

Neotropical Acanthoderini (Coleoptera, Cerambycidae, Lamiinae): Synonymies and new status in some genera, new species, transferences and new distributional records

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Abstract. Taxonomic notes are provided in some Acanthoderini genera. Three new species are described: *Scythropopsis intricata* Santos-Silva, Botero and Wappes from Mexico, *Aegomorphus robustus* Santos-Silva, Botero and Wappes and *Eupromerella boliviana* Santos-Silva, Botero and Wappes from Bolivia. The following synonymies are proposed: *Psapharochrus* Thomson, 1864 as synonym of *Aegomorphus* Haldeman, 1847; *Acanthoderes* (*Psapharochrus*) *albomaculatus* Fuchs, 1963 and *Acanthoderes griseomaculata* Zajciw, 1971 as synonyms of *Symperasmus alboniger* (Bates, 1861); *Pteridotelus contaminatus* Thomson, 1865 as synonym of *Scythropopsis melanostictica* (White, 1855); *Psapharochrus jameswappesi* Tavakilian, 2018 as synonym of *Aegomorphus wappesi* (Galileo, Martins & Santos-Silva, 2015); and *Psapharochrus histrio* Casey, 1913 and *Psapharochrus guatemalensis* Casey, 1913 as synonyms of *Aegomorphus circumflexus* (Jacquelin du Val, 1857). *Acanthoderes circumflexus* Jacquelin DuVal, 1857 is proposed as *nomen protectum* and *Acanthocinus rusticus* Klug, 1829 as *nomen oblitum*. *Aegomorphus contaminatus* (Thomson, 1965) is revalidated. *Scythropopsis* Thomson, 1864 and *Symperasmus* Thomson, 1864 are herein elevated to genus rank. The geographical distribution of ten species is expanded and a key to differentiate *Acanthoderes*, *Aegomorphus*, *Scythropopsis*, and *Symperasmus* is provided.

Key-Words. *Eupromerella*; *Mirochrus*; *Plagiosaurus*; *Sychnomerus*; Taxonomy.

INTRODUCTION

The tribe Acanthoderini has a cosmopolitan distribution and is comprised of 66 genera and more than 560 species making it one of the largest tribes in the Lamiinae (Tavakilian & Chevillotte, 2019). Most Acanthoderini genera and species are geographically restricted to the Neotropical Region. In this region there is a group of closely related genera whose taxonomy is confusing, which has resulted in species allocated to them that do not fit the current definition of the genera further complicating an already confusing situation.

Herein, the definition and limits of these closely related genera: *Acanthoderes* Audinet-Serville, 1835, which is subdivided in four subgenera, *A.* (*Acanthoderes*), *A.* (*Pardalisia*) Casey, 1913, *A.* (*Scythropopsis*) Thomson, 1864, and *A.* (*Symperasmus*) Thomson, 1864; *Aegomorphus*

Haldeman, 1847; *Eupromerella* Fisher, 1938; *Mirochrus* Galileo & Martins, 2012; *Plagiosaurus* Bates, 1880; *Psapharochrus* Thomson, 1864; and *Sychnomerus* Bates, 1885 are defined to validate each. As a result, numerous taxonomic changes are proposed. Additionally, three new species are described, and new geographical records listed for ten species.

MATERIAL AND METHODS

Photographs were taken in the MZSP with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65 mm f/2.8 1-5X macro lens, controlled by Zerene Stacker AutoMontage software. Measurements were taken in "mm" using a measuring ocular Hensoldt/Wetzlar – Mess 10 in the Leica MZ6 stereomicroscope, also used in the study of the specimens.

Pap. Avulsos Zool., 2020; v.60: e20206006

<http://doi.org/10.11606/1807-0205/2020.60.06>

<http://www.revistas.usp.br/paz>

<http://www.scielo.br/paz>

Edited by: Simone Policena Rosa

Received: 29/08/2019

Accepted: 16/12/2019

Published: 31/01/2020

ISSN On-Line: 1807-0205

ISSN Printed: 0031-1049

ISNI: 0000-0004-0384-1825

<http://zoobank.org/D696AE0B-C11E-4A5A-B42B-2A841532E842>



The acronyms used in the text are as follows:

- ACMT:** American Coleoptera Museum (James Wappes), San Antonio, Texas, USA
FSCA: Florida State Collection of Arthropods, Gainesville, FL, USA
FWSC: Fred W. Skillman collection, Pearce, Arizona, USA
MNKM: Museo de História Natural Noel Kempff Mercado, Santa Cruz, Santa Cruz, Bolivia
MNRJ: Museu Nacional do Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil
MZSP: Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil
RFMC: Roy F. Morris Collection, Lakeland, FL, USA
SWLC: Steven W. Lingafelter Collection, Hereford, AZ, USA

TAXONOMY

**On *Acanthoderes* Audinet-Serville, 1835,
Aegomorphus Haldeman, 1847,
Psapharochrus Thomson, 1864,
Scythropopsis Thomson, 1864,
Symperasmus Thomson, 1864, and
Sychnomerus Bates, 1885**

The definitions of the above acanthoderine genera have seen numerous changes since their original descriptions. These changes have resulted in modifications in their status (some more than once) making for a complicated history. In the most part their original descriptions have not allowed taxonomists to clearly understand their limits, and as a result new species have been assigned to them that changed their limits. Thus, making the study of this group of genera even more complicated and difficult. Santos-Silva & Nascimento (2018) commented about the differences between *Acanthoderes* and *Psapharochrus*, and reported: "Apart from those controversial classifications, it was Lacordaire (1872) who provided the best feature to separate *Psapharochrus* (including *Symperasmus* as a synonym) from *Acanthoderes* (*Acanthoderes*) and *A. (Scythropopsis)*. The former was included among the genera with eyes coarsely or sub-coarsely granulated, while the two latter genera were included among those with eyes finely granulated." Lacordaire (1872) also included *Aegomorphus* in the group of genera with eyes coarsely granulated, and separated it from *Psapharochrus* in the key by the genal length (translated): "Genae very short, sometimes nearly absent", leading to *Aegomorphus*; and "Genae, at least, with medium size", leading to *Psapharochrus*. However, the genae in the type species of *Aegomorphus*, *Aegomorphus decipiens* Haldeman, 1847 (Figs. 4-6) (= *Lamia modesta* Gyllenhal, 1817 (Figs. 41) = *Aegomorphus modestus*), is distinctly long, as in the type species of *Psapharochrus*, *Acanthoderes cylindricus* Bates, 1861 (Figs. 7-9). Actually, the genal length is longer in the type species of the former than in the latter, and *Aegoschema sensu* Lacordaire (1872) is equal to *Aegoschema* Aurivillius, 1923. This becomes clear when seeing the species originally includ-

ed in *Aegomorphus* by Lacordaire (1872): *A. adspersus* Thomson, 1861; *A. moniliferus* White, 1855; and *A. obesus* Bates, 1861. But it is curious to note that *Aegomorphus decipiens* Haldeman, 1847, type species of the genus, was included in *Psapharochrus* by Lacordaire (1872). The problem here was that Lacordaire (1872) wrongly attributed the authorship of the *Aegomorphus* to Thomson (1861), considering *Aegomorphus* Haldeman as a synonym of *Psapharochrus*. Evidently, thus occurred because Haldeman (1847) attributed the authorship of *Aegomorphus* to Dejean (1835) followed by a correction of this by Aurivillius (1923): "*Aegoschema* n. nom. / *Aegomorphus* Thoms. (nec Hald. 1847)."

Santos-Silva & Nascimento (2018) summarized their conclusions as follows: "1. *Symperasmus* – probably a synonym of *Psapharochrus*, but, at least, it must be considered a subgenus of *Psapharochrus*. 2. *Pardalisia* – probably a synonym of *Acanthoderes*; 3. *Scythropopsis* – probably a genus distinct from *Acanthoderes*; 4. Eyes very finely granulated – *Acanthoderes (Acanthoderes)*; *A. (Pardalisia)*; *A. (Scythropopsis)*. 5. Eyes coarsely or moderately coarsely granulated – *Psapharochrus*; *A. (Symperasmus)*." However, at that time they did not formally propose any change in the status of those genera.

A study of the type species of *Aegoschema* and *Psapharochrus* has revealed that they share the same features and lack significant differences hence we conclude that they are not different genera. Accordingly, we formally synonymise *Psapharochrus* with *Aegomorphus*. Although, it is important to note that several species currently placed in *Psapharochrus* which have the lateral tubercles of the prothorax with the apex blunt (not acute as in the type species) and the elytra gradually narrowed from humerus to apex (not parallel-sided as in the type species), indicates that they are likely not true *Psapharochrus* and thus, not *Aegomorphus* either.

Following the conclusions of Santos-Silva & Nascimento (2018), we also consider both *Scythropopsis* and *Symperasmus* to be genera distinct from *Acanthoderes*. However, *Pardalisia* Casey, 1913 will provisionally be kept as a subgenus of *Acanthoderes*.

The following key incorporates the primary characters needed to separate the genera involved in this study:

1. Eyes very finely granulated (Figs. 1-3, 10-12) 2
— Eyes slightly to distinctly coarsely granulated (Figs. 4-6, 7-9, 13-15) . 3
- 2(1). Elytra distinctly carinate dorsally (Fig. 10)..... *Scythropopsis*
— Elytra without carina distant from suture (Fig. 1)
..... *Acanthoderes (Acanthoderes)* and *A. (Pardalisia)*
- 3(1). Pronotum (Figs. 13-14) with distinct central depression, from near base to near apex or only in anterior region, margined by lateral elevation (carinate or not) *Symperasmus*
— Pronotum (Figs. 4-5, 7-8) not distinctly depressed centrally, nor with lateral tubercles forming carina..... *Aegomorphus*

It is important to note that this study does not exclude the possibility that some species currently included (or transferred herein) in *Acanthoderes*, *Aegomorphus*, *Symperasmus*, and *Scythropopsis* may need a new genus. This is especially true regarding some species current-

ly included in *Psapharochrus*, which differ considerably from the type species of the genus, and hence also *Aegomorphus*.

The species of *Aegomorphus* from the Palearctic region agree well with the type species of the genus in the body shape, prothoracic lateral tubercles, and pronotal tubercles. However, at least in one of the species examined by us, *Aegomorphus clavipes* (Schrank, 1781) (Figs. 16-18), the eyes are finely granulated as in the species of *Acanthoderes* and *Scythropopsis*.

According to Bates (1885) on *Sychnomerus*: "Two species of Central-American 'Acanthoderini' are here separated from *Acanthoderes* on the same grounds as *Discopus*, *Pteridotelus*, *Tetrasarus*, and *Plagiosarus* are separated, namely, differences in the clothing and structure of the antennae... The typical species of *Sychnomerus* has greatly the appearance of a *Pteridotelus*; but it wants the dentiform process of the undersides of joints 7, 9, and 10 which characterize that genus; what seem to be such processes in *Sychnomerus* prove on close examination to be dense tufts of hair." He defined the genus as follows (translated): "A genus differing from *Acanthoderes* only by the antennomeres III-XI densely setose ventrally, and VII-XI very short in females, and with long and abundant setae." Examination of photographs of the types of the two species included in *Sychnomerus*, *S. hirticornis* Bates, 1885, and *S. barbiger* Bates, 1885 (see photographs at Bezark, 2019), suggests that they do not belong to the same genus because the body shape differs, and the erect setae on the ventral surface of their antennomeres is noticeably different. *Sychnomerus hirticornis*, the type species, appears to be *Scythropopsis*. However, in the species of this latter genus examined by us, the distal antennomeres in males often have the setae as in *S. hirticornis* and, usually, the "tufts" in the basal antennomeres also very similar or identical. However, the females of *Sychnomerus* do not have long setae on distal antennomeres. Accordingly, while specimens of *S. hirticornis* and *S. barbiger* were not available for study, we are keeping *Sychnomerus* as a distinct genus.

***Symperasmus* Thomson, 1864 resurrection of the original status**

***Symperasmus alboniger* (Bates, 1861), comb. nov. (Figs. 19-34)**

Acanthoderes alboniger Bates, 1861: 218.

Acanthoderes albonigra; Gemminger, 1873: 3145 (cat.); Blackwelder, 1946: 610 (checklist); Zajciw, 1969a: 198 (distr.); 1970a: 5 (distr.).

Acanthoderes (Psapharochrus) albonigra; Aurivillius, 1923: 385 (cat.); Gilmour, 1965: 615 (cat.); Monné, 1994: 57 (cat.); Monné & Giesbert, 1994: 229 (checklist).

Psapharochrus alboniger; Lacordaire, 1872: 751; Monné, 2005: 200 (cat.); Monné et al., 2012: 32 (distr.); Monné, 2019: 289 (cat.).

Psapharochrus albonigrus; Wappes et al., 2006: 33 (distr.); Monné & Hovore, 2006: 209 (checklist).

Acanthoderes (Psapharochrus) albomaculatus Fuchs, 1963: 10; Gilmour, 1965: 615 (cat.). Syn. nov.

Acanthoderes albomaculata; Monné, 1994: 57 (cat.); Monné & Giesbert, 1994: 229 (checklist).

Psapharochrus albomaculatus; Monné, 2005: 200 (cat.); Monné & Monné, 2006: 42; Wappes et al., 2006: 33 (distr.); Monné, 2019: 289 (cat.).

