, 32, 35, 51, 57, 62, 69, 77, 94, 108, 126, 131, 134-142).

Physaloptera sp.

Hosts: *Ameiva ameiva*; *Cercosaura argulus*; *Cnemidophorus littoralis*; *C. ocellifer*; *Hemidactylus mabouia*; *Hoplocercus spinosus*; *Mabuya agilis*; *M. macrorhyncha*; *Polychrus acutirostris*; *Tropidurus etheridgei*; *T. torquatus*; *Tupinambis meriana* Duméril & Bibron, 1839; *Tupinambis teguixin*; *Dicrodon guttulatum*.

Site of infection: Stomach

Distribution: Brazil – Abrolhos and Salvador (Restinga), BA; Linhares and Praia das Neves (Restinga) and Sooretama (Cerrado), ES; Salobra (Pantanal) and Bodoquena (Cerrado), MS; Ilha Grande, Cabo Frio, Grussaí, Grumari and Jurubatiba (Restinga), RJ; Valinhos (Cerrado), SP; Alegria (Pampas), RS; Argentina – SA and FO (Chaco); Peru – Rio Nanay, LO, LI.

Comments: *Tupinambis teguixin* of Salobra,

MS corresponds to *T. meriana* (personal observation) (8, 16, 22, 31, 33, 35, 41, 45, 92, 96, 101, 102, 143).

Physalopteroides Wu & Liu, 1940

Physalopteroides venancioi Lent, Freitas & Proença, 1946

Hosts: *Alopoglossus atriventris* Duellman, 1973; *Ameiva ameiva*; *Cercosaura ocellata* Wagler, 1830; *Cnemidophorus native*; *Kentropyx altamazonica*; *K. calcarata*; *K. pelviceps*; *Mabuya agilis*; *M. bistrata*; *M. macrorhyncha*; *Thecadactylus solimoensis*; *Tropidurus torquatus*; *Tupinambis teguixin*

Site of infection: Stomach

Distribution: Brazil – AC (Amazon); Trancoso and Guaratiba (Restinga), BA; Juara (Amazon), MT; Magé, Grussaí and Jurubatiba (Restinga), RJ; Praia das Neves (Restinga), ES; Peru – CU (Amazon) (1, 17, 23, 33, 35, 52, 77, 131, 144).

Thubunaea Seurat, 1914

Thubunaea iguanae (Telford, 1965)

Hosts: *Microlophus peruvianus*; *M. thoracicus* (Tschudi, 1845)

Site of infection: Stomach

Distribution: Peru – R. N. Paracas, IC (117).

Thubunaea parkeri Baylis, 1926

Hosts: *Microlophus occipitalis* Peters, 1871; *Dicrodon heterolepis*

Site of infection: Stomach

Distribution: Peru (7, 145).

Spiruridae

Spirurinae

Hosts: *Hemidactylus mabouia*

Site of infection: Stomach, lungs, and small intestine

Distribution: Brazil – Rio de Janeiro, RJ (146).

Micropleuridae

Micropleura Linstow, 1906

Hosts: *Tupinambis meriana*

Site of infection: Body cavity

Distribution: Brazil – Salobra (Pantanal), MS.

Comments: This record must be discharged, because no mention of *Micropleura* was found in Travassos and Freitas (147) as cited by Vicente *et al.* (8). Moreover, *Micropleura* may be found in the body cavity of crocodiles and turtles (148).

Diaphanocephalidae

***Diaphanocephalus* Diesing, 1851**

Diaphanocephalus diesingi Freitas & Lent, 1938

Hosts: *Tupinambis teguixin*

Site of infection: Small intestine

Distribution: Brazil – Belém (Amazon), PA (149).

Diaphanocephalus galeatus Rudolphi, 1819

Hosts: *Tupinambis merianae*, *T. rufescens*, *T. teguixin*

Site of infection: Intestine

Distribution: Brazil – Salobra (Pantanal) and Bodoquena (Cerrado), MS; Manguinhos, Angra dos Reis e Rio de Janeiro (Atlantic Forest), RJ; Pedras Altas, RS; Ilha Grande and São Paulo, SP; Fernando de Noronha, PE; Argentina; Bolivia; Surinam – PR (8, 28, 46, 149, 150).

Diaphanocephalus jacuruxi Alho, 1965

Hosts: *Dracaena guianensis*

Site of infection: Small intestine

Distribution: Brazil – Ilha das Onças (Amazon), PA (8, 151).

***Kalicephalus* Molin, 1861**

Kalicephalus sp.

Hosts: *Tupinambis teguixin*

Site of infection: Intestine

Distribution: Brazil – Sooretama, ES; Alegria (Pampas), RS.

Comments: Species of *Kalicephalus* spp. have been reported in several snake species, and seems to be restricted to these animals, likewise *Diaphanocephalus* spp. (a sister taxon) in Tupinambinae. Published records could be a misidentification or a case of incidental infection in a *Tupinambis teguixin* (8, 143, 152).

Molineidae

***Kentropyxia* Baker, 1982**

Kentropyxia sauria Baker, 1982

Hosts: *Kentropyx calcarata*

Site of infection: Small intestine

Distribution: Brazil – Belém and Novo Progresso (Amazon), PA (136, 153).

***Oswaldocruzia* Travassos, 1917**

Oswaldocruzia bainaie Ben-Slimane & Durette-Desset, 1996

Hosts: *Anolis chrysolepis* Duméril & Bibron, 1837; *A. fuscoauratus*

Site of infection: Small intestine

Distribution: Ecuador – San Pablo (Amazon), LR (154).

Oswaldocruzia benslimanei Durette-Desset, Anjos & Vrcibradic, 2006

Hosts: *Enyalius bilineatus* Duméril & Bibron, 1837

Site of infection: Small intestine

Distribution: Brazil – Marechal Floriano, ES (24, 155).

Oswaldocruzia brasiliensis Lent & Freitas, 1935

Hosts: *Hemidactylus mabouia*

Site of infection: Small intestine

Distribution: Brasil – Rio de Janeiro, RJ (8, 156).

Oswaldocruzia burseyi Durette-Desset, Anjos & Vrcibradic, 2006

Hosts: *Enyalius perditus*

Site of infection: Stomach

Distribution: Brazil – São Sebastião (Atlantic Forest), SP (3, 155).

Oswaldocruzia fredii Durette-Desset, Anjos & Vrcibradic, 2006

Hosts: *Enyalius iheringii*

Site of infection: Stomach

Distribution: Brazil – São Sebastião (Atlantic Forest), SP (3, 155).

Oswaldocruzia peruensis Ben-Slimane, Verhaag & Durette-Desset, 1995

Hosts: *Stenocercus roseiventris*, *Cercosaura argulus*, *Anolis punctatus*

Site of infection: Stomach

Distribution: Peru – HA and CU (Amazon) (1, 45, 157).

Oswaldocruzia vittii Bursey & Goldberg, 2004

Hosts: *Alopoglossus angulatus*, *A. atriventris*, *Anolis fuscoauratus*, *A. punctatus*, *Cercosaura eigenmanni*, *C. oshaugnessyi*

Site of infection: Intestine

Distribution: Brazil – AC, AM, PA, RO (Amazon); Ecuador – SU (Amazon); Peru – CU (Amazon) (1, 18, 40, 51, 52).

Oswaldocruzia sp.

Hosts: *Tropidurus torquatus*, *Enyalius catenatus* (Wied, 1821), *E. perditus*, *K. calcarata*

Site of infection: Intestine

Distribution: Brazil – Bodoquena (Cerrado)

and Salobra (Pantanal), MS; Juara (Amazon), MT; Cachimbo (Amazon), PA; Rio de Janeiro, RJ; Ibitipoca, MG; Argentina – SA, JU and TA (Chaco); Paraguay – Assuncion and Chaco.

Comments: The aforementioned records of *T. torquatus* are those of *O. mazzai* and that of *Enyalius* spp. are from *O. subauricularis*, both considered *species inquirenda* (48, 131, 155, 158).

Cestoda

Cyclophyllidea

Linstowiidae

***Oochoristica* Lühe, 1898**

Oochoristica ameivae (Beddard, 1914)

Hosts: *Ameiva ameiva*, *Cnemidophorus natio*, *Mabuya agilis*, *M. macrorhyncha*

Site of infection: Intestine

Distribution: Brazil – Cachimbo (Amazon), PA; Serra do Navio (Amazon), AP; Manaus (Amazon), AM; Guaratiba and Trancoso (Restinga), BA; Praia das Neves (Restinga), ES; Grussaí (Restinga), RJ; Peru – CU (Amazon) (1, 17, 23, 77, 159-161).

Oochoristica bressalui Fürhmann, 1927

Hosts: *Tropidurus guarani*, *T. hispidus*, *T. torquatus*

Site of infection: Small intestine

Distribution: Brazil – Salvador and Serrinha, BA; Cachimbo (Amazon), Arraial do Cabo, Cabo Frio and Rio de Janeiro, RJ; Bolivia – El Carmen; Paraguay (19, 65, 160, 162).

Oochoristica freitasi Rego & Ibañez, 1965

Hosts: *Dicrodon heterolepis*; *D. guttulatum* Duméril & Bibron, 1893

Site of infection: Small intestine

Distribution: Peru – Trujillo, LL; TU, PI.

Comments: Bursey and Goldberg (163) and Arizmendi-Espinosa *et al.* (164) apparently ignored the presence of *O. freitasi* in their findings of *Oochoristica* spp. infecting lizards in neotropical regions. The species is not included in any synonymy of the revisions made by Bursey *et al.* (165). Moreover, the species was cited in Guillén-Hernández *et al.* (166).

Oochoristica iguanae (Baylis, 1919) Bursey & Goldberg, 1996

Hosts: *Iguana iguana*, *Dicrodon guttulatum*

Site of infection: Small intestine

Distribution: Venezuela – Isla Margarita; Peru – LA, PI (92,163).

Oochoristica travassosi Rego & Ibañez, 1965

Hosts: *Leiocephalus* sp.; *Liolaemus vallecurensis* Pereira, 1992; *Phyllodactylus johnwrigthi* Dixon & Huey, 1970

Site of infection: Small intestine

Distribution: Peru – Moche; Argentina – SJ (2, 97, 167).