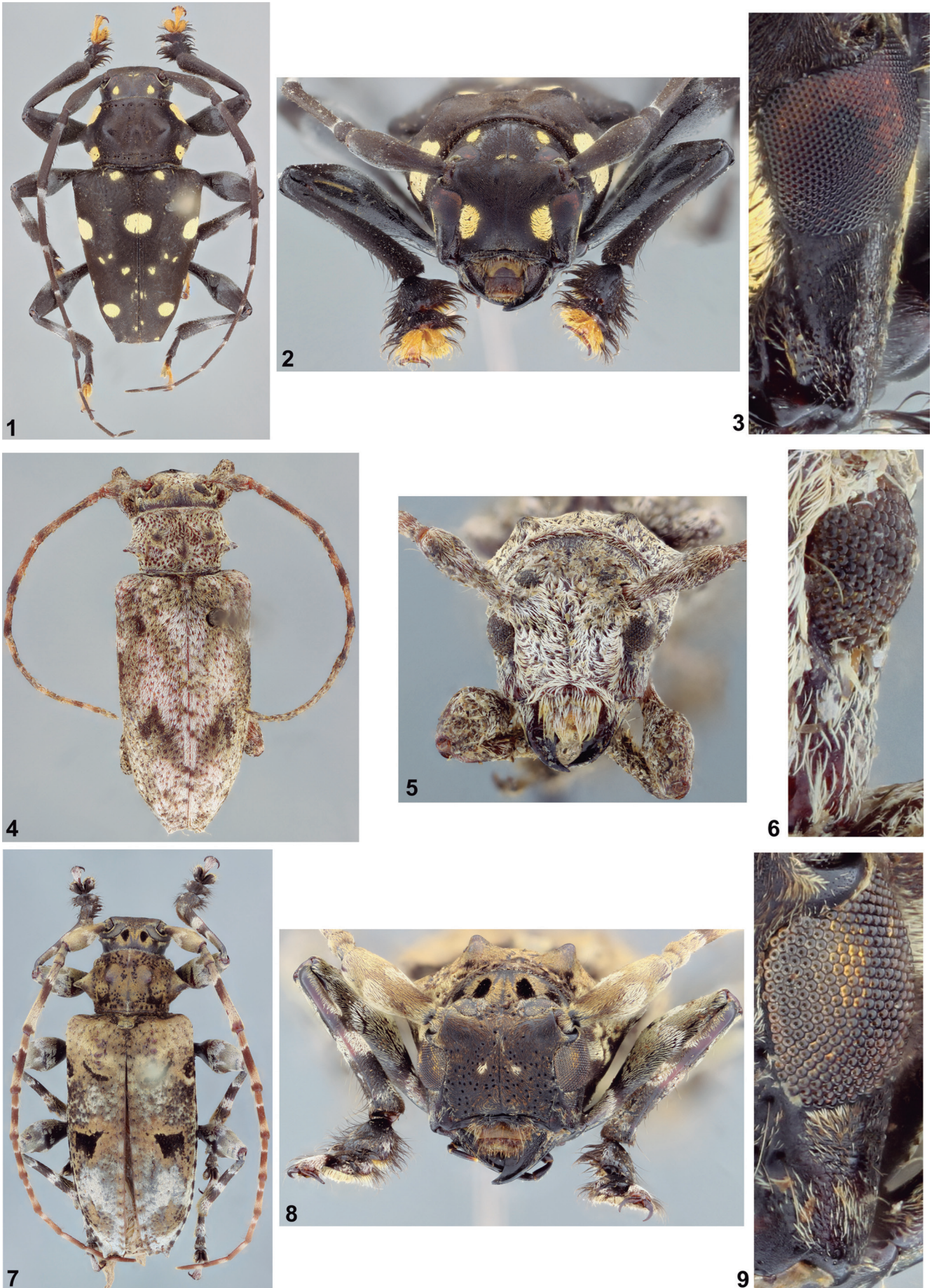
Acanthoderes griseomaculata Zajciw, 1971: 306; Julio et al., 2000: 30 (holotype). Syn. nov.

Acanthoderes (Psapharochrus) griseomaculata; Monné, 1994: 61 (cat.); Monné & Giesbert, 1994: 230 (checklist).

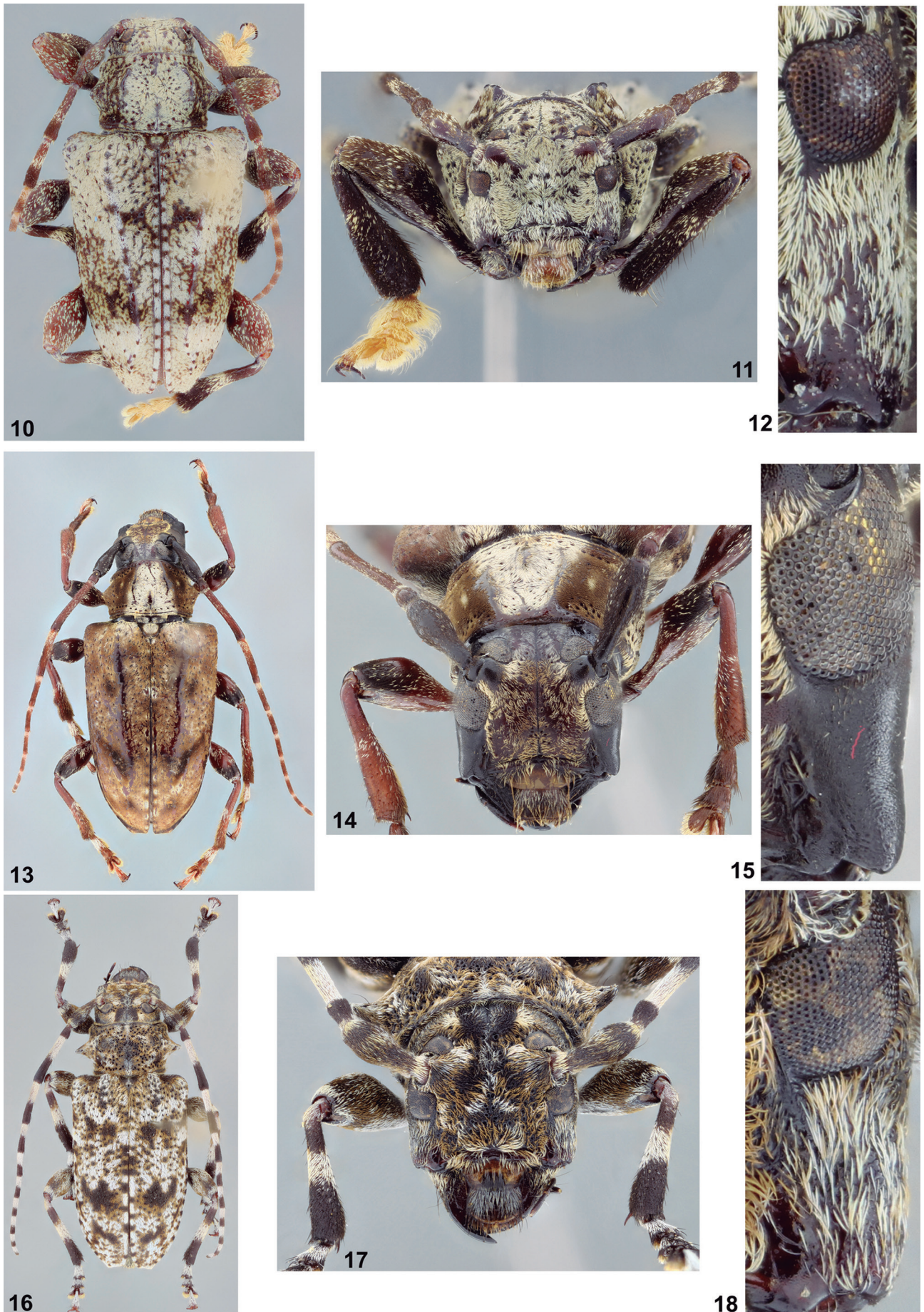
Psapharochrus griseomaculatus; Monné, 2005: 204 (cat.); Monné et al., 2012: 32 (distr.); Monné & Chaboo, 2015: 90 (distr.); Monné & Monné, 2016: 57 (holotype); Monné, 2019: 295 (cat.).

Bates (1861) described *Acanthoderes alboniger* (see photograph of the holotype at Bezark, 2019) based on a single female from Brazil (Pará). The holotype, as well as a series of specimens from Brazil (Amazonas), has the elytral apex covered with light yellowish-brown pubescence (but not obscuring punctures). Later, Fuchs (1963) described *Acanthoderes (Psapharochrus) albomaculatus* (see photograph of the holotype at Bezark, 2019) based on a single female from Bolivia and reported (translated): "A very excellent species, with a certain similarity to *A. laetifica* Bat. According to the description, the new species is related to *A. albonigra* Bat., unknown to me, from which it differs by the blunt lateral tubercles of the prothorax and by drawing of the elytra." However, although the elytral pubescent pattern is somewhat different in the holotypes of *A. albonigra* and *A. albomaculata* (*Acanthoderes* is feminine gender, and both names were originally wrongly utilized), the lateral tubercles of the prothorax are identical in both specimens. Bates (1861) incorrectly recorded them as acute, suggesting an apex much narrower than they really are in the species. Finally, Zajciw (1971) described *Acanthoderes griseomaculata* (see photograph of the holotype at Bezark, 2019) based on a single male from Peru but did not provide any comparison with other species.

Symperasmus alboniger has a noticeable variation of the elytral pubescent pattern color, as it can be yellow, partially yellow and partially pale-yellow, pale-yellow, partially pale-yellow and partially white, or white. Also, the centrodorsal pubescent macula of the elytra may be entire or fragmented, and the anterolateral pubescent spots may be small or large, fused or not with the centrodorsal macula. Furthermore, the apical elytral pubescence is also noticeably variable (Figs. 27-34). The specimens examined from Brazil have the elytral pubescence of the apex as in the holotypes of *A. albonigra* and *A. albomaculata*, while the specimens from Bolivia also have this area as in those holotypes as well as in the holotype of *A. griseomaculata* but with an intermediate pattern. Accordingly, we do not find a reliable feature that allows separation of the three species hence both *A. albomaculata* and *A. griseomaculata* are synonymized with *S. alboniger*.



Figures 1-9. (1-3) *Acanthoderes daviesii*, male. (1) Dorsal habitus. (2) Head, frontal view. (3) Lower eye lobe. (4-6) *Aegomorphus modestus*, female. (4) Dorsal habitus. (5) Head, frontal view. (6) Lower eye lobe. (7-9) *Aegomorphus cylindricus*, male. (7) Dorsal habitus. (8) Head, frontal view. (9) Lower eye lobe.



Figures 10-18. (10-12) *Scythropopsis albitarsis*, male. (10) Dorsal habitus. (11) Head, frontal view. (12) Lower eye lobe. (13-15) *Sympersasmus thoracicus*, female. (13) Dorsal habitus. (14) Head, frontal view. (15) Lower eye lobe. (16-18) *Aegomorphus clavipes*, female. (16) Dorsal habitus. (17) Head, frontal view. (18) Lower eye lobe.

The eyes in *S. albonigra* are not finely granulated, thus, it does not belong to *Acanthoderes*. Additionally, the pronotum is distinctly depressed centrally, a feature that also supports its placement in *Symperasmus*, not in *Acanthoderes* or *Psapharochrus* (= *Aegomorphus*) as previously placed.

Material examined (all from MZSP): PERU, *Junín*: Satipo, 1 female, 18.XII.1937, Paprzycki col.; 1 male, 1938-39, Meskendahl col.; (Sanibeni; 340 m), 1 male, 13.XI.1935, F. Woytowski col. *Huánuco*: Tingo Maria (Rio Huallaga), 2 males, X.1960, formerly Dirings collection. BOLIVIA, *Beni* (new department record): Uyapi (Guanay), 1 male, X-XI.1992, no collector indicated. *La Paz* (new department record): Ixiamas, 1921-22, M.R. Lopez col. *Cochabamba* (new department record): "Região de Chaparé", 400 m), 1 female, 05.IX.1956, Zischka col. BRAZIL, *Amazonas*: São Paulo de Olivença, 1 female, II.1923, S. Klages col.; 1 male, IV.1923, S. Klages col.; 1 male, V.1923, S. Klages col.; (Rio Javari), 1 female, III.1951, formerly Dirings collection; 1 female, VII.1956, formerly Dirings collection; 1 female, X.1960, formerly Dirings collection; 1 female, II.1961, formerly Dirings collection; 3 males, 2 females, X.1961, formerly Dirings collection; 2 males, XI.1961, formerly Dirings collection; 1 female, II.1962, formerly Dirings collection; 1 female, XI.1962, formerly Dirings collection; 1 male, 1 female, X.1963, formerly Dirings collection; (Rio Solimões), 1 male, XI.1960, formerly Dirings collection.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new records): Ecuador, Peru, Bolivia (Santa Cruz, Beni, La Paz, Cochabamba), Brazil (Amazonas, Pará).

***Scythropopsis* Thomson, 1864 resurrection of the original status**

***Scythropopsis melanostictica* (White, 1855), comb. nov. (Figs. 35-39)**

Acanthoderes melanosticticus White, 1855: 361.
Psapharochrus melanosticticus; Lacordaire, 1872: 751; Monné, 2005: 208 (cat.); Monné et al., 2010: 246 (distr.); Monné, 2019: 301 (cat.).
Acanthoderes melanosticta; Gemminger, 1873: 3146 (cat., error); Blackwelder, 1946: 611 (cat.); Zajciw & Seabra, 1968: 72 (distr.); Zajciw, 1969b: 610 (distr.), 1970a: 5 (distr.).
Acanthoderes (Psapharochrus) melanosticta; Aurivillius, 1923: 387 (cat.); Buck, 1959: 604 (distr.); Gilmour, 1965: 615 (cat.); Monné, 1994: 65 (cat.); Mermudes & Monné, 2001: 331 (error); Monné et al., 2017: 30 (morphology).
Acanthoderes (Psapharochrus) melanostictus; Monné & Giesbert, 1994: 231 (checklist).
Pteridotelus contaminatus Thomson, 1865: 544; Gemminger, 1873: 3144 (cat.); Thomson, 1878: 15 (type); Aurivillius, 1923: 383 (cat.); Blackwelder,

1946: 610 (checklist); Gilmour, 1965: 611 (cat.); Monné, 1994: 53 (cat.); Monné & Giesbert, 1994: 237 (checklist); Monné, 2005: 215 (cat.); Monné & Hovore, 2006: 211 (checklist); Monné, 2019: 308. Syn. nov.

Audinet-Serville (1835) included some species in his new genus *Acanthoderes* which indicated it was masculine gender (*Acanthoderes varius* (Fabricius, 1787), and *Acanthoderes griseus* (Fabricius, 1792)). However, it is actually feminine gender, as previously correctly indicated by Gemminger (1873) and Blackwelder (1946). Gemminger (1873) corrected the gender in only part of the species neglecting to do so for *Acanthoderes melanosticticus*.

Lacordaire (1872) transferred *A. melanosticticus* to *Psapharochrus* Thomson, 1864. As Aurivillius (1923) considered *Psapharochrus* a subgenus of *Acanthoderes*, *A. melanosticticus* was mentioned in this subgenus until recently by nearly all authors, even after *Psapharochrus* had been considered a distinct genus. In *Acanthoderes melanostictica* the size of the ommatidia is very small (Figs. 37-38) and the elytra have distinct carina (Santos-Silva & Nascimento, 2018), hence it belongs to *Scythropopsis* Thomson, 1864. It also has the protibiae distinctly widened toward apex (Fig. 39), as in the type species of the genus.

Thomson (1865) described two species, in different genera, with the same name: *Psapharochrus contaminatus* (Fig. 40), and *Pteridotelus contaminatus* (Fig. 36). Monné & Giesbert (1992) synonymized *Psapharochrus contaminatus* with *Acanthoderes melanostictus* [sic]: "*Acanthoderes melanostictus* White, 1855: 361/ *Acanthoderes* [sic] *contaminatus* Thomson, 1865: 543, new synonymy." However, *Psapharochrus contaminatus* is not equal to *Scythropopsis melanostictica* (Fig. 35), rather it is *Pteridotelus contaminatus* that is the true synonym of the latter.

Material examined (all from MZSP): BRAZIL, *Minas Gerais* (new state record): Poços de Caldas, 1 female, 11.XI.1954, J.J. Ferracioli col.; Passa Quatro, 1 female, 14.XI.1916, Zikán col.; (Fazenda dos Campos), 1 male, 30.XII.1917, Zikán col. *São Paulo*: São Paulo, 1 female, 05.XII.1914, collector illegible; 1 female, 1915, no collector indicated; (Ipiranga), 1 male, 2 females, no date and collector indicated; 1 female, 25.I.1957, collector illegible; (Jabaquara), 1 female, 13.II.1974, L.R. Fontes col.; Campinas, 1 female, XI.1919, collector illegible. *Paraná*: Guarapuava, 1 female, III.1953, H. Schneider col.; 1 female, I.1960, H. Schneider col.; Ponta Grossa, 1 female, XII.1938, Camargo col.; 1 male, 1941, Machado col.; Rio Negro, 1 female, 07.I.1924, no collector indicated; 1 male, 22.XII.1924, no collector indicated; 1 female, I.1926, no collector indicated; 1 female, 09.III.1927, no collector indicated; Curitiba, 1 male, 05.II.1936, no collector indicated. *Santa Catarina*: São Bento do Sul, 1 male, XI.1924, A. Maller col.; 1 female, II.1950, formerly Dirings col.; 1 male, II.1952, formerly Dirings col.; 2 females, II.1960, formerly Dirings col.; (Rio Vermelho), 1 male, XII.1949, formerly Dirings col.; 2 females, I.1950, formerly Dirings col.; 1 male, I.1952, formerly Dirings col.; 1 male, II.1952,

formerly Dirings col.; 1 female, X.1952, formerly Dirings col.; 1 male, XII.1952, formerly Dirings col.; 1 male, I.1958, formerly Dirings col.; 6 males, 4 females, III.1960, formerly Dirings col.; 2 females, I.1961, formerly Dirings col.; 1 female, III.1961, formerly Dirings col.; 1 female, I.1962, formerly Dirings col.; Timbó, 4 males, 3 females, X.1958, formerly Dirings col.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new record): Brazil (Minas Gerais, Espírito Santo, Rio de Janeiro, São Paulo, Paraná, Santa Catarina, Rio Grande do Sul).

***Scythropopsis lacrymans* (Thomson, 1865),
comb. nov.
(Fig. 42)**

Pteridotelus lacrymans Thomson, 1865: 544; Gemminger, 1873: 3145 (cat.); Thomson, 1878: 15 (type).

Acanthoderes lacrymans; Bates, 1880: 140; Blackwelder, 1946: 610 (checklist); Ordóñez-Reséndiz & Martínez-Ramos, 2017: 828 (distr.).

Acanthoderes (*Psapharochrus*) *lacrymans*; Aurivillius, 1923: 386 (cat.); Gilmour, 1965: 613 (cat.); Monné, 2005: 164 (cat.).

Pteridotelus lachrymosus; Bates, 1880: pl. 11, fig. 6 (error). *Acanthoderes lachrymans*; Chemsak *et al.*, 1992: 130 (checklist); Noguera & Chemsak, 1996: 406 (distr.).

Acanthoderes (*Pardalisia*) *lacrymans*; Monné, 1994: 70 (cat.); Monné & Giesbert, 1994: 229 (checklist); Vargas-Cardoso *et al.*, 2018: 96 (hosts); Monné, 2019: 225 (cat.).

Thomson (1865) described *Pteridotelus lacrymans* without explaining its inclusion in the genus. Since the main feature of *Pteridotelus* (last antennal segments distinctly widened), is not present in this species it is incorrect to maintain it in the genus. The species *Pteridotelus lacrymans* appears in Monné & Giesbert (1994), and Monné (1994) as an *Acanthoderes*, without explanation in either reference. Factually, *Pteridotelus lacrymans* cannot be placed in *Acanthoderes* (*Pardalisia*), because the antennae are distinctly longer, with distinct ventral pubescence, clearly forming tufts in antennomeres III and IV (distinctly shorter and glabrous in *A. (P.) funeraria* Bates, 1861, elytra carinate toward distal area (not so in *A. (P.) funeraria*). Due to the antennal setae *Pteridotelus lacrymans* also has affinities with *Tetrasarus* Bates, 1880. However, *Tetrasarus* is another problematic genus with species assigned to but not belonging to it. The presence of a dense tuft of long setae on the apex of antennomeres III and IV is a definitive character of *Tetrasarus* and is lacking in *P. lacrymans*. Although *Pteridotelus lacrymans* differs somewhat in general appearance to other *Scythropopsis* species it is provisionally placed in the genus because of the common characters previously commented on.

Material examined: MEXICO, Morelos Cuernavaca, 1 male, no date indicated, Martins col. (MZSP).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new record): Mexico (Oaxaca, Puebla, Guerrero, Morelos).

***Scythropopsis nigratarsis* (White, 1855), comb. nov.
(Figs. 43-54)**

Acanthoderes nigratarsis White, 1855: 363; Gemminger, 1873: 3146 (cat.); Bates, 1880: 141 (distr.); Gahan, 1892: 264 (syn.); Blackwelder, 1946: 611 (checklist); Chemsak *et al.*, 1992: 130 (cat.); Noguera & Chemsak, 1996: 406 (cat.); Chemsak & Hovore, 2002b: 11.

Acanthoderes (*Psapharochrus*) *nigratarsis*; Aurivillius, 1923: 387 (cat.); Gilmour, 1965: 613 (cat.); Monné & Giesbert, 1994: 231 (checklist); Monné, 1994: 231 (cat.).

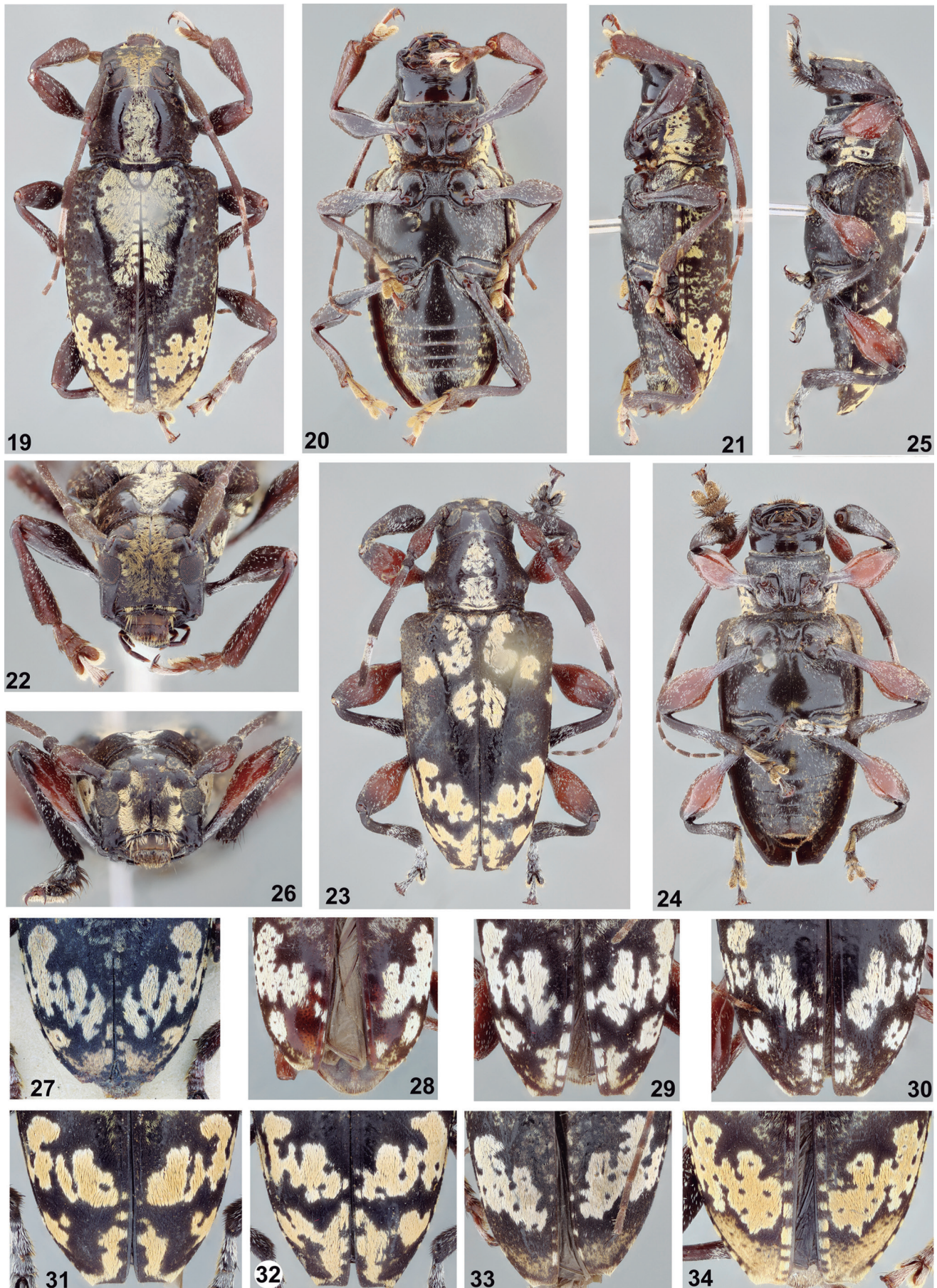
Psapharochrus nigratarsis; Turnbow *et al.*, 2003: 29 (distr.); Monné, 2005: 209 (cat.); Hovore, 2006: 376 (distr.); Swift *et al.*, 2010: 46 (distr.); Monné, 2019: 302 (cat.).

Acanthoderes sylvanus Bates, 1880: 141, 1885: 378.

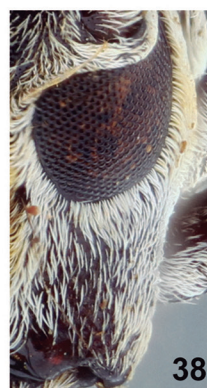
Acanthoderes sylvana; Lameere, 1883: 62 (checklist).

Description: Female (Figs. 48-52): Integument black, slightly more dark brown on some areas.

Head: Frons, vertex, and area behind eyes with dense, white, yellowish-brown, and pale-yellow pubescence mixed; with sparse, long, erect dark setae close to eyes. Genae with dense pubescence as on dorsal surface of head close to eye, narrow area close to frons glabrous, distinctly sparser on wide remaining surface, with a few long, erect dark setae interspersed. Postclypeus with dense, long white, yellowish-brown, and pale-yellow pubescence intermixed, with sparse, long, erect dark setae interspersed, central area and sides glabrous. Posterior $\frac{2}{3}$ of labrum coplanar with anteclypeus, inclined at anterior third; with dense white, yellowish-brown, and pale-yellow pubescence intermixed on sides of posterior $\frac{2}{3}$ (whiter and shorter toward central area), distinctly sparser centrally, sparse on inclined area, especially centrally, and with fringe of yellowish-brown setae at anterior margin; with transverse, sparse row of long, erect, dark setae centrally. Gula mentum slightly convex, posterior $\frac{2}{3}$ glabrous, depressed, finely punctate, anterior third with short yellowish-brown pubescence not obscuring integument. Distance between upper eye lobes 0.62 times length of scape; in frontal view, distance between lower eye lobes 0.94 times length of scape. Antennae 1.2 times elytral length, reaching posterior seventh of elytra. Scape clavate, flattened dorsally at basal third; with brown pubescence not obscuring integument, with short, moderately abundant white pubescence interspersed on posterior $\frac{2}{3}$, except narrow posterior area with dense white pubescence; with a few long, erect dark setae on posterior region of ventral surface. Antennomeres with basal white pubescent ring, gradually narrower toward posterior segments (pubescence sparser on III), and remaining surface with brown pubescence not obscuring integument; pedicel and antennomeres III-VI with sparse,



Figures 19-34. *Symperasmus alboniger*. (19-22) Male from Brazil (Amazonas, Benjamin Constant). (19) Dorsal habitus. (20) Ventral habitus. (21) Lateral habitus. (22) Head, frontal view. (23-26) Male from Peru (Huánuco, Tingo María). (23) Dorsal habitus. (24) Ventral habitus. (25) Lateral habitus. (26) Head, frontal view. (27-34) Elytral apex. (27) Female, Bolivia. (28) Female, Bolivia. (29) Female, Peru. (30) Male, Peru. (31) Male, Peru. (32) Male, Peru. (33) Male, Bolivia. (34) Male, Brazil.



Figures 35-42. (35-39) *Acanthoderes melanosticticus*, (35) Syntype, dorsal view. (36) *Pteridotelus contaminatus*, holotype, dorsal habitus. (37) Male from Brazil (Santa Catarina), head, frontal view. (38) Idem, lower eye lobe. (39) Idem, protibia. (40) *Psapharochrus contaminatus*, holotype male, dorsal habitus. (41) *Lamia modesta*, holotype, dorsal habitus. (42) *Pteridotelus lacrymans*, holotype, dorsal habitus.

long, erect dark setae ventrally (gradually shorter, sparser toward VI). Antennal formula (ratio) based on length of antennomere III: scape = 0.80; pedicel = 0.23; IV = 0.77; V = 0.46; VI = 0.36; VII = 0.35; VIII = 0.27; IX = 0.23; X = 0.19; XI = 0.19.

Thorax: Lateral tubercle of prothorax large, conical, slightly curved upward, with apex blunt. Pronotum with large, wide tubercle on each side, gradually elevated from posterior quarter toward anterior quarter, with more elevated area slightly bifid at top; with narrow, carina-shaped tubercle centrally, from posterior sulcus to anterior margin, gradually widened toward posterior sulcus; posterior sulcus well-marked; coarsely, sparsely punctate between tubercles, forming transverse row in posterior sulcus, and a few coarse punctures on dorsal surface of lateral tubercles of prothorax; with dense, white, yellowish-brown, and pale-yellow pubescence intermixed, with a few long, erect dark setae interspersed on sides of posterior area, greenish-brown pubescent band adjacent to outer side of lateral tubercles of pronotum, widened anteriorly and posteriorly, anterior $\frac{3}{4}$ of central tubercle glabrous, anterior apex of lateral tubercles of pronotum nearly glabrous; posterior fourth with white pubescent band centrally. Sides of prothorax coarsely, sparsely punctate; with dense, white, yellowish-brown, and pale-yellow pubescence intermixed. Prosternum with white, yellowish-brown, and pale-yellow pubescence intermixed, denser on some areas. Prosternal process 0.75 times width of procoxal cavity, coarsely rugose-punctate, longitudinally sulcate centrally, with apex strongly emarginate; pubescence with same color as on prosternum, distinctly sparser toward apex. Ventral surface of mesothorax with dense, white, yellowish-brown, and pale-yellow pubescence intermixed laterally, yellowish-brown centrally. Mesoventral process 1.5 times width of mesocoxal cavity, with one distinct tubercle each side of anterior area; with white, yellowish-brown, and pale-yellow pubescence intermixed, not obscuring integument. Ventral surface of metathorax with dense, white, yellowish-brown, and pale-yellow pubescence intermixed, sparser centrally. Scutellum with dense yellowish-brown pubescence on anterocentral area, with white pubescence interspersed, narrow white pubescent band on central area of posterior third, and brown pubescence on remaining surface. **Elytra:** Humeri somewhat projected forward; with elevated centrobasal crest covered with small tubercles between humeri and scutellum, and distinct carina from apex of centrobasal crest to near apex; posterior margin concave, with outer angle distinctly triangularly projected; pubescence dense, mostly white, yellowish-brown, and pale-yellow pubescence intermixed, with dense, slightly oblique, wide white pubescent band laterally on anterior third, small greenish-brown spots surrounding punctures, and irregular areas with greenish-brown pubescence; with sparse, moderately long, erect dark setae throughout. **Legs:** Femora with dense, white, yellowish-brown, and pale-yellow pubescence intermixed. Tibiae with moderately dense, white, yellowish-brown, and pale-yellow

pubescence intermixed on dorsal and lateral sides of basal area, and wide central area (this later area somewhat projected toward apex along outer surface), with white pubescence on ventral surface of basal area and dorsal surface of apex, and brownish pubescence on remaining surface, not obscuring integument; with long, erect, sparse dark setae. Tarsomeres I and V with mostly white pubescence dorsally, and remaining segments with brownish pubescence not obscuring integument.