Oochoristica insulamargaritae López-Neyra and Diaz-Ungría, 1957

Hosts: *Ameiva ameiva*

Site of infection: Small intestine

Distribution: Venezuela – Isla Margarita (163, 168).

Oochoristica vanzolinii Rego & Rodrigues, 1965

Hosts: *Hemidactylus mabouia*, *Eurolophosaurus nanuzae*

Site of infection: Small intestine

Distribution: Brazil – Rio de Janeiro, RJ; Serra do Cipó, MG (66, 162).

Oochoristica sp.

Hosts: *Alopoglossus atriventris*, *Amphisbaena ridleyi*, *Mabuya frenata*, *Tupinambis merianae*, *Trachylepis atlantica*

Site of infection: Small intestine

Distribution: Ecuador – SU; Brazil – Valinhos, SP; Fernando de Noronha Islands, PE (34, 46, 52).

***Semenoviella* Spasskii, 1951**

Semenoviella amphisbaenae Rudolphi, 1819

Hosts: *Amphisbaena fuliginosa* Linnaeus, 1758; *A. alba*

Site of infection: Intestine

Distribution: Brazil – Belém (Amazon), PA; Pirassununga (Cerrado), SP (169).

Mesocestoididae

***Mesocestoides* Vaillant, 1863**

Mesocestoides sp.

Hosts: *Anolis transversalis*

Site of infection: Not informed

Distribution: Brazil – RO (Amazon) (40).

Proteocephalidea

Proteocephalidae

***Cairaella* Coquille & De Chambrier, 2008**

Cairaella henrii Coquille & De Chambrier, 2008

Hosts: *Anolis trachyderma* Cope, 1876

Site of infection: Intestine

Distribution: Ecuador – San Pablo de Kantesyia, SU (170).

Ophiotaenia La Rue, 1911

Ophiotaenia flava Rudin, 1917

Hosts: *Kentropyx pelviceps*

Site of infection: Not informed

Distribution: Peru – CU (Amazon) (1).

Ophiotaenia nicoleae Coquille & De Chambrier, 2008

Hosts: *Thecadactylus solimoensis*

Site of infection: Intestine

Distribution: Ecuador – San Pablo de Kantesyia, SU.

Comments: *Thecadactylus* was recently subjected to revision, and a *T. solimoensis* was found in from localities that encompass San Pablo de Kantesyia, Ecuador (113, 170).

Ophiotaenia sp.

Hosts: *Anolis fuscoauratus*

Site of infection: Small intestine

Distribution: Brazil – Santarém (Amazon), PA (51).

Tejidotaenia Freze, 1965

Tejidotaenia appendiculata (Baylis, 1947)

Hosts: *Tupinambis teguixin*

Site of infection: Small intestine

Distribution: Surinam – PR (Amazon); Brazil – Serra do Navio (Amazon), AP; Linhares, ES (28, 171).

Trematoda

Digenea

Echinostomata

Echinostomida

Superfamily Echinostomatoidea Looss, 1899

Cathaemasiidae

Pulchrossomoides Freitas & Lent, 1937

Pulchrossomoides elegans Freitas & Lent, 1937

Hosts: *Tupinambis teguixin*, *Iguana iguana*, *Mabuya macrorhyncha*

Site of infection: Stomach

Distribution: Brazil – Porto Esperança (Pantanal), MS; Praia das Neves (Restinga), ES (6, 23, 172, 173).

Plagiorchiida

Dicrocoeliidae

Brachycoelium (Dujardin, 1845)

Brachycoelium salamandrae (Frolich, 1789) Dujardin, 1845

Hosts: *Leposoma osvaldoi* Ávila-Pires, 1995; *Anolis nitens* Wagler, 1830

Site of infection: Small intestine

Distribution: Brazil – Novo Progresso (Amazon), PA.

Comments: Cited as *B. mesocoeliiformis* when found infecting *Anolis scypheus* (*A. nitens*) (6, 137).

Platynosomum

Platynosomum sp. Looss, 1906

Hosts: *Amphisbaena ridleyi*, *Trachylepis atlantica*

Site of infection: Gallbladder, bile duct

Distribution: Brazil – Fernando de Noronha Island, PE (46).

Harmotrematidae

Helicotrema Odhner, 1912

Helicotrema asymmetricum (Travassos, 1922) Viana, 1924

Hosts: *Iguana iguana*

Site of infection: Intestine

Distribution: Brazil – Pantanal (174, 175).

Helicotrema magniovatum Odhner, 1912

Hosts: *Iguana iguana*

Site of infection: Intestine

Distribution: Brazil (6).

Helicotrema spirale (Diesing, 1850) Odhner, 1912

Hosts: *Iguana iguana*

Site of infection: Intestine

Distribution: Brazil (6).

Pronocephalidae

Iguanacola Gilbert, 1938

Iguanacola navicularis Gilbert, 1938

Hosts: *Amblyrhynchus cristatus* Bell, 1825

Site of infection: Not informed

Distribution: Ecuador – Galapagos Islands (14).

Myosaccus Gilbert, 1938

Myosaccus amblyrhynchi Gilbert, 1938

Hosts: *Amblyrhynchus cristatus*

Site of infection: Not informed

Distribution: Ecuador – Galapagos Islands (14).

Cetiosaccus Gilbert, 1938

Cetiosaccus galapagensis Gilbert, 1938

Hosts: *Amblyrhynchus cristatus*
 Site of infection: Not informed
 Distribution: Ecuador – Galapagos Islands (14).

Paradistomum Kossack, 1910

Paradistomum parvissimum (Travassos, 1918)
 Hosts: *Ameiva ameiva*, *Hemidactylus mabouia*,
Iguana iguana, *Liolaemus lutzae*, *Mabuya*
macrorhyncha, *M. agilis*, *Tropidurus torquatus*,
Tupinambis teguixin

Site of infection: Gallbladder, liver, small intestine
 Distribution: Brazil – Arraial do Cabo, Grussaí,
 Nova Iguaçu, Manguinhos, Maricá, RJ; Praia das
 Neves (Restinga) and Santa Teresa, ES; Salvador
 and Trancoso (Restinga), BA (17, 19, 21, 23, 27,
 100, 146, 176, 177).

Paradistomum rabusculum Kossack, 1910

Hosts and records: *Gymnodactylus geckoides* Spix,
 1825
 Site of infection: Not informed
 Distribution: Brazil (6).

Macroderidae

***Pneumotrema* Bhalerao, 1937**

Pneumotrema travassosi Bhalerao, 1937
 Hosts and records: *Amphisbaena alba*
 Site of infection: Kidney, ureter, intestine.
 Distribution: Brazil (172).

Family Plagiorchiidae

***Allopharynx* (Strom, 1928)**

Allopharynx daileyi Bursey, Goldberg & Vitt, 2005
 Hosts: *Uranoscodon superciliosus*
 Site of infection: Small intestine
 Distribution: Brazil – PA, RO, RR (Amazon) (50).

***Dasymetra* Nicoll, 1911**

Dasymetra tupinambis Nasir & Diaz, 1971
 Hosts: *Tupinambis teguixin*
 Site of infection: Intestine
 Distribution: Venezuela – Sucre (178).

***Styphlodora* Looss, 1899**

Styphlodora condita Faria, 1911
 Hosts: *Tupinambis rufescens* (Günther, 1871)
 Site of infection: Kidney, ureter, intestine
 Distribution: Argentina (9).

***Plagiorchis* Lühe, 1899**

Plagiorchis freitasi Vicente, 1978
 Hosts: *Tropidurus torquatus*

Site of infection: Small intestine
 Distribution: Brazil – Conceição da Barra, ES
 (19).

Plagiorchis vicentei Rodrigues, 1994

Hosts: *Hemidactylus mabouia*
 Site of infection: Small intestine
 Distribution: Brazil – Teresópolis, RJ (179).

Family Brachycoeliidae

***Mesocoelium* Odhner, 1910**

Mesocoelium monas (Rudolphi, 1819) Freitas,
 1958

Hosts: *Alopoglossus angulatus*, *Amphisbaena* sp.,
Amphisbaena ridleyi, *Cercosaura eigenmanni*,
Diploglossus lessonae, *Trachylepis atlantica*,
Leposternon microcephalum, *Tropidurus*
torquatus, *Uranoscodon superciliosus*

Site of infection: Intestine
 Distribution: Brazil – PA, RO, RR (Amazon); João
 Alfredo (Caatinga) and Fernando de Noronha,
 PE; Maricá and Rio de Janeiro (Restinga), RJ;
 Ecuador – SU (Amazon) (6, 18, 46, 50, 52, 100).

Family Urotrematidae

Urotrema Braun, 1900

Urotrema shirleyae Zamparo, Brooks & Tkach,
 2005

Hosts: *Anolis fuscoauratus*
 Site of infection: Small intestine
 Distribution: Brazil – Santarém (Amazon), PA
 (51).

HOST-PARASITE LIST

Family Amphisbaenidae

Amphisbaena alba

Maracaya belemensis
Maracaya graciai
Aplectana albae
Aplectana raillieti
Physaloptera retusa
Semenoviella amphisbaenae
Pneumotrema travassosi

Amphisbaena bolivica

Aplectana tucumanensis

Amphisbaena fuliginosa

Semenoviella amphisbaenae

Amphisbaena ridleyi

Aplectana albae
Parapharyngodon alvarengai
Mesocoelium monas
Platynosomum sp.
Oochoristica sp.

***Amphisbaena* sp.**
Maracaya pusilla
Mesocoelium monas

Leposternon microcephalum
Cosmocerca rara
Mesocoelium monas
Paradollfusnema amphisbaenia

Family Iguanidae

Amblyrhynchus cristatus
Iguanacola navicularis
Myosaccus amblyrhynchi
Cetiosaccus galapagensis

Iguana iguana
Alaeuris caudatus
Alaeuris vogelsangi
Ozolaimus cirratus
Ozolaimus megatyphlon
Oswaldofilaria brevicaudata
Physaloptera retusa
Oochoristica iguanae
Pulchrosomoides elegans
Paradistomum parvissimum
Helicotrema asymmetricum
Helicotrema magniovatum
Helicotrema spirale

Conolophus subcristatus
Alaeuris conolophi
Alaeuris galapagensis
Alaeuris labicula
Alaeuris longispicula
Paralaeuris dorochila

Family Hoplocercidae

Enyalioides praestabilis
Spinicauda spinicauda

Hoplocercus spinosus
Africana dardanelosi
Cruzia rudolphii
Physaloptera sp.

Morunasaurus annularis

Raillietnema brachyspiculatum

Family Polychrotidae

Anisolepis undulatus
Rhabdias sp.

Anolis chrysolepis
Oswaldocruzia bainaie

Anolis fuscoauratus
Cosmocerca vrcibradici
Strongyluris oscar
Rhabdias sp.
Physaloptera retusa
Oswaldocruzia bainaie
Oswaldocruzia vitti
Ophiotaenia sp.
Urotrema shirleyae

Anolis nitens
Brachycoelium salamandrae

Anolis punctatus
Cyrtosomum longicaudatum
Strongyluris oscar
Falcaustra sp.
Skrjabinellazia intermedia
Rhabdias anolis
Rhabdias elegans
Rhabdias sp.
Oswaldofilaria brevicaudata
Piratuba lainsoni
Physaloptera retusa
Oswaldocruzia peruensis
Oswaldocruzia vitti

Anolis trachyderma
Cairaella henrii

Anolis transversalis
Cyrtosomum longicaudatum
Strongyluris oscar
Mesocestoides sp.

Polychrus acutirostris
Gynaecometra bahiensis
Physaloptera sp.

Polychrus marmoratus
Pseudostrongyluris polychrus
Oswaldofilaria azevedoi
Piratuba lainsoni

Family Leiosauridae

Enyalius bilineatus

Centrorhynchidae

Rhabdias sp.

Acuariidae

Physaloptera lutzi

Physaloptera retusa

Oswaldocruzia benslimanei

Enyalius catenatus

Oswaldocruzia sp.

Enyalius iheringii

Strongyluris oscari

Rhabdias sp.

Oswaldocruzia fredii

Enyalius perditus

Acanthocephalus sp.

Aplectana vellardi

Cosmocerca sp.

Strongyluris oscari

Rhabdias sp.

Oswaldocruzia burseyi

Oswaldocruzia sp.

Leiosaurus belli

Physaloptera retusa

Leiosaurus catamarcensis

Physaloptera retusa

Family Liolaemidae

Liolaemus alticolor

Physaloptera lutzi

Liolaemus andinus

Spauligodon maytacapaci

Liolaemus buergeri

Parapharyngodon riojensis

Liolaemus capillitas

Spauligodon lobo

Liolaemus chilensis

Spauligodon maytacapaci

Liolaemus elongatus

Spauligodon maytacapaci

Liolaemus huacahuasicus

Spauligodon lobo

Liolaemus lemniscatus

Spauligodon maytacapaci

Liolaemus lutzae

Ichthyocapillaria (= *Pseudocapillaria*) *maricaensis*

Parapharyngodon sceleratus

Pharyngodon cesarpintoi

Physaloptera retusa

Paradistomum parvissimum

Liolaemus neuquensis

Physaloptera retusa

Liolaemus ornatus

Spauligodon lobo

Physaloptera lutzi

Liolaemus pictus

Spauligodon maytacapaci

Liolaemus quilmes

Spauligodon lobo

Physaloptera lutzi

Liolaemus ramirezae

Spauligodon lobo

Liolaemus signifer

Parapharyngodon senisfaciecaudus

Liolaemus tenuis

Spauligodon maytacapaci

Liolaemus vallecurensis

Oochoristica travassosi

Phymaturus palluma

Parapharyngodon riojensis

Phymaturus punae

Parapharyngodon riojensis

Family Tropiduridae

Eurolophosaurus nanuzae

Strongyluris oscari

Subulura lacertilia

Parapharyngodon sceleratus

Physaloptera lutzi

Oochoristica vanzolinii

***Leiocephalus* sp.**

Spauligodon maytacapaci
Oochoristica travassosi

Microlophus albermalensis

Parapharyngodon sceleratus

Microlophus occipitalis

Thubunaea parkeri

Microlophus peruvianus

Parapharyngodon arequipensis
Parapharyngodon moqueguensis
Parapharyngodon yurensis
Thubunaea iguanae

Microlophus thoracicus

Thubunaea iguanae

Plica plica

Strongyluris oscari
Physaloptera retusa

Plica umbra

Strongyluris oscari
Hastospiculum sp.
Piratuba digiticauda
Physaloptera retusa

Stenocercus caducus

Skrjabinellazia intermedia
Strongyluris oscari

Stenocercus roseiventris

Atractis cruciata
Strongyluris oscari
Oswaldofilaria azevedoi
Physaloptera retusa
Oswaldocruzia peruensis

Tropidurus etheridgei

Parapharyngodon sp.
Thelandros sp.
Physaloptera sp.

Tropidurus guarani

Strongyluris oscari
Skrjabinellazia intermedia
Typhlonema sp.
Parapharyngodon sceleratus
Spauligodon oxkutzcabiensis
Piratuba digiticauda

Physaloptera lutzi

Physaloptera retusa

Oochoristica bressalui

Tropidurus hispidus

Parapharyngodon sceleratus
Pharyngodon sp.
Rhabdias sp.
Oswaldofilaria petersi
Physaloptera retusa
Oochoristica bressalui

Tropidurus itambere

Parapharyngodon sceleratus
Pharyngodon cesarpintoi
Physaloptera lutzi

Tropidurus melanopleurus

Strongyluris oscari
Parapharyngodon sceleratus
Physaloptera retusa

Tropidurus semitaeniatus

Parapharyngodon sceleratus

Tropidurus spinulosus

Strongyluris oscari

Tropidurus torquatus

Acanthocephala
Echinorhynchus sp.
Centrorhynchus tumidulus
Capillaria (= *Amphibiocapillaria*) *freitaslenti*
Hexametra boddaertii
Strongyluris oscari
Skrjabinellazia intermedia
Subulura sp.
Parapharyngodon sceleratus
Parapharyngodon sp.
Pharyngodon sp.
Acuariidae
Piratuba digiticauda
Physaloptera lutzi
Physaloptera retusa
Physaloptera sp.
Physalopteroides venancioi
Oswaldocruzia sp.
Oochoristica bressalui
Paradistomum parvissimum
Plagiorchis freitasi
Mesocoelium monas

Tropidurus sp.

Strongyluris oscar
Parapharyngodon sceleratus

Uracentron flaviceps

Skrjabinodon dixon
Physaloptera retusa

Uranoscodon superciliosus

Cosmocerca vrcibradici
Africana chabaudi
Allopharynx dailey
Mesocoelium monas

Family Gekkonidae

Hemidactylus mabouia

Centrorhynchidae
Parapharyngodon largitor
Parapharyngodon sceleratus
Parapharyngodon sp
Strongyloides cruzi
Acuariidae
Physaloptera sp.
Spirurinae
Oswaldocruzia brasiliensis
Oochoristica vanzolinii
Paradistomum parvissimum
Plagiorchis vicentei

Family Phyllodactylidae

Bogertia lutzae

Spauligodon oxkutzcabiensis

Gymnodactylus geckoides

Paradistomum rabusculum

Phyllodactylus angustidigitus

Spauligodon viracochai

Phyllodactylus inaequalis

Spauligodon oxkutzcabiensis

Phyllodactylus gerrhopygus

Spauligodon viracochai

Phyllodactylus johnwrighti

Oochoristica travassosi
Parapharyngodon sceleratus
Spauligodon oxkutzcabiensis

Phyllodactylus lepidopygus

Acuariidae

Phyllodactylus microphyllus

Spauligodon oxkutzcabiensis

Phyllodactylus reissii

Spauligodon oxkutzcabiensis

Phyllopezus pollicaris

Macdonaldius grass
Parapharyngodon sceleratus

Thecadactylus solimoensis

Spauligodon oxkutzcabiensis
Physaloptera retusa
Physalopteroides venancioi
Ophiotaenia nicoleae

Family Sphaerodactylidae

Gonatodes humeralis

Skrjabinelazia galliardi

Family Anguidae

Diploglossus lessonae

Parapharyngodon verrucosus
Mesocoelium monas

Ophiodes striatus

Cyrtosomum sp.
Physaloptera retusa

Family Teiidae

Ameiva ameiva

Oligacanthorhynchus sp.
Centrorhynchus tumidulus
Capillaria (= Amphibiocapillaria) freitaslenti
Spinicauda spinicauda
Strongyluris oscar
Parapharyngodon alvarengai
Parapharyngodon largitor
Parapharyngodon sceleratus
Parapharyngodon sp.
Pharyngodon cesarpinto
Pharyngodon travassosi
Chabirenina cayennensis
Piratuba scaff
Physaloptera lutzi
Physaloptera retusa
Physaloptera sp.
Physalopteroides venancioi
Oochoristica ameivae
Oochoristica insulamargaritae
Paradistomum parvissimum

Ameiva festiva
Physaloptera retusa

Cnemidophorus abaetensis
Physaloptera lutzi
Physaloptera retusa

Cnemidophorus lemniscatus
Physaloptera retusa

Cnemidophorus littoralis
Hexametra boddaertii
Parapharyngodon sceleratus
Physaloptera lutzPhysaloptera retusa
Physaloptera sp.

Cnemidophorus nativo
Skrjabinellazia intermedia
Subulura lacertilia
Parapharyngodon sp.
Physaloptera retusa
Physalopteroides venancioi
Oochoristica ameivae

Cnemidophorus ocellifer
Hexametra boddaertii
Physaloptera retusa
Physaloptera sp.

Cnemidophorus sp.
Pharyngodon cesarpintoii

Dicrodon heterolepis
Pharyngodon micrurus
Thubunaea parkeri
Oochoristica freitasi

Dicrodon guttulatatum
Oochoristica freitasi
Oochoristica iguanae
Pharyngodon micrurus
Parapharyngodon moqueguensis
Parapharyngodon lagitor
Parapharyngodon sp.
Thelandros capacityupanquii
Physaloptera sp.