Abdomen: Ventrites with white, yellowish-brown, and pale-yellow pubescence intermixed, sparser on wide central area of V; apex of ventrite V nearly truncate.

Dimensions (mm): Total length, 16.7; prothoracic length, 3.2; anterior prothoracic width, 4.2; posterior prothoracic width, 4.4; maximum prothoracic width, 5.9; humeral width, 7.2; elytral length, 11.9.

Material examined: MEXICO, *Chiapas* (new state record): Reserva Biosfera El Triunfo, 15°39'N, 92°49'W, 1 female, 15.XI.2001, C.W. O'Brien col. (FWSC). *Oaxaca*: MX175, 5 km N Portillo de Rayo, 2 males, 20.X.2005, F. Skillman & B. Eya col. (FWSC; MZSP).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new record): Mexico (Oaxaca, Chiapas), Guatemala, Honduras, Costa Rica.

Remarks: The outer angle of the elytral apex in the female examined is more triangularly projected than in the two males examined. However, based on the study of other species of the genus, as well as similar genera, this feature is considered a variation. The elytral pubescence, in *S. nigritarsis* varies widely as it does in other species of the genus.

The finely granulate eyes and distinct elytral carina observed in photographs of the holotype and the other material examined ensure us that the species belongs to *Scythropopsis*.

***Scythropopsis sallei* (Thomson, 1865), comb. nov.
(Fig. 55)**

Psapharochrus sallei Thomson, 1865: 543, 1878: 15 (type); Monné, 2005: 212 (cat.), 2019: 305 (cat.).

Psapharochrus saillei (error); Lacordaire, 1872: 751.

Acanthoderes sallaei (error); Bates, 1880: 141 (distr.); Noguera & Chemsak, 1996: 406 (checklist).

Acanthoderes (Psapharochrus) sallei; Aurivillius, 1923: 388 (cat.); Gilmour, 1965: 613 (cat.); Monné & Giesbert, 1994: 231 (checklist); Monné, 1994: 67 (cat).

Acanthoderes sallei; Blackwelder, 1946: 611 (checklist); Zajciw, 1970b: 187; Chemsak *et al.*, 1992: 130 (checklist).

Thomson (1865) described the species from Mexico, without a specific locality. Subsequently Bates (1880) provided: Córdoba (= Córdoba) in the Mexican state of

Veracruz as a collection locality. Noguera & Chemsak (1996) reiterated the Bates (1880) information.

Although the outer angle of the elytral apex is distinctly projected on the holotype of *Psapharochrus sallei*, and not so in the holotypes of *Acanthoderes nigratarsis* and *Acanthoderes sylvanus*, it is still possible that they belong to the same species. For example, the female of *Scythropopsis nigratarsis* examined by us has the elytral apex intermediate between the three holotypes. This kind of variation is not unusual in Acanthoderini as, for example, in *Aegomorphus jaspideus* (Germar, 1823) where the outer angle of the elytra is usually distinctly projected but, can also be slightly or even not at all projected.

The finely granulate eyes and distinct elytral carina, as shown in the photograph of the holotype, support the species transference to *Scythropopsis*.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Mexico (Veracruz).

***Scythropopsis cornuta* (Bates, 1880), comb. nov.**
(Figs. 56-61)

Acanthoderes cornutus Bates, 1880: 142.

Acanthoderes cornuta; Lameere, 1883: 62 (cat.); Blackwelder, 1946: 610 (checklist); Chemsak *et al.*, 1992: 130 (checklist).

Acanthoderes (Psapharochrus) cornuta; Aurivillius, 1923: 386 (cat.); Gilmour, 1965: 614 (cat.); Monné & Giesbert, 1994: 230 (checklist); Monné, 1994: 60 (cat.).

Psapharochrus cornuta; Turnbow *et al.*, 2003: 29 (distr.).

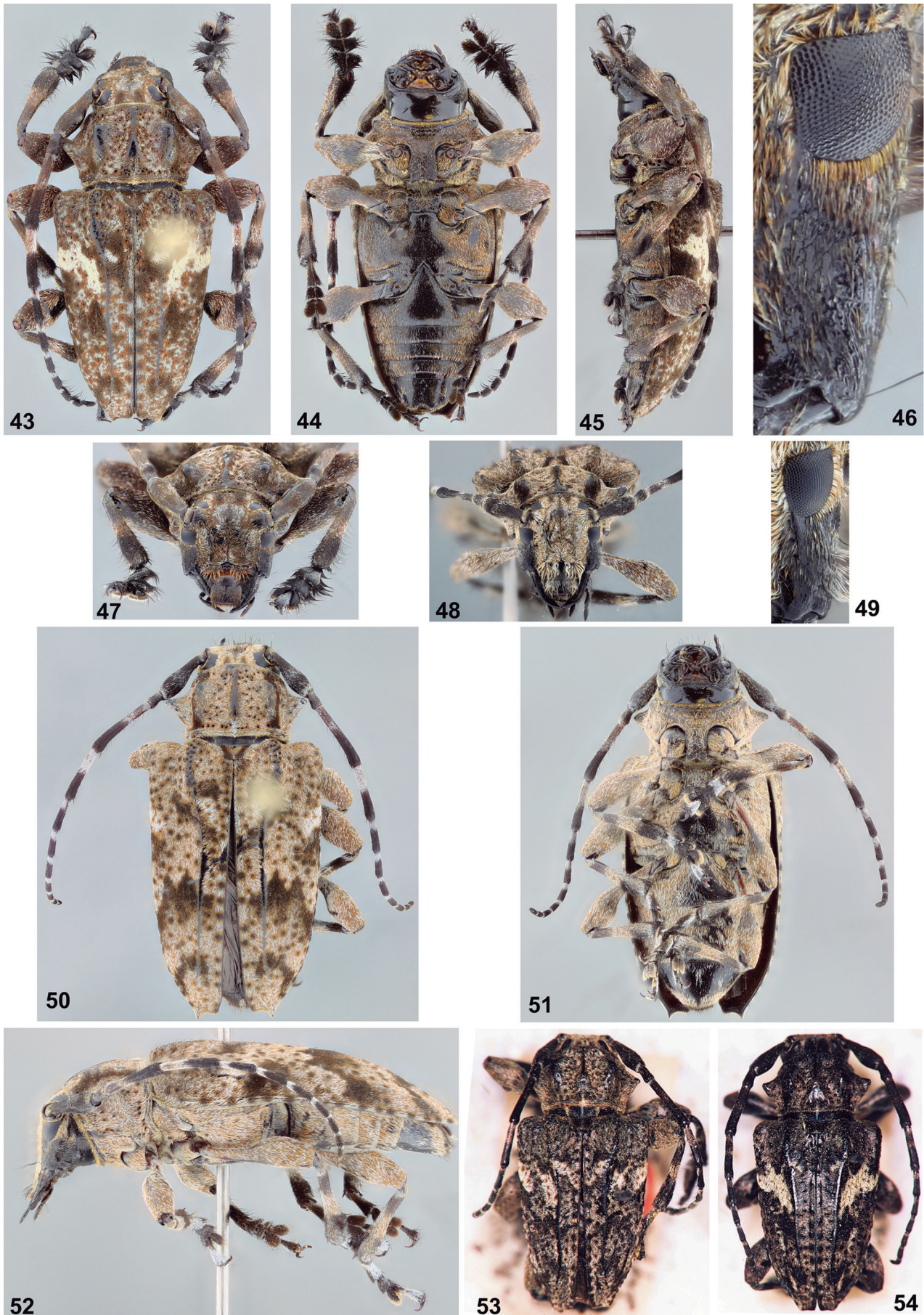
Psapharochrus cornutus; Monné, 2005: 203 (cat.); Hovore, 2006: 376 (distr.); Monné, 2019: 293 (cat.).

Description: Male (Figs. 57-61): Integument mostly dark brown, almost black on some areas; anteclypeus, apex of labrum, parts of mouthparts, and apex of palpomeres reddish-brown; antennomeres V-XI with orange ring on basal third; central area of anterior third of elytra with large, triangular reddish-brown macula.

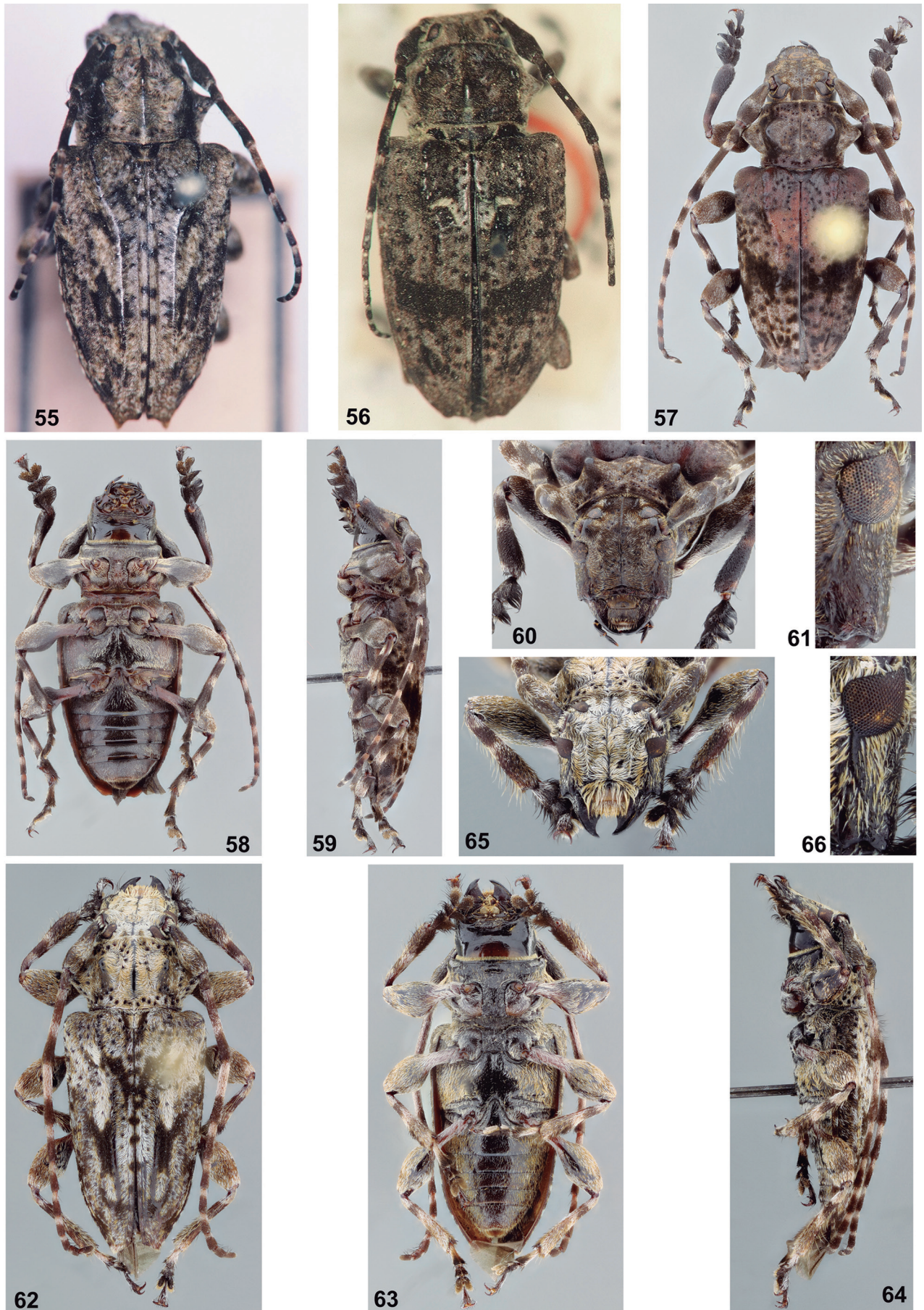
Head: Frons, area between antennal tubercles and beginning of upper eye lobes finely, sparsely punctate; with moderately dense brownish pubescence (more reddish-brown close to eyes and postclypeus), with white pubescence interspersed. Remaining surface of vertex smooth, with dark brown pubescence centrally, partially obscuring integument, and longitudinal yellowish-white pubescent band laterally. Area behind upper eye lobes with sparse fringe of yellowish-white pubescence close to eye, glabrous on narrow sulcate area close to eye, with yellowish-brown pubescence close to sulcate area, nearly glabrous toward prothorax. Area behind lower eye lobes glabrous on sulcus adjacent to eye, with narrow (widened near inferior curvature of eye) yellowish-brown pubescent band close to sulcus (pubescence sparser than behind lower eye lobe), remaining surface glabrous. Genae almost twice length of lower eye lobe; with fringe of yellowish-brown pubescence close to eye,

and sparse yellowish-brown pubescence, with white pubescence interspersed on remaining surface, narrow distal area glabrous. Postclypeus coarsely, sparsely punctate on wide central area, smooth laterally; with short, bristly reddish-brown pubescence not obscuring integument on wide central area more so on center of this area, laterally glabrous; with long, sparse, erect dark setae on wide central area. Posterior $\frac{2}{3}$ of labrum coplanar with anteclypeus, inclined at anterior third; with yellowish-brown pubescence not obscuring integument, on posterior $\frac{2}{3}$, anterior third nearly glabrous, anterior margin with fringe of brown setae (apex of nearly all setae yellowish); with transverse row of long, erect, dark setae near curvature between coplanar and inclined area. Gulamentum glabrous, wide posterior area smooth, except transverse striae on center of anterior region of this area; anterior area depressed, transversely striate, with sparse reddish-brown pubescence close to anterior margin. Distance between upper eye lobes 0.55 times length of scape; in frontal view, distance between lower eye lobes equal to length of scape. Antennae 1.65 times elytral length, reaching elytral apex near apex of antennomere VIII. Scape clavate, distinctly sulcate dorsally at anterior third; dorsally with reddish-brown pubescence except wide transverse white pubescent band centrally, and narrow white pubescent band at apex; remaining surface with white pubescence not obscuring integument. Pedicel with transverse white pubescent band dorsally near base, brown on remaining dorsal surface except yellowish-white on outer side close to apex; ventral and lateral surfaces with white pubescence. Antennomeres III and IV with two white pubescent rings, one basally, another after middle, ventrally fused by longitudinal band; remaining surface with brown pubescence. Antennomeres V-XI with white pubescence on basal half, brown on distal half; distal third of antennomere IX and basal $\frac{2}{3}$ of antennomere X with long, erect dark setae. Antennal formula (ratio) based on length of antennomere III: scape = 0.73; pedicel = 0.29; IV = 0.76; V = 0.45; VI = 0.39; VII = 0.35; VIII = 0.31; IX = 0.27; X = 0.21; XI = 0.24.