Dicrodon holmbergi
Thelandros capacityupanquii

Dracaena guianensis
Oswaldofilaria belemensis

Diaphanocephalus jacuruxi

Kentropyx altamazonica
Physaloptera retusa
Physalopteroides venancioi

Kentropyx calcarata
Kentropyxia sauria
Piratuba digiticauda
Piratuba shawi
Physaloptera retusa
Physalopteroides venancioi
Oswaldocruzia sp.

Kentropyx pelviceps
Dujardinascaris sp.
Parapharyngodon sceleratus
Physaloptera retusa
Physalopteroides venancioi
Ophiotaenia flava

Tupinambis longilineus
Physaloptera retusa

Tupinambis merianae
Cruzia travassosi
Physaloptera sp.
Diaphanocephalus galeatus
Spinicauda spinicauda
Oochoristica sp.

Tupinambis rufescens
Physaloptera retusa
Diaphanocephalus galeatus
Styphlodora condita

Tupinambis teguixin
Centrorhynchus tumidulus
Centrorhynchus sp.
Contraecum sp.
Dujardinascaris sp.
Freitasascaris alata
Atractis cruciata
Spinicauda spinicauda
Cruzia fulleborni
Cruzia tentaculata
Oswaldofilaria petersi
Oswaldofilaria sp.
Physaloptera retusa
Physaloptera sp.
Physalopteroides venancioi
Diaphanocephalus diesingi

Diaphanocephalus galeatus
Tejidotaenia appendiculata
Pulchrosomoides elegans
Paradistomum parvissimum
Dasymetra tupinambis

Family Gymnophthalmidae

Alopoglossus angulatus
Cosmocerca vrcibradici
Oswaldocruzia vitti
Mesocoelium monas

Alopoglossus atriventris
Cosmocerca vrcibradici
Physalopteroides venancioi
Oswaldocruzia vitti
Oochoristica sp.

Cercosaura argulus
 Cosmocercidae
 Acuariidae
Physaloptera sp.
Oswaldocruzia peruensis

Cercosaura eigenmanni
Cosmocerca vrcibradici
Oswaldocruzia vitti
Mesocoelium monas

Cercosaura ocellata
Physalopteroides venancioi

Cercosaura oshaugnessyi
Acanthocephalus saurius
Cosmocerca vrcibradici
Oswaldocruzia vitti

Leposoma osvaldoi
Brachycoelium salamandrae

Neusticurus bicarinatus
Falcaustra belemensis

Potamites ecleopus
Falcaustra belemensis
Physaloptera retusa

Potamites strangulatus
Falcaustra belemensis

Family Scincidae

Mabuya agilis

Centrorhynchidae
Hexametra boddaertii
Strongyluris oscari
Parapharyngodon largitor
Parapharyngodon sceleratus
 Acuariidae
Physaloptera retusa
Physaloptera sp.
Physalopteroides venancioi
Oochoristica ameivae
Paradistomum parvissimum

Trachylepis atlantica
Moaciria alvarengai
Parapharyngodon alvarengai
Mesocoelium monas
Spinicauda spinicauda
Platynosomum sp.
Oochoristica sp.

Mabuya bistrata
Parapharyngodon sceleratus
Oswaldofilaria spinosa
Piratuboides zeae
Physaloptera retusa
Physalopteroides venancioi

Mabuya caissara
Parapharyngodon sceleratus

Mabuya dorsivittata
Skrjabinodon spinulosus
Physaloptera retusa

Mabuya frenata
Hexametra boddaertii
Parapharyngodon sceleratus
Skrjabinodon heliocostai
Oswaldofilaria sp.
Oochoristica sp.

Mabuya macrorhyncha
 Echinorhynchidae
 Centrorhynchidae
Hexametra boddaertii
 Acuariidae
Physaloptera sp.
Physalopteroides venancioi
Oochoristica ameivae
Pulchrosomoides elegans
Paradistomum parvissimum

FINAL REMARKS

The literature on helminth parasites of lizards and amphisbaenians from South America is extensive, dating back to the 1920's with the studies by Lauro Travassos. However, the information is scattered, and reviews of helminth parasites are necessary because they provide a basis for a better understanding on parasitism in these groups for future research (180). The need of updated knowledge becomes clear considering the valuable information that parasites provide because it is synergistic with all the information about the natural history of the hosts (181).

Helminths are high diverse in vertebrates, but the species richness are poorly known (182, 183). For example, less than a half of nematode species are believed to be currently described (148). In this paper, a total of 644 records of helminths collected from lizards are reported. Since many species descriptions were made in the last decade, the diversity of helminth parasites of lizards from South America should be increased with further studies.

Of the 644 records in the present study, at least nine species of acantocephalans, 15 cestodes, 20 trematodes and 111 nematodes were reported, including records in which family or genus were not identified. From the 155 helminth species reported, one was recorded from Chile and French Guiana, three were from Colombia, three were from Uruguay, eight were from Bolivia, nine were from Surinam, 13 were from Paraguay, 12 were from Venezuela, 27 were from Ecuador, 17 were from Argentina, 39 were from Peru and 103 were from Brazil. Five species considered *species inquirenda* were also reported.

Lizards and amphisbaenians comprises approximately 420 genera and more than 3,300 species, most of which can be found in South America (184, 185). Many South American countries exhibit great lizard diversity, for example Argentina presents 167 species while Brazil has 240 species, but data presented herein includes only 114 lizard species, which indicates that much work has to be done (186, 187).

ACKNOWLEDGEMENTS

Both authors are thankful for the financial support of The State of São Paulo Research Foundation (FAPESP) and Robson W. Ávila is grateful for the grant provided by Coordination for the

Improvement of Higher Education Personnel (CAPES).

COPYRIGHT

© CEVAP 2010

SUBMISSION STATUS

Received: June 1, 2010.

Accepted: September 16, 2010.

Abstract published online: September 17, 2010.

Full paper published online: November 30, 2010.

CONFLICTS OF INTEREST

There is no conflict.

FINANCIAL SOURCE

FAPESP (processes n. 04/03628-1 and 06/59692-5) and CAPES provided the financial grants.

CORRESPONDENCE TO

ROBSON WALDEMAR ÁVILA, Departamento de Parasitologia, Instituto de Biociências, UNESP, Distrito de Rubião Jr., s/n, CEP 18618-000, Botucatu, SP, Brazil. Email: robsonavila@gmail.com.

REFERENCES

1. Bursey CR, Goldberg SR, Parmelee JR. Gastrointestinal helminths from 13 species of lizards from Reserva Cuzco Amazónico, Peru. *Comp Parasitol.* 2005;72(1):50-68.
2. Goldberg SR, Bursey CR, Morando M. Metazoan endoparasites of 12 species of lizards from Argentina. *Comp Parasitol.* 2004;71(2):208-14.
3. Vrcibradic D, Anjos LA, Vicente JJ, Bursey CR. Helminth parasites of two sympatric lizards, *Enyalius iheringii* and *E. perditus* (Leiosauridae), from an Atlantic Rainforest area of southeastern Brazil. *Acta Parasitol.* 2008;53(2):222-5.
4. Travassos LP. Contribuições para o conhecimento da fauna helmintológica brasileira. IX. Sobre as espécies do gênero *Spinicauda*. *Mem Inst Oswaldo Cruz.* 1920;12:41-50.
5. Travassos LP. Pesquisas helminthológicas realizadas em Hamburgo. IX. Ensaio monográfico da família Cosmoceridae Travassos, 1925 (Nematoda). *Mem Inst Oswaldo Cruz.* 1931;25:237-98.
6. Travassos LP, Freitas JFT, Kohn A. Trematodeos do Brasil. *Mem Inst Oswaldo Cruz* 1969;67:1-886.
7. Baker MR. Synopsis of the Nematoda parasitic in amphibians and reptiles. *Mem Univ Newfoundland Occas Pap Biol.* 1987;11:1-325.