Thorax: Lateral tubercle of prothorax large, conical. Pronotum with large, nearly conical tubercle each side, with wide, rounded apex; with narrow, carina-shaped tubercle centrally, from posterior sulcus to anterior margin, slightly widened posteriorly; posterior sulcus distinct laterally, nearly indistinct toward center; coarsely, sparsely punctate between tubercles, slightly coarser and more abundant between anterior margin and base of lateral tubercles, and along posterior sulcus, shallower, sparser on lateral tubercles of prothorax; with white, yellowish-brown, and pale-yellow pubescence intermixed, partially obscuring integument in some areas. Sides of prothorax coarsely, sparsely punctate (punctures distinctly coarser close to posterior margin); pubescence as on pronotum. Prosternum with pale-yellow pubescence not obscuring integument. Prosternal process with widest central area about as wide as procoxal cavity; lateral margins sinuous and elevated; central area longitudinally slightly elevated; with sparse pale-yellow pubescence.



Figures 43-54. *Scythropopsis nigritarsis*. (43-47) Male. (43) Dorsal habitus. (44) Ventral habitus. (45) Lateral habitus. (46) Lower eye lob. (47) Head, frontal view. (48-52) Female. (48) Head, frontal view. (49) Lower eye lob. (50) Dorsal habitus. (51) Ventral habitus. (52) Lateral habitus. (53) Holotype male, dorsal habitus. (54) *Acanthoderes sylvanus*, holotype male, dorsal habitus.



Figures 55-66. (55) *Psapharochrus sallei*, holotype female, dorsal habitus. (56-61) *Scythropopsis cornuta*. (56) Holotype female, dorsal habitus. (57) Dorsal habitus, male. (58) Ventral habitus, male. (59) Lateral habitus, male. (60) Head, frontal view, male. (61) Lower eye lobe, male. (62-66) *Scythropopsis wappesi*, male. (62) Dorsal habitus. (63) Ventral habitus. (64) Lateral habitus. (65) Head, frontal view. (66) Lower eye lobe.

Ventral surface of meso- and metathorax with pale-yellow pubescence not obscuring integument, sparser on center of meso- and metaventrite. Mesoventral process with apex slightly wider than mesocoxal cavity; with one moderately large tubercle each side of anterior area; lateral margins slightly narrowed centrally, distinctly widened at apex. Scutellum with yellowish-white pubescence not obscuring integument. **Elytra:** Humeri rounded, somewhat projected forward; with elevated centrobasal crest between humeri and scutellum, nearly conically elevated anteriorly, covered with small tubercles, and distinct carina from apex of centrobasal crest to near apex; with another carina between the crest and humeri; elytral carina fused at their apex; coarsely, sparsely punctate, punctures sparser, shallower toward apex, with basal punctures anteriorly bordered by small tubercle; apex truncate, slightly concave centrally; pubescence mostly yellowish-white, less so between centrobasal crests, except wide, irregular, slightly oblique dark-brown pubescent band about middle (with irregular yellowish-white macula on inclined area), and dark-brown spots surrounding punctures; U-shaped white pubescent band on basal third partially lost in the specimen examined. **Legs:** Femora with yellowish-white pubescence partially obscuring integument on some areas, with brown pubescent areas interspersed on club. Tibiae with three rings of yellowish-white pubescence, one basally, one centrally, another at apex; remaining surface with brown pubescence. Protarsi with dark-brown pubescence dorsally, with a few whitish setae interspersed on tarsomere I. Tarsomere I of meso- and metatarsi with dense white pubescence on posterior $\frac{2}{3}$ of dorsal surface, dark-brown pubescence basally; tarsomeres II-IV with dark-brown pubescence; tarsomere V with white pubescence not obscuring integument.

Abdomen: Ventrites with yellowish-white pubescence not obscuring integument, sparser centrally, with distal area of ventrites I-IV glabrous; distal margin of ventrite V concave.

Dimensions (mm): Total length, 12.80; prothoracic length, 2.60; anterior prothoracic width, 3.20; posterior prothoracic width, 3.15; maximum prothoracic width, 4.55; humeral width, 5.15; elytral length, 8.20.

Material examined: NICARAGUA (new country record): *Nueva Segovia*: Cerro Jesus (13°58'N, 86°10'W; 1,100-1,200 m), 1 male, VI-VII.2016, E. van den Berghe col. (ACMT).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new record): Guatemala, Honduras, Nicaragua.

Remarks: Bates (1880) described the species from Guatemala, based on a single female. Subsequently, the species has only been mentioned in catalogs and checklists, except for Turnbow *et al.* (2003), who provided Honduras as a new country record. Examination of

the photograph of the holotype, as well as the male described here, reveals that the eyes are finely granulated, and the elytra distinctly carinate. Accordingly, the species is transferred to *Scythropopsis*.

***Scythropopsis wappesi* (Chemsak & Hovore, 2002),
comb. nov.
(Figs. 62-66)**

Acanthoderes wappesi Chemsak & Hovore, 2002b: 7; Monné, 2005: 163 (cat.); Hovore, 2006: 376 (distr.).
Acanthoderes (Acanthoderes) wappesi Monné, 2005: 163 (cat.); Monné & Hovore, 2006: 203 (checklist).
Psapharochrus wappesi; Tavakilian, 2018: 39 (comb. nov.); Monné, 2019: 307 (cat.).

According to Tavakilian (2018): "The photography of the type available on Bezark's Catalog (2018)... shows clearly that the species described as *Acanthoderes wappesi* Chemsak & Hovore, 2002... belongs to the genus *Psapharochrus* Thomson, 1863 and not to the genus *Acanthoderes* Audinet-Serville, 1835. This necessary new combination induces a new homonym and the latest species described in honour of James Wappes must be renamed." However, Chemsak & Hovore (2002b) reported the eyes as being "finely faceted", which was confirmed in the specimen examined by us. As the species also has distinctly elytral carina, it belongs to *Scythropopsis* and thus negates the need for a replacement name.

Material examined: MEXICO (new country record): *Tamaulipas*: 10 km E Tula (3900'), 1 male, 10.V.1994, J.E. Wappes col. (ACMT).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new record): Guatemala, Mexico (*Tamaulipas*).

***Scythropopsis barrerai* (Chemsak & Hovore, 2002),
comb. nov.
(Figs. 67-71)**

Acanthoderes barrerai Chemsak & Hovore, 2002a: 13.
Acanthoderes (Acanthoderes) barrerai Monné, 2005: 160 (cat.); Monné & Hovore, 2006: 202 (checklist); Monné, 2019: 220 (cat.); Santos-Silva *et al.*, 2018: 205 (distr.).

As in the photograph of the holotype, as well as the female from the MZSP collection, the eyes are finely granulated and the elytra are distinctly carinate, the species is appropriately transferred to *Scythropopsis*.

Material examined: MEXICO, *Veracruz*: Los Tuxtlas (900 m), 1 female, 01-15.VI.2016, J.H. Garcia col. (MZSP).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Mexico (*Querétaro*, *Hidalgo*, *Veracruz*).

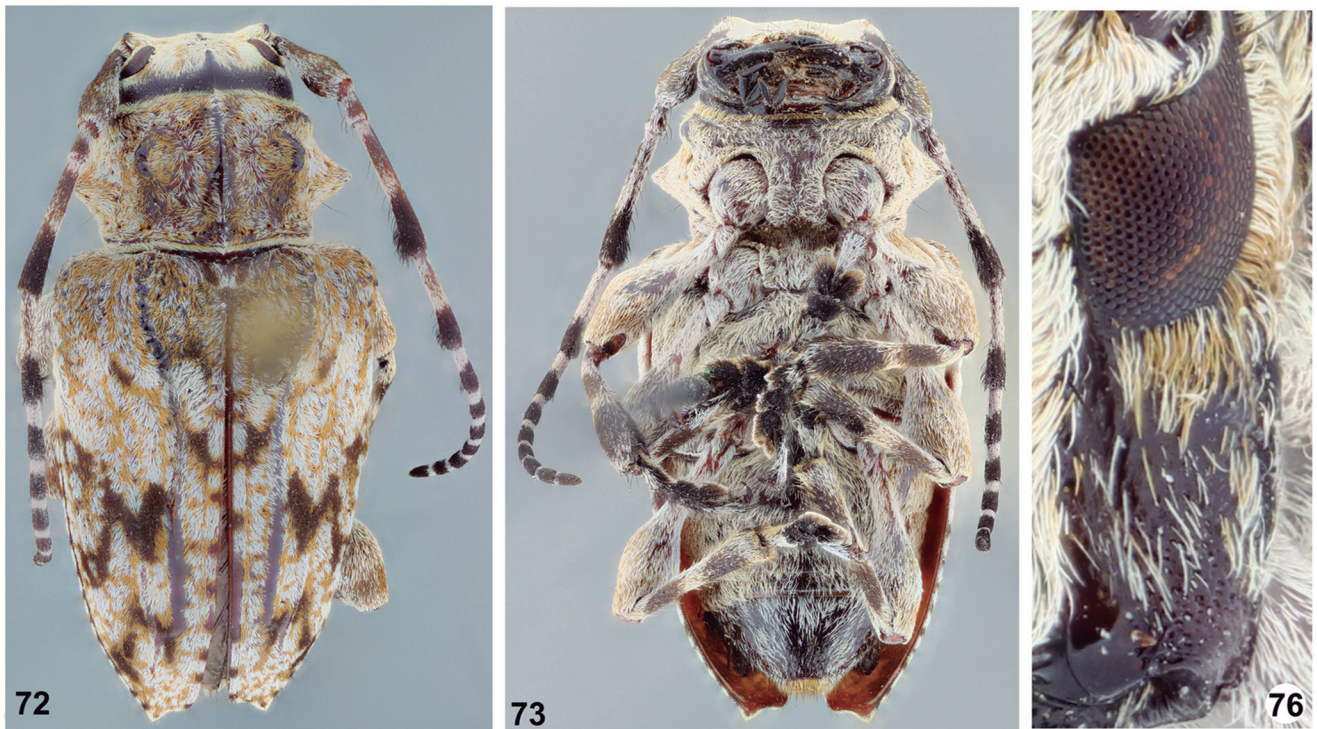
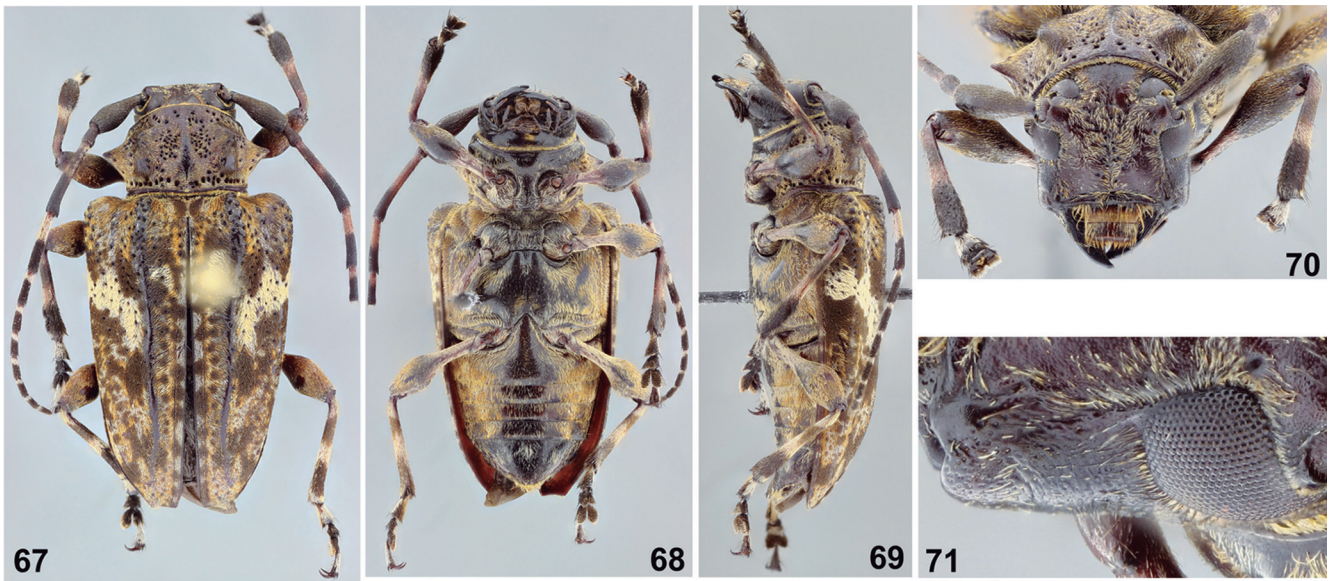
***Scythropopsis intricata* sp. nov.**
(Figs. 72-76)

Description: Female: Integument mostly black; mouth-parts dark reddish-brown with darkened areas, except palpi of last palpomeres black with yellowish-brown apex.

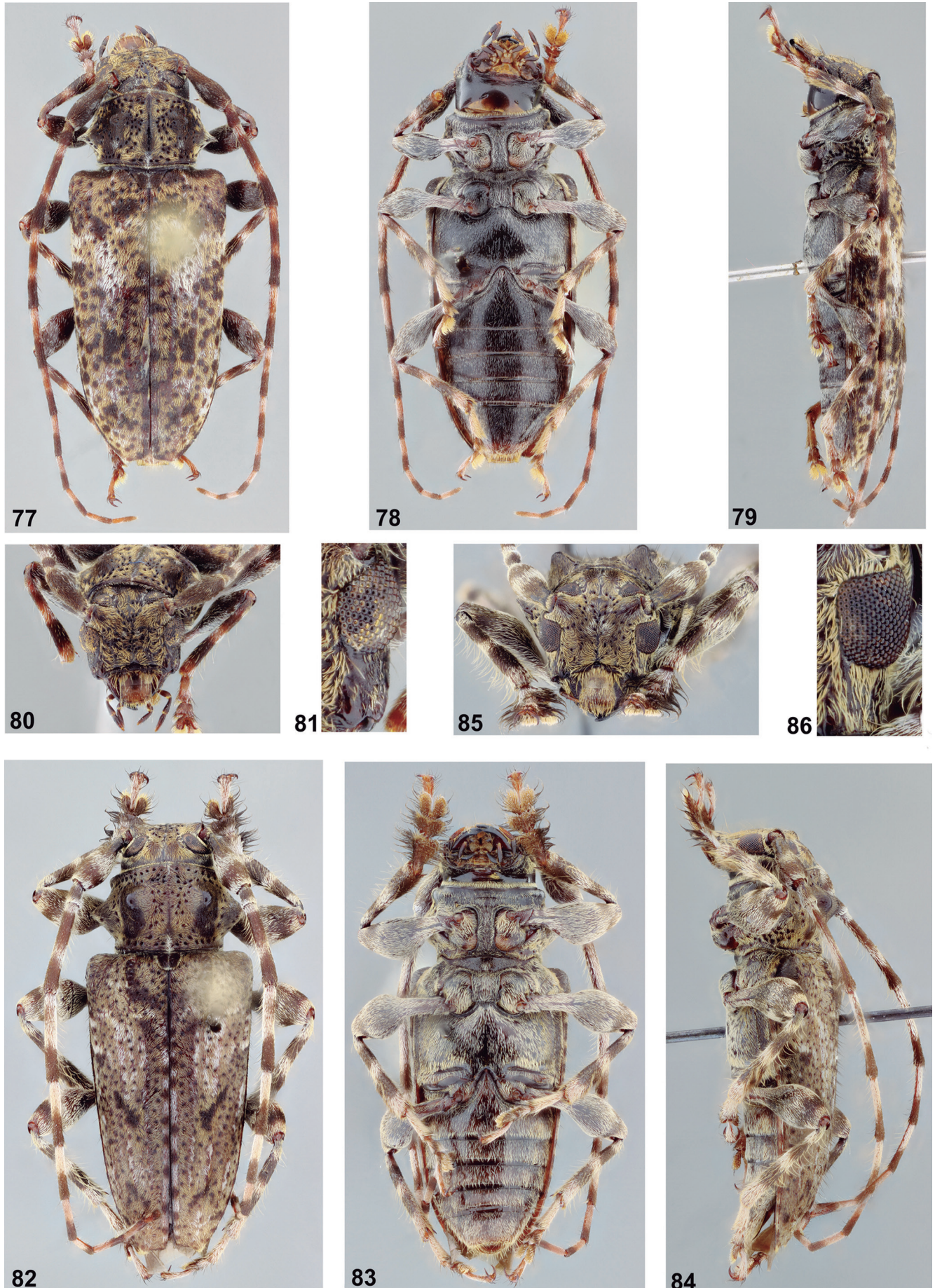
Head: Frons dense white pubescent with narrow, longitudinal, central glabrous band close to clypeus; with a few long, erect dark setae close to lower eye lobes. Area between antennal tubercles and upper eye lobes, and behind upper eye lobes with dense white pubescence, except moderately narrow, longitudinal, glabrous central band between antennal tubercles and upper eye lobes, orange pubescence interspersed in area adjacent to inner side of upper eye lobes, orange pubescent macula behind beginning of the upper eye lobes, and oblique orange pubescent band each side of median groove between antennal tubercles and upper eye lobes, and triangular glabrous indent on posterior central area; area of vertex and behind upper eye lobes close to prothorax glabrous; with a few long, erect dark setae close to eyes. Area behind lower eye lobes (this area widened toward ventral surface) with dense white pubescence close to wide superior area of eye, with orange pubescence interspersed, orange pubescence on inferior area close to eye, with white pubescence interspersed. Genae slightly shorter than twice length of lower eye lobe; with dense orange pubescence close to eye and white pubescence interspersed, distinctly sparser toward glabrous apex. Postclypeus with dense, long, decumbent white pubescence on wide central area, glabrous laterally; with sparse, long, erect dark setae on pubescent area. Posterior $\frac{2}{3}$ of labrum coplanar with anteclypeus, inclined at anterior third; with dense, long, decumbent white pubescence on posterior $\frac{2}{3}$, sparser, shorter on anterior third; with fringe of pale-yellow setae in anterior margin; with sparse, long, erect dark setae in central area of posterior $\frac{2}{3}$. Gulamentum glabrous except narrow anterior area with yellowish-white pubescence not obscuring integument. Distance between upper eye lobes 0.9 times length of scape; in frontal view, distance between lower eye lobes 1.2 times length of scape. Antennae 1.25 times elytral length, reaching posterior fifth of elytra. Scape clavate, longitudinally sulcate dorsally in basal half; with yellowish brown pubescence partially obscuring integument dorsally and laterally, with white pubescence interspersed, except white distal pubescent ring, which surrounds the entire circumference of the scape; ventral surface with white pubescence not obscuring integument; with a few long, erect dark setae ventrally. Pedicel with abundant white pubescence, except yellowish-brown pubescence on center of dorsal and lateral surfaces; with a few long, erect dark setae ventrally. Antennomere III with abundant white pubescence on wide anterior area, except sparsely pubescent on center of dorsal and lateral surfaces of this area; remaining surface with dark pubescence partially obscuring integument; anterior $\frac{3}{4}$ of ventral surface with sparse, long, erect dark setae, becoming noticeable denser, forming

distinct tuft on posterior quarter. Antennomere IV with anterior $\frac{2}{3}$ densely white pubescent, dark on remaining surface; with long, erect dark setae on ventral surface of posterior third, forming distinct tuft in posterior quarter of antennomere (shorter than in III). Antennomere V with dense white pubescence on basal half, dark on distal half; ventral third of surface with moderately short and abundant, erect dark setae. Remaining antennomeres with basal white pubescent ring, and dark pubescence on remaining surface. Antennal formula (ratio) based on length of antennomere III: scape = 0.75; pedicel = 0.25; IV = 0.75; V = 0.42; VI = 0.30; VII = 0.25; VIII = 0.20; IX = 0.17; X = 0.15; XI = 0.20.

Thorax: Lateral tubercle of prothorax large, conical, slightly curved upward, with blunt apex. Pronotum with large, elevated tubercle on each side, with top truncate and somewhat bifid; with large tubercle centrally, from posterior sulcus to near anterior margin, triangular-shaped posteriorly, carina-shaped anteriorly; slightly well-marked centrally, more distinct laterally; with coarse, sparse punctures, forming transverse row in posterior sulcus; with white and orange pubescence intermixed, denser laterally, with white pubescence prevailing in some areas, orange in others, apex of lateral pronotal tubercles, and parts of central tubercle; with a few long, erect dark setae posteriorly. Sides of prothorax with dense white pubescence, with orange pubescence interspersed on some areas. Prosternum with white pubescence, denser laterally, with orange pubescence interspersed on area under procoxal cavities. Prosternal process about as wide as procoxal cavity, coarsely rugose-punctate, longitudinally sulcate centrally, with apex strongly emarginate; with abundant white pubescence not obscuring integument. Ventral surface of mesothorax with abundant yellowish-white pubescence centrally, not obscuring integument, dense, white pubescence, obscuring integument laterally, except large area of mesanepisternum with orange pubescence interspersed. Mesoventral process slightly wider than mesocoxal cavity, with distinctly, elevated tubercle each side of anterior area; with white pubescence nearly obscuring integument, top of lateral tubercles glabrous. Ventral surface of metathorax with dense yellowish-white pubescence, with orange pubescence interspersed in some areas of sides. Scutellum with white pubescence centrally, orange on sides, margins with white and orange pubescence intermixed. **Elytra:** Humeri projected slightly forward; with elevated centrobasal crest between humeri and scutellum, covered with small tubercles, and distinct carina from apex of centrobasal crest to near apex; with another slightly distinct basal crest, also between humeri and scutellum but placed more laterally than the former crest, covered with small tubercles (sparser than in the other crest), followed by another carina, fused with the innermost near apex; humeral area with small, sparse tubercles; apex truncated with outer angle triangularly projected; with sparse, erect dark setae throughout; pubescence dense, mostly white with orange pubescent maculae and bands interspersed, except seven, notice-



Figures 67-76. (67-71) *Scythropopsis barrerai*, female. (67) Dorsal habitus. (68) Ventral habitus. (69) Lateral habitus. (70) Head, frontal view. (71) Lower eye lobe. (72-76) *Scythropopsis intricata* sp. nov. holotype female. (72) Dorsal habitus. (73) Ventral habitus. (74) Lateral habitus. (75) Head, frontal view. (76) Lower eye lobe.



Figures 77-86. (77-81) *Aegomorphus ramirezi*, male. (77) Dorsal habitus. (78) Ventral habitus. (79) Lateral habitus. (80) Head, frontal view. (81) Lower eye lobe. (82-86) *Aegomorphus maccartyi*, male. (82) Dorsal habitus. (83) Ventral habitus. (84) Lateral habitus. (85) Head, frontal view. (86) Lower eye lobe.

able dark-brown pubescent areas: one small, oblique, placed centrally on anterior third; one before middle laterally; one arched before middle close to suture; one zig-zag, large, placed about middle, not reaching suture; one inverted V-shaped dorsally on posterior fifth; another oblique, placed laterally on posterior fifth; sparse small maculae along posterior half of suture and posterior margin. **Legs:** Femora with dense white pubescence on peduncle and base of club, with white and orange pubescence intermixed on remaining surface. Tibiae with three yellowish-white pubescent rings, one basally, one centrally, another narrower, less conspicuous on apex; remaining surface with dark-brown pubescence; entire surface with short, sparse orange setae interspersed. Tarsomeres I and V mostly with white pubescence dorsally, and remaining segments with brownish pubescence not obscuring integument.

Abdomen: Ventrites I-IV with yellowish-white pubescence laterally, gradually whiter, sparser toward central area; ventrite V mostly with white pubescence, distinctly sparser on some large areas, with sparse, long, erect dark setae interspersed posteriorly; apex of ventrite V truncate.

Dimensions (mm): Total length, 12.9; prothoracic length, 2.7; anterior prothoracic width, 4.1; posterior prothoracic width, 3.9; maximum prothoracic width, 5.3; humeral width, 6.1; elytral length, 9.0.

Type material: Holotype female from MEXICO, Chiapas: Lago de Colores, 12.V.1969, J.M. Campbell col. (MZSP).

Remarks: *Scythropopsis intricata* has a feature of *Tetrasarus*: tuft of setae on ventral surface of the antennomeres III and IV. However, the tuft of setae is much denser, and the antennomere IV is distinctly longer in species of *Tetrasarus*. Notwithstanding, it could be compared with *Tetrasarus inops* Bates, 1880 (see photograph of the lectotype at Bezark, 2019), by the similar elytral pubescence pattern, from which it differs, besides the two features pointed out before, by the proportions between elytra and head + prothorax (elytra distinctly shorter in *S. intricata*). In *Scythropopsis*, *S. intricata* is most similar to *S. sallei*, but differs by the tuft of setae on ventral surface of the antennomeres III and IV, which is absent in females of *S. sallei*.

Etymology: The name of this new species: "intricata" is in reference to the intricate pubescent pattern of the elytra.

***Scythropopsis lugens* (Thomson, 1865), comb. nov.
(Fig. 100)**

Psapharochrus lugens Thomson, 1865: 543; Lacordaire, 1872: 543; Thomson, 1878: 15 (type); Monné, 2019: 300 (cat.).

Acanthoderes lugens; Gemminger, 1873: 3146 (cat.); Bates, 1880: 140, 1885: 378; Blackwelder, 1946: 611 (checklist); Chemsak et al., 1992: 130 (checklist); Noguera & Chemsak, 1996: 406 (cat.).

Acanthoderes (*Psapharochrus*) *lugens*; Aurivillius, 1923: 386 (cat.); Gilmour, 1965: 613 (cat.); Monné & Giesbert, 1994: 231 (checklist); Monné, 1994: 64 (cat.). *Aegomorphus lugens*; Chemsak & Noguera, 1995: 99; Monné, 2005: 166 (cat.).

Bates (1880) transferred *Psapharochrus lugens* to *Acanthoderes* but did not provide any explanation. Chemsak & Noguera (1995) transferred *Acanthoderes lugens* to *Aegomorphus* also without explanation. However, Noguera & Chemsak (1996) listed the species as *Acanthoderes lugens*. Probably this latter work was already in press when the transference to *Aegomorphus* was done. Nevertheless, examination of photograph of the holotype shows that the eyes are finely granulated, and the elytra have distinct carina. Accordingly, the species is transferred to *Scythropopsis*. Monné (2019), without any explanation, listed the species as *Psapharochrus lugens*.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Mexico (Mexico, Puebla, Veracruz).

***Aegomorphus* Haldeman, 1847**

Psapharochrus Thomson, 1864: 18. Syn. nov.

***Aegomorphus wappesi* (Galileo, Martins & Santos-Silva, 2015), comb. nov.**

Psapharochrus wappesi Galileo et al., 2015: 87; Wappes & Arias, 2016: 8 (holotype).

Psapharochrus jameswappesi Tavakilian, 2018: 40; Monné, 2019: 296 (cat.). Syn. nov.

As indicated previously, Tavakilian (2018) transferred *Acanthoderes wappesi* Chemsak & Hovore, 2002 to *Psapharochrus*. Accordingly, this created a secondary homonymy with *Psapharochrus wappesi* Galileo et al., 2002. Tavakilian (2018) provided a new name for the younger homonymy: *P. jameswappesi*. However, with the transference of *Acanthoderes wappesi* to *Scythropopsis*, *Psapharochrus wappesi* Galileo et al., 2002 is reinstated as valid, and the former becomes its synonym (ICZN, 1999: Article 59.4).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Bolivia (Tarija).

***Aegomorphus contaminatus* (Thomson, 1865),
revalidation, comb. nov.
(Fig. 40)**

Psapharochrus contaminatus Thomson, 1865: 543, 1878: 15 (type).

Acanthoderes contaminata; Gemminger, 1873: 3145 (cat.); Blackwelder, 1946: 610 (checklist); Zajciw, 1969a: 197 (distr.), 1970b: 187 (distr.).

Acanthoderes (Psapharochrus) contaminatus; Aurivillius, 1923: 386 (cat.).

Acanthoderes (Psapharochrus) contaminata; Gilmour, 1965: 615 (cat.).

Acanthoderes melanostictus; Monné & Giesbert, 1992: 252 (syn.).

Even based on examination of the photograph of the holotype (Fig. 37) we were not able to place this species as a synonym (senior or junior) of another species currently included in *Psapharochrus*, *Acanthoderes* (including the subgenera), or other genera with species similar in general appearance to it. Thus, the species is considered valid and remains known only from the type locality indicated in the original description (Brazil). The original description and photograph of the holotype is not diagnostic enough to know whether the species really belongs to *Aegomorphus*. However, we doubt that the species is from Brazil and, most likely belongs in *Scythropopsis*. For now, until more information or specimens become available, it is transferred to *Aegomorphus* simply because of its original placement in *Psapharochrus*.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Brazil.

***Aegomorphus ramirezi* (Chemsak & Hovore, 2002),
comb. nov.
(Figs. 77-81)**

Acanthoderes ramirezi Chemsak & Hovore, 2002b: 5.

Acanthoderes (Acanthoderes) ramirezi; Monné, 2005: 162 (cat.); Monné & Hovore, 2006: 203 (checklist); Monné, 2019: 223 (cat.).

Examination of photograph of the holotype, as well as the specimens examined, indicates that the eyes are coarsely granulated. Accordingly, the species is transferred to *Aegomorphus*.

Material examined: MEXICO, *Sonora* (new state record): MX16 km 155, 1 male, 18.VII.2007, Skillman, Ribardo and Hildebrandt col. (FWSC). *Michoacán* (new state record): Hwy MX37, 98 km S Nueva Italia, 1 female, 13.VII.2006, F.W. Skillman and D.C. Hildebrandt col. (MZSP, formerly FWSC). *Jalisco*: MX200, 21 km N Melaque, 1 female, 06.VII.2006, F.W. Skillman & D.C. Hildebrandt col. (ACMT, formerly FWSC).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new records): Mexico (Sonora, Jalisco, Michoacán).