8. Vicente JJ, Rodrigues HO, Gomes DC, Pinto RM. Nematóides do Brasil. Parte III: Nematóides de répteis. Rev Bras Zool. 1993;10(1):19-168.
9. Lunaschi LI, Drago FB. Checklist of digenean parasites of amphibians and reptiles from Argentina. Zootaxa. 2007;1476:51-68.
10. Smales LR. Acanthocephala in amphibians (anura) and reptiles (Squamata) from Brazil and Paraguay with description of a new species. J Parasitol. 2007;93(2):392-8.
11. Yamaguti S. Systema helminthum. The nematodes of vertebrates. v. III. parts I and II. New York: Interscience Publishers; 1961.
12. Yamaguti S. Systema helminthum. The cestodes of vertebrates. v. II. New York: Interscience Publishers; 1959.
13. Schmidt GD. CRC Handbook of tapeworm identification. Florida: CRC Press; 1986. 675 p.
14. Yamaguti S. Systema helminthum. The digenetic trematodes of vertebrates. V. I. New York: Interscience Publishers; 1958.
15. Yamaguti S. Systema helminthum. Acanthocephala. v. V. New York: Interscience Publishers; 1963.
16. Lamas MF, Zaracho VH. *Tropidurus torquatus*. Endoparasites. Natural History. Herpetol Rev. 2006;37:474-5.
17. Vrcibradic D, Rocha CFD, Van Sluys M, Bursey CR. *Mabuya macrorhyncha* (NCN) Endoparasites. Herpetol Rev. 2001; 32:256.
18. Bursey CR, Goldberg SR. *Cosmocerca vrcibradici* n. sp. (Ascaridida: Cosmocercidae), *Oswaldocruzia vittii* n. sp. (Strongylida: Molineidae), and other helminths from *Prionodactylus eigenmanni* and *Prionodactylus oshaughnessyi* (Sauria: Gymnophthalmidae) from Brazil and Ecuador. J Parasitol. 2004;90(1):140-5.
19. Vicente JJ. Helminths de *Tropidurus* (Lacertilia, Iguanidae) da Coleção Helminológica do Instituto Oswaldo Cruz I. Trematoda, Cestoda, Acanthocephala, Linguatulida. Atas Soc. Biol. Rio de Janeiro 1978;19:71-78.
20. Vrcibradic D, Rocha CFD. Observations on the natural history of the lizard *Mabuya macrorhyncha* Hoge (Scincidae) in Queimada Grande island, São Paulo. Rev Bras Zool. 2005;22(4):1185-90.
21. Rodrigues HO. Contribuição ao estudo da fauna helminológica de vertebrados de Nova Iguaçu, RJ. Atas Soc Biol. Rio de Janeiro. 1986;26:27-8.
22. Anjos LA, Rocha CFD, Vrcibradic D, Vicente JJ. Helminths of the exotic lizard *Hemidactylus mabouia* from a rock outcrop area in southeastern Brazil. J Helminthol. 2005;79(4):307-13.
23. Vrcibradic D, Rocha CFD, Bursey CR, Vicente JJ. Helminth communities of two sympatric skinks (*Mabuya agilis* and *Mabuya macrorhyncha*) from two 'restinga' habitats in southeastern Brazil. J Helminthol. 2002;76(4):355-61.
24. Vrcibradic D, Vicente JJ, Bursey CR. Helminths infecting the lizard *Enyalius bilineatus* (Iguanidae, Leiosaurinae), from an Atlantic Rainforest area in Espírito Santo state, southeastern Brazil. Amphib-Reptil. 2007;28:166-169.
25. Neiva A, Cunha AM, Travassos LP. Contribuições parasitológicas. Mem Inst Oswaldo Cruz. 1914;6(3):180-91.
26. Travassos L. Contribuições para o conhecimento da fauna helminthologica brasileira. XX Revisão dos Acanthocephalos brasileiros. Parte II. Familia Echinorhynchidae Hamann, 1892, sub-fam. Centrorhynchinae Travassos, 1919. Mem Inst Oswaldo Cruz. 1926;19(1):31-125.
27. Rodrigues HO. *Pseudocapillaria (Ichthyocapillaria) maricaensis* n. sp. (Nematoda, Capillariidae) and remarks on the helminthological fauna of *Liolaemus lutzae* Mertens, 1938 (Lacertilia, Iguanidae). Mem Inst Oswaldo Cruz. 1992; 87:297-300.
28. Baylis HA. Some Roundworms and Flatworms from the West Indies and Surinam. I. Nematodes and Acanthocephala. J Linn Soc London. 1947;41:394-405.
29. Lent H, Freitas JFT. Uma coleção de nematódeos de vertebrados, do Museu de Historia Natural de Montevideo. Mem Inst Oswaldo Cruz. 1948;46(1):1-71.
30. Sprent JFA. Ascaridoid nematodes of amphibians and reptiles: *Freitasascaris* n.g. J Helminthol. 1983;57:283-90.
31. Dias EJR, Vrcibradic D, Rocha CFD. Endoparasites infecting two species of whiptail lizard (*Cnemidophorus abaetensis* and *C. ocellifer*; Teiidae) in a restinga habitat of northeastern Brazil. Herpetol J. 2005;15(2):133-7.
32. Ribas SC, Rocha CFD, Teixeira-Filho PF, Vicente JJ. Nematode infection in two sympatric lizards (*Tropidurus torquatus* and *Ameiva ameiva*) with different foraging tactics. Amphib-Reptil. 1998;19(3):323-30.
33. Rocha CFD, Vrcibradic D. Nematode assemblages of some insular and continental lizard hosts of the genus *Mabuya* Fitzinger (Reptilia, Scincidae) along the eastern Brazilian coast. Rev Bras Zool. 2003;20(4):755-9.
34. Vrcibradic D, Rocha CFD, Ribas SC, Vicente JJ. Nematodes infecting the skink *Mabuya frenata* in Valinhos, São Paulo State, southeastern Brazil. Amphib-Reptil. 1999;20:333-9.
35. Vrcibradic D, Cunha-Barros M, Vicente JJ, Galdino CAC, Hatano FH, Van Sluys M, et al. Nematode infection patterns in four sympatric lizards from a restinga habitat (Jurubatiba) in Rio

- de Janeiro state, southeastern Brazil. *Amphib-Reptil.* 2000; 21(3):307-16.
36. Barus V, Coy Otero A. Systematic survey of nematodes parasitizing lizards (Sauria) in Cuba. *Helminthol.* 1969;10(3):329-46.
 37. Vicente JJ. Redescricao de *Atractis cruciata* Linstow, 1902, em novo hospedeiro (Nematoda, Atractoidea). *Atas Soc Biol Rio de Janeiro.* 1966;10(2):41-3.
 38. Bursey CR, Flanagan JP. *Atractis marquezii* n. sp. (Nematoda: Atractidae) and a revision of *Atractis* Dujardin, 1845, sensu Baker, 1987. *J Parasitol.* 2002;88:320-4.
 39. McAllister CT, Bursey CR, Freed PS. Helminth parasites of selected amphibians and reptiles from the Republic of Ecuador. *Comp Parasitol.* 2010;77(1):52-66.
 40. Goldberg SR, Bursey CR, Vitt LJ. Parasites of two lizard species, *Anolis punctatus* and *Anolis transversalis* (Squamata: Polychrotidae) from Brazil and Ecuador. *Amphib-Reptil.* 2006;27(4):575-9.
 41. Ávila RW, Souza FL, Silva RJ. Helminths from seven species of lizards (Reptilia: Squamata) at the Cerrado of Mato Grosso do Sul State, Brazil. *Comp Parasitol.* 2010;77(1):67-71.
 42. Adamson ML, Baccam D. Systematic position of the Atractidae sensu Chabaud (1978) (Nematoda: Cosmocercidae): *Maracaya belemensis* n. sp. and *Aplectana albae* n. sp. from *Amphisbaena alba* in Brazil. *Can J Zool.* 1988;66:1857-64.
 43. Boamer S, Morand S. A new nematode (Nematoda: Cosmocercidae) from the lizard, *Chamaeleo inturensis* (Squamata: Chamaeleonidae) from the Democratic Republic of Congo. *J Parasitol.* 2006;92(2):346-9.
 44. Ramallo GR, Bursey CR, Goldberg SR. A new species of Cosmocercidae (Ascaridida) in the worm lizard, *Amphisbaena bolivica* (Squamata: Amphisbaenidae), from Argentina. *J Parasitol.* 2008;94(6):1361-3.
 45. Goldberg SR, Bursey CR. *Cercosaura argulus* endoparasites. *Herpetol Rev.* 2007;38(4):451.
 46. Ramalho ACO, Silva RJ, Schwartz H, Péres AK. Helminths from an introduced species (*Tupinambis merianae*), and two endemic species (*Trachylepis atlantica* and *Amphisbaena ridleyi*) from Fernando de Noronha archipelago, Brazil. *J Parasitol.* 2009;95(4):1026-8.
 47. Travassos LP. *Aplecana raillietii*, n. sp. *C R Seanc Soc Biol.* 1925;93:973-4.
 48. Sousa BM, Lima SS, Oliveira A. Gastrointestinal helminth fauna of *Enyalius perditus* (Reptilia: Leiosauridae): Relation to host age and sex. *J Parasitol.* 2007; 93(1):211-3.
 49. Freitas JFT, Vicente JJ. Nôvo nematódeo do gênero *Cosmocerca* Diesing, 1861, parasito de anfisbaenideo. *Atas Soc Biol. Rio de Janeiro.* 1966;10:109-11.
 50. Bursey CR, Goldberg SR, Vitt LJ. New species of *Allopharynx* (Digenea: Plagiorchiidae) and other helminths in *Uranoscodon superciliosus* (Squamata: Tropicuridae) from Amazonian Brazil. *J Parasitol.* 2005;91(5):1395-8.
 51. Goldberg SR, Bursey CR, Vitt LJ. Helminths of the brown-eared anole, *Norops fuscoauratus* (Squamata, Polychrotidae) from Brazil and Ecuador, South America. *Phyllomedusa.* 2006;5(1):83-6.
 52. Goldberg SR, Bursey CR, Vitt LJ. Parasite communities of two lizard species, *Alopoglossus angulatus* and *Alopoglossus atriventris*, from Brazil and Ecuador. *Herpetol J.* 2007;17(4):269-72.
 53. Baker MR. *Dollfusnema amphisbaenia* n. gen., n. sp. (Nematoda: Cosmocercinae) from the lizard *Leposternon phocaena* (Amphisbaenia) of Brazil. *Can J Zool.* 1981;59(1):138-40.
 54. Baker MR. *Africana chabaudi* n. sp. (Nematoda, Heterakidae) in a Brazilian Iguana. *Bull Mus Nat His Nat Paris.* 1981;3(3):783-7.
 55. Ávila RW, Silva RJ. A new species of *Africana* (Nematoda: Heterakidae) from lizards of southern Amazon, Brazil. *J Parasitol.* 2009;95(5):1156-8.
 56. Freitas JFT. Novo parasito de réptil da Ilha Fernando de Noronha: *Moaciria alvarengai* g. n., sp. n. (Nematoda, Subuluroidea). *Rev Bras Biol.* 1956;16:335-9.
 57. Días-Ungría C. Notas sobre nematodos de reptiles en Venezuela. *Rev Vet Venez.* 1964;17(101):402.
 58. Rodrigues HO, Feijó LMF. Redescricao de *Spinicauda spinicauda* (Olfers, 1919) Travassos, 1920 (Nematoda, Oxyuroidea). *Atas Soc Biol Rio de Janeiro.* 1976; 18:59-63.
 59. Travassos LP. Informações sobre a fauna helminthologica de Mato Grosso. *Folha Med.* 1923;4:58-60.
 60. Pereira C. Os Oxyurata parasitos de Lacertilia do nordeste Brasileiro. *Arch Inst Biol.* 1935;6:5-27.
 61. Bursey CR, Goldberg SR, Telford Jr SR. *Strongyluris panamaensis* n. sp. (Nematoda: Heterakidae) and other helminths from the lizard, *Anolis biporcatus* (Sauria: Polychrotidae), from Panama. *J Parasitol.* 2003;89(1):118-23.
 62. Vicente JJ. Helminths of *Tropidurus* (Lacertilia, Iguanidae) da coleção helmintológica do Instituto Oswaldo Cruz. II. Nematoda. *Atas Soc Biol Rio de Janeiro.* 1981;22:7-18.
 63. Harvey MB, Gutberlet Jr RL. Lizards of the genus *Tropidurus* (Iguania: Tropicuridae) from the serrania de Huanchaca, Bolivia: New species, natural history, and a key to genus.