***Aegomorphus maccartyi* (Chemsak & Hovore, 2002),
comb. nov.
(Figs. 82-86)**

Acanthoderes maccartyi Chemsak & Hovore, 2002a: 30.

Acanthoderes (Acanthoderes) maccartyi; Monné, 2005: 162 (cat.); Monné & Hovore, 2006: 203 (checklist); Monné, 2019: 222 (cat.).

Examination of a photograph of the holotype, as well as available specimens, reveals that the eyes are coarsely granulated. Accordingly, the species is transferred to *Aegomorphus*.

Material examined: MEXICO, *Jalisco*: MX80, 20 km S Autlán (RMO Los Mazos), 1 male, 08.VII.2006, Skillman & Hildebrandt col. (FWSC); El Tuito, 1 female, 14.VII.1993, Morris, Huether & Wappes col. (RFMC); 6 km N El Tuito, 1 male, 15-16.VII.1993, Wappes col. (ACMT).

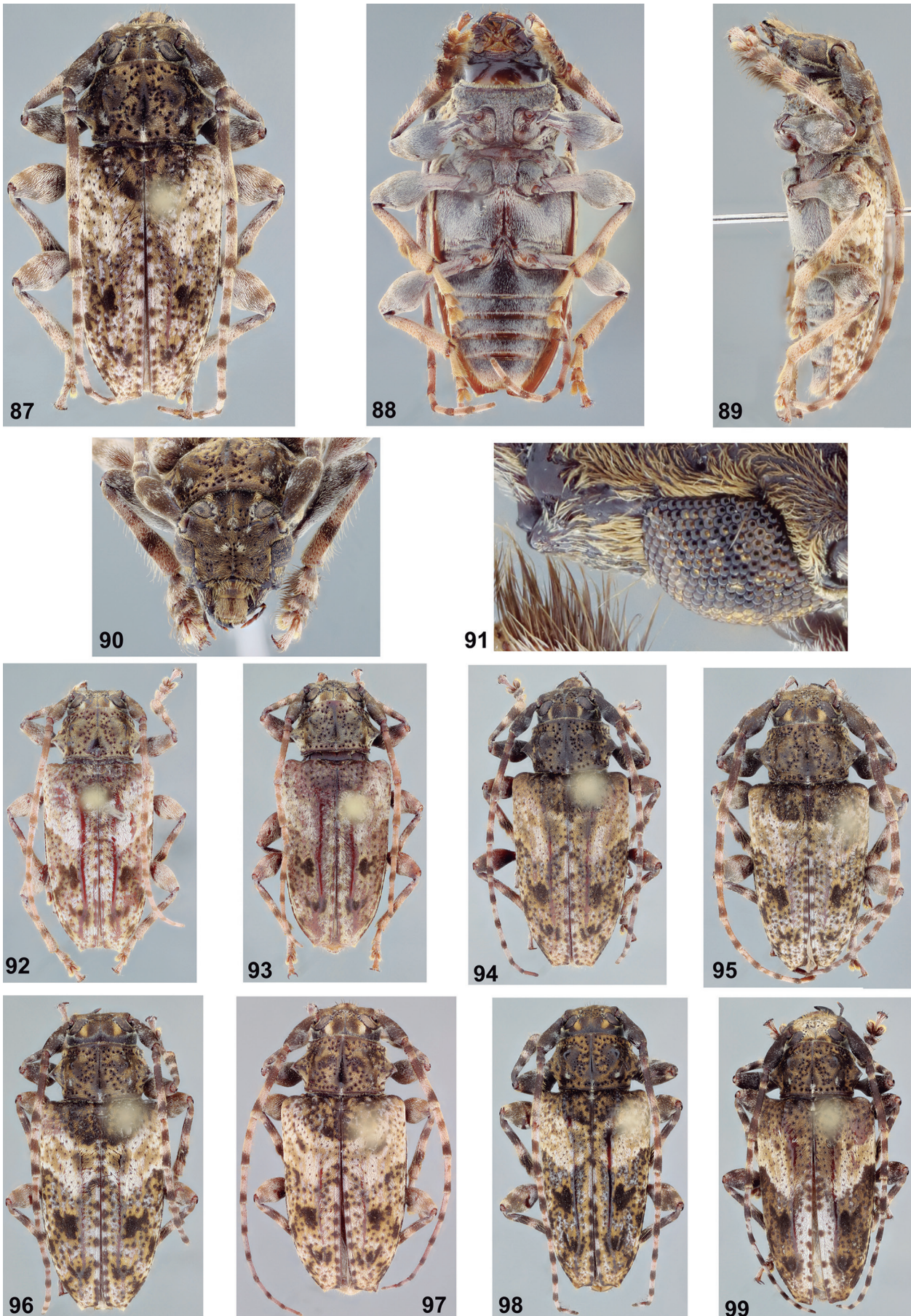
Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Mexico (Jalisco).

***Aegomorphus albosignus* Chemsak & Noguera, 1995
(Figs. 87-99)**

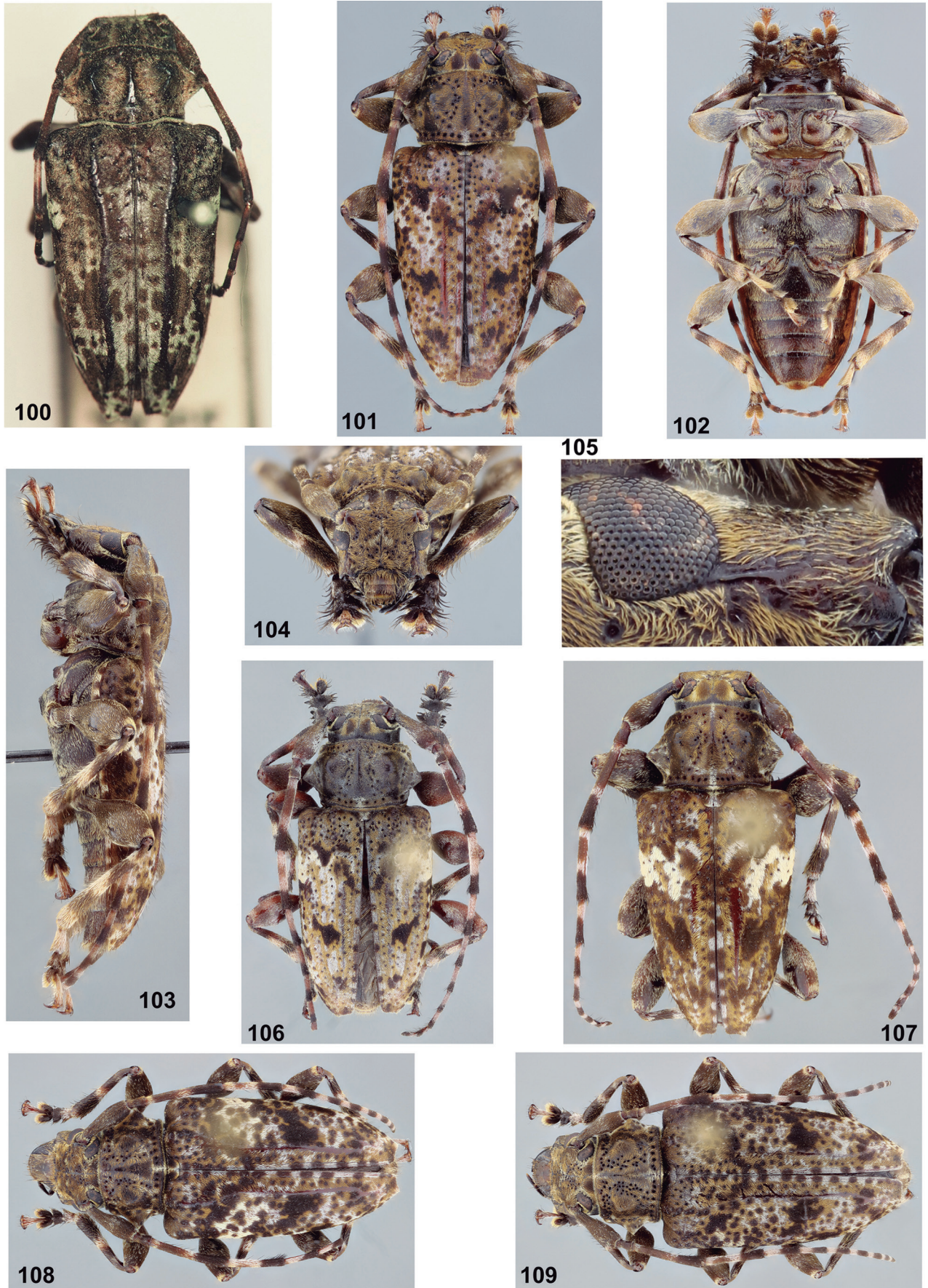
Aegomorphus albosignus Chemsak & Noguera, 1995: 98; Monné, 2001: 38 (cat. hosts); Noguera *et al.*, 2002: 625 (distr.); Monné, 2005: 165 (cat.); Monné & Hovore, 2006: 203 (checklist); Noguera *et al.*, 2009: 89 (distr.); Luna-León *et al.*, 2015: 838 (distr.); Martins *et al.*, 2015: 838 (key); Zaragoza-Caballero & Pérez-Hernández, 2017: 30 (holotype); Noguera *et al.*, 2017: 11 (distr.); Monné, 2019: 227 (cat.).

Aegomorphus albosignus was described from males and females from Mexico (Jalisco). According to Chemsak & Noguera (1995) "this species is readily recognizable by the pale pubescent face and the white oblique and longitudinal fasciae of the elytra" However, as in many other species of *Aegomorphus*, *Acanthoderes*, and *Scythropopsis* the elytral pubescent pattern, as well as the color of the pubescence of the frons are extremely variable. This is clearly shown in the type series (see photographs at Bezark, 2019) where the longitudinal white pubescent band on the elytra varies from being well defined, slightly distinct or even absent or nearly so (Fig. 92-98). Thus, it can be difficult to recognize the species, which is only possible by examining other details provided in the original description.

Material examined: MEXICO, *Michoacán* (new state record): MX 37, km 98, S Nueva Italia, 2 males, 1 female, 13.VII.2006, Skillman & Hildebrandt col. (FWSC); 2 males, 3 females, 15.VII.2006, Skillman & Hildebrandt col. (FWSC). *Colima* (new state record): Hwy MX98, km 33, N Manzanillo, 1 male, 11.VII.2006, Skillman & Hildebrandt col. (FWSC). *Jalisco*: MX200, 5 km N Campo Acosta, 6 males, 3 females, 21.VII.2006, Skillman & Hildebrandt col. (FWSC); MX200, at km marker 59, Chamela Bio Station, 1 female, 09.VII.2006, Skillman & Hildebrandt col. (FWSC); 21 km N Melaque, 1 male, 06.VII.2006, Skillman & Hildebrandt col. (FWSC). *Guerrero*: Hwy. GRO1, 1 km S Taxco El Viejo, 1 male, 26.X.2006, Skillman & Eya col. (FWSC). *Sonora*:



Figures 87-99. (87-91) *Aegomorphus albosignus*, male from Mexico (Jalisco). (87) Dorsal habitus. (88) Ventral habitus. (89) Lateral habitus. (90) Head, frontal view. (91) Lower eye lobe. (92-99) Specimens from Mexico, dorsal habitus. (92) Female from Jalisco. (93) Female from Jalisco. (94) Female from Michoacán. (95) Male from Michoacán. (96) Female from Michoacán. (97) Female from Michoacán. (98) Male from Guerrero. (99) Female from Michoacán.



Figures 100-109. (100) *Psapharochrus lugens*, holotype, dorsal habitus. (101-109) *Aegomorphus quadrigibbus*. (101) Male, specimen 1, from Mexico (Guerrero), dorsal habitus. (102) Male, specimen 1, from Mexico (Guerrero), ventral habitus. (103) Male, specimen 1, from Mexico (Guerrero), lateral habitus. (104) Male, specimen 1, from Mexico (Guerrero), head, frontal view. (105) Male, specimen 1, from Mexico (Guerrero), lower eye lobe. (106) Male, specimen 2, from Mexico (Guerrero), dorsal habitus. (107) Male, specimen 3, from Mexico (Veracruz). (108) Female, specimen 1, from Mexico (Guerrero). (109) Female, specimen 2, from Mexico (Guerrero).

MX16, km 155, 05.VII.2008, 1 male, Skillman, O'Brien & Ribardo col. (FWSC); MX16, km 155, 18.VII.2007, 2 males, Skillman, O'Brien, Ribardo & Hildebrandt col. (ACMT); MX16, km 155, 02.VII.2008, 1 male, Skillman, O'Brien & Ribardo col. (FWSC).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new records): Mexico (Jalisco, Morelos, Guerrero, Sonora, Oaxaca, Michoacán, Colima).

***Aegomorphus quadrigibbus* (Say, 1831)
(Figs. 101-109)**

Acanthocinus quadrigibbus Say, 1831: 9, 1835: 195; LeConte, 1859: 665.

Acanthoderes quadrigibbus; Haldeman, 1847: 45; LeConte, 1852: 175; Melsheimer, 1853: 107 (cat.); Bland, 1861: 97 (distr.); LeConte, 1873: 337, 1880: 237 (hosts); Packard, 1881: 55, 75, 131 (biol.); LeConte & Horn, 1883: 322; Shufeldt, 1884: 334, 338 (distr.); Townsend, 1884: 222; Harrington, 1884: 48 (hosts); Townsend, 1885: 70 (distr.); Chittenden, 1894: 99 (hosts); Knobel, 1895: 34; Hamilton, 1895: 339 (distr.); Leng & Hamilton, 1896: 114; Wickham, 1897: 206; Harrington, 1897: 74 (hosts); Smith, 1900: 294 (distr.); Dury, 1902: 162 (distr.); Ulke, 1903: 26 (distr.); Felt, 1906: 702, 715 (biol.); Smith, 1910: 333; Blatchley, 1910: 1070; Fisher & Kirk, 1912: 314 (distr.); Johnson, 1915: 315 (distr.); Nicolay, 1919: 70 (distr.); Craighead, 1923: 113 (larva); Turnbow & Franklin, 1980: 344 (distr.).

Psapharochrus quadrigibbus; Lacordaire, 1872: 751; Casey, 1913: 301; Champlain et al., 1925: 140 (hosts); Kirk & Knull, 1926: 42 (distr.); Leonard, 1928: 451 (distr.); Beaulne, 1932: 219 (hosts); Pechuman, 1937: 12 (biol.); Brimley, 1938: 217; Kaston, 1938: 239 (biol.); Hoffman, 1942: 11; Knull, 1944: 92 (distr.); Beal & Massey, 1945: 148 (biol.); Loding, 1945: 122 (distr.); Knull, 1946: 243; Fattig, 1947: 33 (distr.); Jaques, 1951: 267; Alexander, 1958: 49 (distr.); Solomon et al., 1976: 290; Monné, 2005: 211 (cat.).