- Herpetologica.1998; 54:493-520.
64. Alho CJR. Oxyurata de lagartos do Planalto Central. Sobre o gênero *Strongyluris* Mueller, 1894 com descrição de duas espécies novas. Rev Bras Biol. 1969;29:65-74.
 65. Bursey CR, Goldberg SR. Helminths of *Tropidurus guarani* (Sauria: Tropiduridae) from Paraguay. Comp Parasitol. 2004;71(2):203-7.
 66. Fontes AF, Vicente JJ, Kiefer MC, Van Sluys M. Parasitism by helminths in *Eurolophosaurus nanuzae* (Lacertilia: Tropiduridae) in an area of rocky outcrops in Minas Gerais state, southeastern Brazil. J Herpetol. 2003;37(4):736-41.
 67. Kohn A, Pinto RM, Fernandes BMM. Contribuição ao conhecimento de *Strongyluris oscar* Travassos, 1923 (Nematoda, Subuluroidea). Mem Inst Oswaldo Cruz. 1973;71(3):219-25.
 68. Sutton CA, Mordeglia C, Cruz F. *Strongyluris oscar* Travassos, 1923 (Nematoda, Heterakidae) en *Tropidurus spinulosus* (Squamata, Tropiduridae) Del Noroeste Argentino. Gayana Zool. 1998;62:171-5.
 69. Roca V. *Tropidurus melanopleurus* (NCN) Parasites. Herpetol Rev. 1997; 28:204.
 70. Khalil M, Vogelsang EG. *Cruzia fulleborni*, a new species of Nematoda from *Tupinambis teguixin*. Zentralbl Bakteriol I Originale. 1930;119(1-2):72-4.
 71. Ruiz JM. Revisão do gênero *Cruzia* (Nematoda: Oxyuroidea) e estudo das espécies brasileiras [doctoral dissertation]. São Paulo (SP): Faculdade de Farmácia e Odontologia da USP; 1947. 105p.
 72. Schuurmans-Stekhoven JH. Nematodos parasitarios del chaco Paraguayo y de Argentina del Museo de Estocolmo. Acta Zool Lilloana.1950;9:325-45.
 73. Ávila RW, Cáceres NC, Ferreira VL, Silva RJ. *Hoplocercus spinosus*. Endoparasites. Herpetol Rev. 2008;39(1):88-9.
 74. Chabaud AG. *Skrjabinelazia Sypliaxov*, 1930 (= *Salobrella* Freitas, 1940), genre de transition entre Cosmocercidés et Spirurides. Ann Parasitol Hum Comp. 1973; 48:329-34.
 75. Chabaud AG, Bain O, Poinar GO. *Skrjabinelazia galliardi* (Nematoda, Seuratoidea): compléments morphologiques et cycle biologique. Ann Parasitol Hum Comp.1988;63(4):278-84.
 76. Freitas JFT. Sobre um interessante nematódeo parasito de reptil (Spiruroidea). Mem Inst Oswaldo Cruz.1940;35(3):603-5.
 77. Menezes VA, Vrcibradic D, Vicente JJ, Dutra GF, Rocha CFD. Helminths infecting the parthenogenetic whiptail lizard *Cnemidophorus nativo* in a restinga habitat of Bahia State, Brazil. J Helminthol. 2004;78(4):323-8.
 78. Vicente JJ, Van Sluys M, Fontes AF, Kiefer MC. *Subulura lacertilia* sp.n. (Nematoda, Subuluridae) parasitizing the Brazilian lizard *Tropidurus nanuzae* Rodrigues (Lacertilia, Tropiduridae). Rev Bras Zool. 2000;17(4):1065-8.
 79. Araújo P. Um novo Oxyurideo *Gynaecometra bahiensis* n. gen. n. sp. (Nematoda: Oxyuroidea: Oxyuridae) encontrado em lacertílio. Mem Inst Butantan. 1976;40/41:251-7.
 80. Tantaléan M. Nuevos registros de nemátodes parasitos de animales de vida silvestre en el Peru. Rev Peru Biol.1998;5(2):103-4.
 81. Lopes SG, Silva LEM, Dantas EF, Almeida WO. Infecção por helmintos em três espécies de lagartos do Nordeste Brasileiro. Cad Cult Ciênc. 2006;1(1):47-51.
 82. Arrojo L. Parásitos de animales silvestres en cautiverio en Lima, Perú. Rev Peru Biol. 2002;9(2):118-20.
 83. Inglis WG, Diaz-Ungría C, Coles WJ. Nematodes de Venezuela. IV. Nematodes parásitos de vertebrados venezolanos, II. Acta Biol Venez. 1960;3:1-24.
 84. Freitas JFT. Sobre os generos *Thelandros* Wedl, 1962 e *Parapharyngodon* Chatteuji, 1933, com descrição de *Parapharyngodon alvarengai* sp. n. (Nematoda, Oxyuroidea). Mem Inst Oswaldo Cruz. 1957;55(1):21-45.
 85. Padilha TN, Duarte MJF. Ocorrência de *Parapharyngodon alvarengai* Freitas, 1957, em *Ameiva ameiva* (L.) no estado do Rio de Janeiro (Nematoda, Oxyuroidea). Atas Soc. Biol. Rio de Janeiro. 1979; 20:21-22.
 86. Ramallo GR, Bursey CR, Goldberg SR. *Parapharyngodon riojensis* n. sp. (Nematoda: Pharyngodonidae) from the lizard *Phymaturus punae* (Squamata: Iguania: Liolaemidae) from northwestern Argentina. J Parasitol. 2002;88(5):979-82.
 87. Bursey CR, Brooks DR. *Parapharyngodon duniae* n. sp. (Nematoda: Pharyngodonidae) in *Phrynohyas venulosa* (Anura: Hylidae) from the Area de Conservación Guanacaste, Guanacaste, Costa Rica. J Parasitol. 2004;90(1):137-9.
 88. Bursey CR, Goldberg SR. Two new species of Pharyngodonidae (Nematoda: Oxyuroidea) and other Nematodes in *Agama caudospina* (Squamata: Agamidae) from Kenya, Africa. J Parasitol. 2005;91(3):591-9.
 89. Calisaya JL, Cordova E. Tres nuevas especies de *Parapharyngodon* (Nematoda, Oxiuroidea) parásitas de *Tropidurus peruvianus* del sur del Perú. Rebiol. 1997;17(1-4):45-54.
 90. Morales E, Sarmiento L, Sánchez L, Florindéz D, Lamas G. Material tipo de helmintos en el Museo de Historia Natural, Universidad Nacional Mayor de San Marcos, (MUSM), Lima, Perú. Rev Peru

- Biología. 2005;12(3):463-72.
91. Rodrigues HO, Pinto RM. Ocorrência de dois faringodonídeos em *Ameiva ameiva* (L.) no estado da Guanabara (Nematoda, Oxyuroidea). Atas Soc Biol Rio de Janeiro. 1967;11(3):123-5.
 92. Vásquez PTS. Helminths parasites of *Dicrodon guttulatum*, Dumeril y Bibron, 1893 (SAURIA: TEIIDAE) de la costa del Perú [dissertation]. Lima, Peru: Facultad Ciencias Biológicas, Universidad Nacional Mayor de San Marcos; 2008. 63p.
 93. Alho CJR, Rodrigues HO. Sobre novo hospedeiro de *Parapharyngodon sceleratus* (Travassos, 1923) Freitas, 1957, com redescoberta da espécie (Nematoda, Oxyuroidea). Atas Soc Biol Rio de Janeiro. 1963; 7:4-6.
 94. Ribas SC, Rocha CFD, Teixeira-Filho PF, Vicente JJ. Helminths (Nematoda) of the lizard *Cnemidophorus ocellifer* (Sauria: Teiidae): assessing the effect of rainfall, body size and sex in the nematode infection rates. Ciênc Cult. 1995;47(1-2):88-91.
 95. Van Sluys M, Rocha CFD, Ribas SC. Nematodes infecting the lizard *Tropidurus itambere* in southeastern Brazil. Amphib-Reptilia. 1994;15(4):405-8.
 96. Van Sluys M, Rocha CFD, Bergallo HG, Vrcibradic D, Ribas SC. Nematode infection in three sympatric lizards in an isolated fragment of restinga habitat in southeastern Brazil. Amphib-Reptil. 1997;18(4):442-6.
 97. Goldberg SR, Bursey CR. Helminths from four species of *Phyllodactylus* (Squamata: Gekkonidae) from Peru. Comp Parasitol. 2010;77(1):91-3.
 98. Freitas JFT. Sobre um novo nematódeo de réptil da Bolívia: *Parapharyngodon senisfaciecaudatus* sp. n. (Nematoda, Oxyuroidea). Rev Bras Biol. 1957;17:451-4.
 99. Freitas JFT, Dobbin Jr, JE. Nova espécie do gênero *Parapharyngodon* Chatterji, 1933 (Nematoda, Oxyuroidea). An Soc Biol Pernambuco. 1959;16:23-33.
 100. Rodrigues HO, Rodrigues SS, Faria Z. Contribution to the knowledge of the helminthological fauna of vertebrates of Maricá, Rio de Janeiro State, Brazil. Mem Inst Oswaldo Cruz. 1990;85(1):115-6.
 101. Zaracho VH, Lamas MF. *Tropidurus etheridgei*. Endoparasites. Natural History. Herpetol Rev. 2006; 37:473.
 102. Cruz FB, Silva S, Scrocchi GJ. Ecology of the lizard *Tropidurus etheridgei* (Squamata: Tropiduridae) from the dry Chaco of Salta, Argentina. Herpetol Nat Hist. 1998;6(1):23-31.
 103. Rocha CFD. Nematode parasites of the Brazilian sand lizard, *Liolaemus lutzae*. Amphib-Reptil. 1995;16(4):412-5.
 104. Alho CJR, Moura VP. Oxyurata de lagartos do Planalto Central. Ocorrência de *Pharyngodon cesarpinto* Pereira, 1935 (Nematoda, Subuluroidea). Atas Soc Biol Rio de Janeiro. 1970;13(1-2):67-9.
 105. Freitas JFT, Ibáñez N. Fauna helmintológica del Perú – Notas sobre “Pharyngodonidae” Travassos, 1920 y descripción de *Pharyngodon micrurus* sp. n. Rev. Universitaria 1963; 23 y 24.
 106. Bursey CR, Goldberg SR. New species of *Skrjabinodon* (Nematoda: Pharyngodonidae) in *Uracentron flaviceps* (Squamata: Iguanidae) from Ecuador and Peru. J Parasitol. 2007;93(4):866-9.
 107. Vicente JJ, Vrcibradic D, Muniz-Pereira LC, Pinto PM. *Skrjabinodon heliocostai* sp. n. (Nematoda, Pharyngodonidae) parasitizing *Mabouya frenata* (Cope) (Lacertilia, Scincidae) in Brazil and the reallocation of *Skrjabinodon capacypanquii* (Freitas, Vicente & Ibanez) in the genus *Thelandros* Wedl. Rev Bras Zool. 2000; 17:361-367.
 108. Rocha CFD, Vrcibradic D, Vicente JJ, Cunha-Barros M. Helminths infecting *Mabouya dorsivittata* (Lacertilia, Scincidae) from a high-altitude habitat in Itatiaia National Park, Rio de Janeiro State, southeastern Brazil. Braz J Biol. 2003;63(1):129-32.
 109. Vicente JJ, Vrcibradic D, Rocha CFD, Pinto RM. Description of *Skrjabinodon spinosulus* sp. n. (Nematoda, Oxyuroidea, Pharyngodonidae) from the Brazilian lizard *Mabouya dorsivittata* Cope, 1862 (Scincidae). Rev Bras Zool. 2002;19(1):157-62.
 110. Ramallo GR, Bursey CR, Goldberg SR. *Spauligodon lobo* n. sp. (Nematoda: Pharyngodonidae) parasite of *Liolaemus* spp. (Iguania: Liolaemidae) from northwestern Argentina. J Parasitol. 2002;88(4):370-4.
 111. Vicente JJ, Ibáñez HN. Nova espécie do gênero *Parathelandros* Baylis, 1930 (Nematoda, Oxyuroidea). Atas Soc Biol Rio de Janeiro. 1968;11:185-7.
 112. Goldberg SR, Muñoz G, Garcias F, Bursey CR. *Liolaemus lemniscatus* (wreath tree iguana), *Liolaemus tenuis* (thin tree iguana). Endoparasites. Herpetol Rev. 2001;32:41.
 113. Bergmann PJ, Russell AP. Systematics and biogeography of the widespread Neotropical gekkonid genus *Thecadactylus* (Squamata), with the description of a new cryptic species. Zool J Linnean Soc. 2007;149:339-70.
 114. Goldberg SR, Bursey CR. *Phyllodactylus reissi* Endoparasites. Herpetol Rev. 2004; 35:395.
 115. Ávila RW, Anjos LA, Gonçalves U, Freire EMX, Almeida WO, Silva RJ. Nematode infection in the lizard *Bogertia lutzae* (Loveridge, 1941)

- from the Atlantic forest in northeastern Brazil. *J Helminthol.* 2010; 89(2):199-201.
116. Freitas JFT, Vicente JJ, Ibáñez N. Fauna helmintológica do Peru: Novo nematódeo do gênero *Parathelandros* Baylis, 1930 (Nematoda, Oxyuroidea). *Atas Soc Biol Rio de Janeiro.* 1968;12:33-35.
 117. Pérez ZJ, Balta K, Salizar P, Sánchez L. Nematofauna de tres especies de lagartijas (Sauria: Tropicuridae y Gekkonidae) de la Reserva Nacional de Paracas, Ica, Perú. *Rev Peru Biol.* 2007;14(1):43-5.
 118. Freitas JFT, Vicente JJ, Ibáñez N. Fauna Helmintológica del Perú: *Parathelandros capacyupanquii* sp. n., parasito de *Dicrodon holmbergi* Schmidt, 1957 (Nematoda, Oxyuroidea). *Atas Soc Biol Rio de Janeiro.* 1968;11:217-9.
 119. Lhermitte-Vallarino L, Bain O, Deharo E, Bertani S, Voza T, Attout T, et al. A new rhabdiasid nematode, *Chabirenia cayennensis* n. g., n. sp., parasitic in the glands of the buccal mucosa of a South American saurian. *Syst Parasitol.* 2005;62(2):151-60.
 120. Muniz-Pereira LC, Vieira FM, Luque JL. Checklist of helminth parasites of threatened vertebrate species from Brazil. *Zootaxa.* 2009;2123:1-45.
 121. Almeida WO, Ribeiro SC, Santana GG, Vieira WLS, Anjos LA, Sales DL. Lung infection rates in two sympatric Tropicuridae lizard species by pentastomids and nematodes in northeastern Brazil. *Braz J Biol.* 2009;69(3):963-7.
 122. Rodrigues HO. Sobre nova espécie do gênero *Strongyloides*. *Atas Soc Biol Rio de Janeiro.* 1968;12:31-32.
 123. Freitas JFT, Rodrigues HO. Nótula helmintológica: sobre o material original de *Filaria multipapilla* Molin, 1858 atualmente existente no Museu de Viena. *Atas Soc Biol Rio de Janeiro.* 1964;8:37-8.
 124. Lent H, Freitas JFT. Sobre os filarídeos parasitas de lacertídeos neotropicos. *Rev Bras Biol.* 1941;1(4):383-6.
 125. Vicente JJ, Jardim CR. Filarídeos da Coleção helmintológica do Instituto Oswaldo Cruz. I. Peixes, anfíbios e répteis. *Atas Soc Biol Rio de Janeiro.* 1980; 21:47-57.
 126. McAllister CT, Bursey CR, Freed PS. Helminth parasites (Cestoda: Nematoda) of selected herpetofauna from Paraguay. *J Parasitol.* 2010;96(1):222-4.
 127. Bain O. Description de nouvelles filaires Oswaldofilariinae de lézards sud-américains; hypothese sur l'évolution des filaires de reptiles. *Bull Mus natn Hist nat Paris, Zool.* 1974;138(108):169-200.
 128. Freitas JFT, Lent H. Sobre *Oswaldofilaria brevicaudata* (Rhodian and Vuylssteke, 1937) n. comb. (Nematoda; Filarioidea). *Mem Inst Oswaldo Cruz.* 1937;32:439-42.
 129. Díaz-Ungria C. Helminths parásitos de vertebrados en el estado Zulia (Venezuela) algunas especies nuevas para Venezuela. *Vet Trop.* 1978;3:15-37.
 130. Silva RJ, Kohlsdorf T. *Tropicurus hispidus* Spix 1825 (Sauria, Tropicuridae): a new host for *Oswaldofilaria petersi* Bain & Sulahian 1974 (Nematoda, Onchocercidae). *Arq Bras Med Vet Zootec.* 2003;55(3):377-9.
 131. Ávila RW, Silva RJ. Helminths of the teiid lizard *Kentropyx calcarata* (Squamata) from an Amazonian site in western Brazil. *J Helminthol.* 2009;83(3):267-9.
 132. Ramallo GR, Díaz F. *Physaloptera lutzi* (Nematoda, Physalopteridae) parasite de *Liolaemus* (Iguania, Tropicuridae) del noroeste Argentino. *Bol Chil Parasitol.* 1998;53(1-2):19-22.
 133. Cristofaro R, Guimarães JF, Rodrigues HO. Alguns nematódeos de *Tropicurus torquatus* (Wied) e *Ameiva ameiva* (L.)—Fauna Helmintológica de Salvador, Bahia. *Atas Soc Biol Rio de Janeiro.* 1976;18:65-70.
 134. Díaz-Ungria C, Gallardo MF. Nematodes de reptiles Venezolanos, con descripción de varias especies nuevas. *Bol Soc Vem Cienc Nat.* 1968; 27(113-114):550-70.
 135. Vicente JJ, Santos E. Ocorrência de *Physaloptera retusa* Rudolphi, 1819 em novo hospedeiro (Nematoda, Physalopteridae). *Atas Soc Biol Rio de Janeiro.* 1967; 11:75-7.
 136. Goldberg SR, Bursey CR, Caldwell JP, Vitt LJ, Costa GC. Gastrointestinal Helminths from six species of frogs and three species of lizards, sympatric in Pará State, Brazil. *Comp Parasitol.* 2007;74:327-42.
 137. Goldberg SR, Bursey CR. *Uracentron flaviceps*. Endoparasites. *Herpetol Rev.* 2007;38(1):85.
 138. Noronha D, Bragança R, Vicente JJ, Pereira LCM. Coleções particulares incorporadas à coleção helmintológica do Instituto Oswaldo Cruz (CHIOC). I: Coleção do Instituto Pasteur de São Paulo. *Rev Bras Zool.* 2004;21(2):303-5.
 139. Goldberg SR, Bursey CR. *Ameiva festiva* Endoparasites. *Herpetol Rev.* 2009;40(1):81-2.
 140. Prieto AS. Note on parasites of the tropical lizard *Tropicurus hispidus*. *J Herpetol.* 1980;14(2):190-2.
 141. Caballero E, Vogelsang EG. Fauna helmintológica venezolana. I. *Ochetosoma miladelarocai* n. sp. de *Bothrops atrox* L. y hallazgo de *Physaloptera retusa* (Rud., 1819) en *Cnemidophorus lemniscatus lemniscatus*. *Rev Med Vet Parasitol.* 1947;6:53-62.
 142. Ávila RW, Anjos LA, Silva RJ, Costa HC, São Pedro VA, Feio RN. *Tupinambis longilineus*. Endoparasites. *Herpetol Rev.* 2010;41(1):87-8.
 143. Silva AS, Zanette RA, Tochetto C, Oliveira

- CB, Soares JB, Otto MA, et al. Parasitismo por *Physaloptera* sp., *Kalicephalus* sp. e *Cryptosporidium* sp. em lagarto (*Tupinambis teguixin*) no Rio Grande do Sul, Brasil. *Zoociências*. 2008;10(3):269-72.
144. de Fabio SP, Rolas FJT. Contribuição ao conhecimento de *Thubunaea dactyluris* Karve, 1938 (Nematoda, Spiruroidea). *Mem Inst Oswaldo Cruz*. 1974; 72(3-4):283-290.
145. Baylis HA. On a new Species of the Nematode Genus *Thubunaea*. *Ann Mag Nat Hist*. 1926;9(18):361-4.
146. Rodrigues HO. Estudo da fauna helmintológica de *Hemidactylus mabouia* (M. de J.) no Estado da Guanabara. *Atas Soc Biol Rio de Janeiro*. 1970;12:15-23.
147. Travassos LP, Freitas JFT. Relatório da terceira excursão a zona da estrada de Ferro Noroeste do Brasil realizada em fevereiro e março de 1940. *Mem Inst Oswaldo Cruz*. 1941;35(3):607-96.
148. Anderson RC. (2000) Nematode Parasites of Vertebrates. Their Development and Transmission. Wallingford, Oxon, U.K.: CAB International; 2000. 650 pp.
149. Freitas JFT, Lent H. Pesquisas helmintológicas realizadas no estado do Pará. V. gênero *Diaphanocephalus* Diesing, 1851 (Nematoda: Strongyloidea). *Mem Inst Oswaldo Cruz*. 1938;33(3):423-32.
150. Spinelli CM, Fiorito LE, Stiebel C. Alteraciones histológicas en el intestino delgado en *Tupinambis rufescens* (Sauria, Teiidae) causadas por *Diaphanocephalus galeatus* (Nematoda, Diaphanocephalidae). *Cuad Herpetol*. 1992;7(6):38-40.
151. Alho CJR. Sobre uma espécie nova do gênero *Diaphanocephalus* Diesing, 1851, parasita de lagarto da Amazônia (*Dracaena guianensis* Daudin) (Nematoda, Strongyloidea). *Bol Mus Par Emilio Goeldi Zool*. 1965;59:1-5.
152. Travassos LP, Freitas JFT, Mendonça JM. Relatório da excursão do Instituto Oswaldo Cruz ao Parque de Reserva e Refúgio Sooretama, no estado do Espírito Santo, em outubro de 1963. *Bol Mus Biol Mello-Leitão*. 1964;23(1):1-26.
153. Baker MR. On two new nematode parasites (Trichostrongyloidea: Molineidae) from amphibians and reptiles. *Proc Helm Soc Wash*. 1982;49:252-7.
154. Ben Slimane B, Durette-Desset MC. Four new species of *Oswaldocruzia* (Nematoda: Trichostrongylinea, Molineoidea) parasitizing amphibians and lizards from Ecuador. *Mem Inst Oswaldo Cruz*. 1996;91(3):317-28.
155. Durette-Desset MC, Anjos LA, Vrcibradic D. Three new species of the genus *Oswaldocruzia* Travassos, 1917 (Nematoda, Trichostrongylinea, Molineoidea) parasites of *Enyalius* spp. (Iguanidae) from Brazil. *Parasite*. 2006;13(2):115-25.
156. Lent H, Freitas JFT. Sobre uma nova espécie do gênero *Oswaldocruzia* Travassos, 1917. *Mem Inst Oswaldo Cruz*. 1935;30(3):379-86.
157. Ben Slimane B, Verhaagh M, Durette-Desset MC. *Oswaldocruzia peruensis* n. sp. (Nematoda: Trichostrongylinea) parasite d'un Iguanidae du Pérou. *Bull Mus Nat Hist Nat Paris*. 1995;17:77-82.
158. Freitas JFT. Nota sobre a fauna helmintológica de répteis brasileiros. *Rev Bras Biol*. 1955;15(3):279-84.
159. Vicente JJ, Santos E. Contribuição ao conhecimento de *Oochoristica fuhrmanni* Hughes, 1940. *Atas Soc Biol Rio de Janeiro*. 1971;15:29-31.
160. Rego AA. Contribuição ao conhecimento dos Cestóides do Brasil. I-Cestóides de peixes, anfíbios e répteis. *Atas Soc Biol Rio de Janeiro*. 1973;16(2):97-129.
161. Pinto RM, Gomes DC. Contribuição ao conhecimento da fauna helmintológica da região Amazônica - Cestódeos. *Mem Inst Oswaldo Cruz*. 1976; 74(1):53-64.
162. Rego AA, Rodrigues HO. Sobre duas *Oochoristica* parasitas de lacertílios (Cestoda, Cyclophyllidae). *Rev Bras Biol*. 1965; 25(1):59-65.
163. Bursley CR, Goldberg SR. *Oochoristica maccoyi* n. sp. (Cestoda: Linstowiidae) from *Anolis gingivinus* (Sauria: Polychrotidae) collected in Anguilla, Lesser Antilles. *Caribbean J Sci*. 1996;32(4):390-4.
164. Arizmendi-Espinosa MA, García-Prieto L, Guillén-Hernández S. A new species of *Oochoristica* (Eucestoda: Cyclophyllidae) parasite of *Ctenosaura pectinata* (Reptilia: Iguanidae) from Oaxaca, Mexico. *J Parasitol*. 2005; 91(1):99-101.
165. Bursley CR, Goldberg SR, Telford Jr SR. Gastrointestinal helminths of 14 species of lizards from Panama with descriptions of five new species. *Comp Parasitol*. 2007;74(4):108-40.
166. Guillén-Hernández S, Garcia-Prieto L, Arizmendi-Espinosa MA. A new species of *Oochoristica* (Eucestoda: Cyclophyllidae) parasite of *Ctenosaura oaxacana* (Reptilia: Iguanidae) from México. *J Parasitol*. 2007;93(5):1136-9.
167. Rego AA, Ibañez HN. Duas novas espécies de *Oochoristica*, parasitas de lagartixas do Peru (Cestoda, Anoplocephalidae). *Mem Inst Oswaldo Cruz*. 1965;63:67-73.
168. Lopez-Neyra CR, Diaz-Ungria C. Cestodes de Venezuela.— III. Sobre unos cestodes intestinales

- de reptiles y mamíferos venezolanos. Mem Soc Cienc Nat. La Salle. 1957;17:28-63.
169. Rego AA. Sobre alguns cestódeos parasitos de Répteis. Rev Bras Biol. 1967;27(2):181-7.
170. Coquille SC, De Chambrier A. *Cairaella henrii* n. gen., n. sp., a parasite of *Norops trachyderma* (Polychrotidae) and *Ophiotaenia nicolae* n. sp. (Eucestoda: Proteocephalidea), a parasite of *Thecadactylus rapicauda* (Gekkonidae). Folia Parasitol. 2008; 55:197-206.
171. Rego AA, Chambrier A. Redescription of *Tejidotaenia appendiculata* (Baylis, 1947) (Cestoda: Proteocephalidea), a parasite of *Tupinambis teguixin* (Sauria: Teiidae) from South America. Mem Inst Oswaldo Cruz. 2000; 95(2):161-5.
172. Hughes RC, Higginbotham JW, Clary JW. The trematodes of reptiles-part 1. Am Midl Nat. 1942;27:109-34.
173. Freitas JFT, Lent H. Sobre um novo trematódeo parasita de *Iguana tuberculata* (Laur.). Mem Inst Oswaldo Cruz. 1937;32(1):535-8.
174. Travassos LP. Informações sobre a fauna helmintológica de Mato Grosso. Fol Med. 1922;3:187-190.
175. Travassos LP. Fauna helminthologica de Mato Grosso (Trematódeos – 1ª Parte). Mem Inst Oswaldo Cruz. 1928; 21(2):309-41.
176. Travassos LP. Contribuição para a sistemática dos Dicrocoelinae Looss, 1899. Arc Esc Sup Agric Med Vet. 1919; 3:7-24.
177. Travassos LP. Relatório da excursão do Instituto Oswaldo Cruz ao município de Santa Teresa, no estado do Espírito Santo, em Agosto e Setembro de 1943. Mem Inst Oswaldo Cruz. 1944; 40(2):121-8.
178. Nasir P, Díaz MT. Flukes from Venezuelan reptiles with observations on intraspecific variations. Riv Parassitol. 1971;32(4):231-48.
179. Rodrigues HO. *Plagiorchis vicentei* n. sp. (Trematoda, Plagiorchiidae) a new trematode from *Hemidactylus mabouia* (Moreau de Jonnes) (Lacertilia, Gekkonidae). Rev Bras Zool. 1994;11(4):669-72.
180. Muzzall PM. Parasites of amphibians and reptiles from Michigan: A review of the literature 1916–2003. Michigan Dept. Nat. Res., Fisheries Research Report 2005; 2077:1-30.
181. Rodríguez-Ortiz B, García-Prieto L, Pérez-Ponce de León B. Checklist of the helminth parasites of vertebrates in Costa Rica. Rev Biol Trop. 2004; 52(2):343-54.
182. Hugot JP, Baujard P, Morand S. Biodiversity in helminths and nematodes as a field of study: an overview. Nematol. 2001;3(3):199-208.
183. Gregory RD, Keymer AE, Harvey PH. Helminth parasite richness among vertebrates. Biodiv Conserv. 1996;5(3):985-97.
184. Pough FH, Andrews RM, Cadle JE, Crump ML, Savitzky AH, Wells KD. Herpetology. Second edition. New Jersey: Prentice-Hall; 2001. 612p.
185. Pianka ER, Vitt LJ. Lizards: Windows to the evolution of diversity. Berkeley: University of California Press; 2003. 333pp.
186. Lavilla E, Richard E, Scrocchi G. Categorización de los anfibios y reptiles de la república Argentina. San Miguel de Tucumán: Asociación Herpetológica Argentina; 2000. 97 p.
187. Bérnils RS. Brazilian reptiles – List of species [Internet]. Curitiba: Sociedade Brasileira de Herpetologia; c2004-2010 [updated 2010 March; cited 2010 April 6]. Available from: <http://www.sbherpetologia.org.br/>.