Acanthoderes quadrigibba; Gemminger, 1873: 3146 (cat.); Horn, 1880: 115; Franz, 1954: 226 (distr.); Duffy, 1960: 214 (larva, biol.); Gilmour, 1965: 613 (cat.); Zajciw, 1969b: 609 (distr.); Perrault, 1978: 380 (distr.); Rice & Enns, 1981: 92 (distr., hosts); Chemsak & Hovore, 2002b: 12; Hernández-Fuentes et al., 2018: 544 (distr., host).

Acanthoderes (Acanthoderes) quadrigibba; Swift et al., 2010: 45 (distr.); Maes et al., 2010: 319 (distr.); Audureau, 2010: 8 (distr.); Holt, 2013: 252 (distr.); Klingeman et al., 2017: 298 (distr.); Audureau & Roguet, 2018: 76 (distr.); Monné, 2019: 222 (cat.).

Acanthoderes 4-gibbus; Riley, 1880: 270 (hosts); Packard, 1890: 91, 221, 291, 520 (biol.); Caulfield, 1890: 66 (hosts); Beutenmuller, 1896: 78 (hosts).

Aegomorphus quadrigibbus; Linsley & Chemsak, 1985: 246 (syn.); Hovore et al., 1987: 316 (distr.); Chemsak et al.,

1992: 131 (checklist); Lingafelter & Horner, 1993: 183 (distr.); MacRae, 1993: 244 (distr.); Monné & Giesbert, 1994: 35 (checklist); Monné, 1994: 35 (cat.); Yanega, 1996: 133; Noguera & Chemsak, 1996: 406 (distr.); Linsley & Chemsak, 1997: 339 (hosts); Peck & Thomas, 1998: 122 (distr.); Schiefer, 1998: 125 (distr.); Monné, 2001: 39 (cat. hosts); Vlasak & Vlasakova, 2002: 214 (hosts).

Acanthoderes (Psapharochrus) quadrigibba; Aurivillius, 1923: 387 (cat.).

Psapharochrus quadrigibbus lucidus Knull, 1958: 282; Chemsak, 1977: 178 (type).

Acanthoderes (Psapharochrus) quadrigibba lucidus; Gilmour, 1965: 613 (cat.).

Although originally described in *Acanthocinus* by Say, 1831 and moved to *Acanthoderes* by Haldeman (1847) it was Lacordaire (1872) who finally transferred *Acanthoderes quadrigibbus* to *Psapharochrus*, affirming that this latter genus included most of the species allocated in *Acanthoderes* at that time. Curiously, the species continued to be quoted as being in *Acanthoderes*, and it was Casey (1913) who finally correctly listed it again in *Psapharochrus*. Even so, the species continued to be mentioned as belonging to *Acanthoderes*, until Aurivillius (1923) moved *Psapharochrus* to the status of a subgenus of *Acanthoderes* and included *A. quadrigibba* in it. From Aurivillius (1923) to Rice & Enns (1981) the species was mentioned in *Psapharochrus* (as a distinct genus) or in *Acanthoderes (Psapharochrus)*. More recently, Linsley & Chemsak (1985) transferred the species to *Aegomorphus*, but only because of their doubts: "Since the species of *Acanthoderes sensu lato* are extremely numerous the question of generic or subgeneric assignments is difficult and beyond the scope of this work... We are assigning our five species of 'Acanthoderes' to *Aegomorphus* Haldeman until the problem can be resolved for the entire group." Finally, Chemsak & Hovore (2002b) transferred *Aegomorphus quadrigibbus* to *Acanthoderes*. The thinking behind what these authors were trying to do is somewhat mysterious, since they reported: "We have used the name *Acanthoderes* herein for placement of taxa which possess most of the characters of *Acanthoderes varia* and *Aegomorphus decipiens*, and which fit within the existing parameters of the genus *sensu lato*, without assignment to subgenera." We do not know for sure what "*Acanthoderes varia*" is, but it is likely *Cerambyx varius* Fabricius, 1787 (currently equal to *Cerambyx clavipes* Schrank, 1781, and placed in *Aegomorphus*). As *Aegomorphus decipiens* is the type species of *Aegomorphus*, this affirmation may have indicated their view that it was closely related to *Acanthoderes*. For sure, it was a transference based on doubts, and not on justified or definable features.

Acanthocinus quadrigibbus Say, 1831 has the eyes coarsely faceted, and pronotum not distinctly depressed centrally. Accordingly, it belongs to and is hereby transferred to *Aegomorphus*.

As correctly reported by Chemsak & Hovore (2002b) for this species "the variation within and be-

tween samples is considerable." The elytral pubescence (Figs. 101, 106-109) is extremely variable in concentration, but much less so in position of the maculae, and although the anterior light pubescent macula of the elytra is from slightly distinct to well-marked its placement on the elytra remains the same.

Material examined: MEXICO, *Guerrero*: Hwy 200, 51 km NE Ixtapa, 6 males, 4 females, 18-21.VII.1985, Wappes col. (ACMT); 1 female, 17-20.VII.1985, Wappes col. (ACMT). *Quintana Roo* (new state record): Hwy 307, 5 km S Cancun A.P., 1 male, 09.VI.2005, Skillman Jr. col. (FWSC); 1-5 km S Cancun, 1 male, 04.VI.2009, Skillman & Hildebrant col. (FWSC). *Veracruz*: Laguna Catemaco, 1 male, 24-25.V.1969, J.E.H. Martin col. (MZSP); 1 male, 09.VI.1969, J.E.H. Martin col. (MZSP); 1 male, 25.VI.1969, Bright & Campbell col. (MZSP); 1 male, 08-16.VIII.1969, H.F. Howden col. (MZSP). GUATEMALA (new country record): *Baja Verapaz*: CA14, km 149-151, 1 male, 2 females, 25.VII.2008, Skillman and C. & L. O'Brien col. (FWSC).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new records): Canada, Eastern United States to Florida and Texas, Mexico (Chiapas, Guerrero, Jalisco, Yucatán, Nayarit, Quintana Roo), Guatemala, Nicaragua, Costa Rica.

***Aegomorphus circumflexus* (Jacquelin du Val, 1857),
nomen protectum, comb. nov.
(Figs. 110-114)**

Acanthocinus rusticus Klug, 1829: 13. *Nomen oblitum*.

Acanthoderus circumflexus Jacquelin DuVal, 1857: 270. *Nomen protectum*.

Acanthoderes circumflexus; Pascoe, 1866: 279 (distr.); Bates, 1872: 207, 1880: 140; Berry & Salazar-Vaquero, 1957: 15 (distr.); Gregoire, 1957: 21 (physiol.); Zajciw, 1963: 590, 1964: 160; Linsley & Chemsak, 1966: 242; Chemsak, 1969: 189 (distr.); Chemsak & Linsley, 1980: 310 (distr.).

Acanthoderes circumflexus; Pittier & Biolley, 1895: 28 (distr.).

Acanthoderes circumflexa; Terrón, 1997: 223 (distr.).

Acanthoderes circumflexa; Gemminger, 1873: 3145 (cat.); Gahan, 1895: 130; Leng & Mutchler, 1914: 450 (distr.); Martorell, 1939: 204 (distr.); Fisher, 1944: 10 (distr.); Blackwelder, 1946: 610 (checklist); Wolcott, 1948: 342; Zayas, 1975: 225; Chemsak *et al.*, 1980: 36 (distr.); Chemsak *et al.*, 1992: 130 (cat.); Maes *et al.*, 1994: 28 (distr.); Noguera & Chemsak, 1996: 406 (cat.); Maes, 1998: 922 (distr.); Chemsak & Hovore, 2002b: 10; Lozada Piña *et al.*, 2004: 106 (distr.); Peck, 2005: 177 (distr.); Fernández García *et al.*, 2009: 322 (distr.).

Acanthoderes (Psapharochrus) circumflexa; Aurivillius, 1923: 385 (cat.); Gilmour, 1965: 613 (cat.), 1968: 154 (distr.); Monné, 1994: 28 (cat.); Monné & Giesbert, 1994: 230 (checklist).

Psapharochrus circumflexus; Monné, 2001: 47 (cat. hosts), 2005: 202 (cat.); Hovore, 2006: 376 (distr.); Hubweber,

2008: 255 (distr.); Swift *et al.*, 2010: 46 (distr.); Maes *et al.*, 2010: 368 (distr.); Morales-Morales *et al.*, 2012: 38, 45 (distr.; biol.); Thomas *et al.*, 2013: 20 (distr.); García Morales *et al.*, 2014: 108 (distr.); Lanuza-Garay & Barrios, 2015: 68 (distr.); Audureau & Roguet, 2018: 77 (distr.); Monné, 2019: 292 (cat.).

Psapharochrus circumflexus; Chevrolat, 1862: 247; Gundlach, 1891: 205.

Psapharochrus circumflexa; Turnbow *et al.*, 2003: 28 (distr.).

Psapharochrus circumflexum; Audureau, 2008: 14 (distr.).
Acanthoderus meteorica Gistel, 1857: 79; Aurivillius, 1923: 385 (syn.).

Psapharochrus histrio Casey, 1913: 302; Turnbow *et al.*, 2003: 29 (distr.); Monné, 2005: 205 (cat.); Lingafelter *et al.*, 2014: 76 (holotype); Monné, 2019: 295 (cat.). Syn. nov.

Acanthoderes (Psapharochrus) histrio; Aurivillius, 1923: 386 (cat.); Gilmour, 1965: 614 (cat.); Monné, 1994: 61 (cat.); Monné & Giesbert, 1994: 230 (checklist).

Acanthoderes histrio; Blackwelder, 1946: 610 (checklist); Chemsak *et al.*, 1992: 130 (checklist).

Psapharochrus guatemalensis Casey, 1913: 303; Monné, 2005: 204 (cat.); Hovore, 2006: 376 (distr.); Lingafelter *et al.*, 2014: 73 (holotype); Monné, 2019: 295 (cat.). Syn. nov.

Acanthoderes (Psapharochrus) guatemalensis; Aurivillius, 1923: 386 (cat.); Gilmour, 1965: 614 (cat.); Monné, 1994: 61 (cat.); Monné & Giesbert, 1994: 230 (checklist).

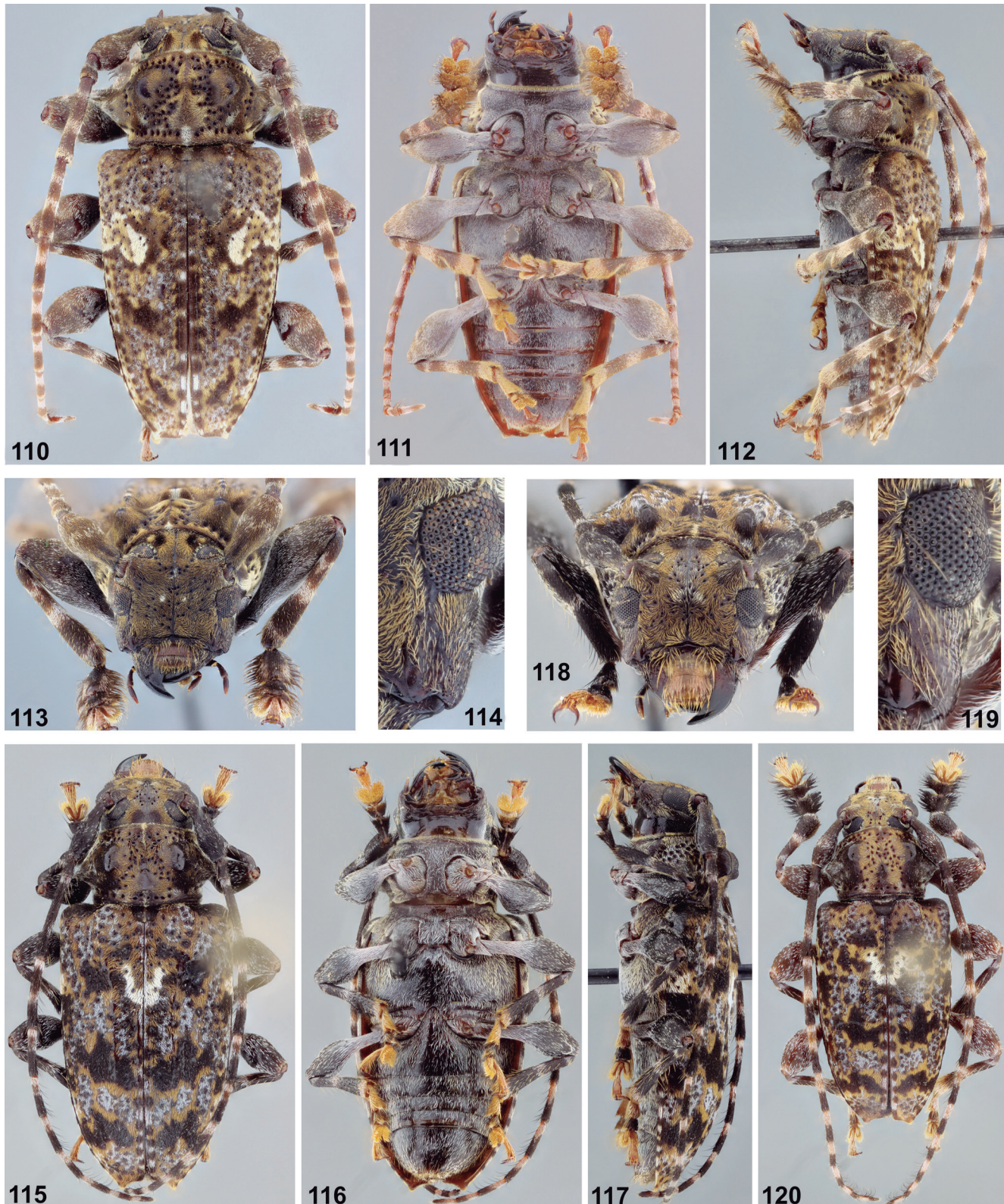
Acanthoderes guatemalensis; Blackwelder, 1946: 610 (checklist); Chemsak *et al.*, 1992: 130 (checklist).

According to Jacquelin du Val (1857): "*Acanthoderus circumflexus*, Klug"; and "*Acanthoderus rusticus*, Dej., Cat. Col., 3^e édit., p. 362." There are some mistakes in those statements: 1. The genus was reported as "*Acanthoderus*" instead to *Acanthoderes*; 2. Klug (1829) described the species as "*Acanthocinus rusticus*: 198. *Acanthocinus rusticus* Dej. (i. litt.) fuscus, nigro-punctatus, coleoptris fascia transversa dentata interrupta grisea. (Minor A. depresso)."

According to Dejean (1835, 1836), "*Acanthoderus. Serville*" is equal to *Acanthocinus*. "*Merzele. Dej. Catal.*"; and "*Rusticus. Dej. Cuba*" is equal to *Circumflexus*. "*Klug. id.*" As there is no species formally described by Johann Christoph Friedrich Klug as "*circumflexus*" in Cerambycidae, it seems evident that Jacquelin du Val (1857) attributed the species to him based on information by Dejean (1835, 1836), indicating ("*Acanthoderus rusticus* equal to "*Acanthoderus circumflexus* Klug). Thus, Klug (1829) described the species as *Acanthocinus rusticus* (with a description provided, making the description taxonomically technically available (ICZN, 1999)). Accordingly, *Acanthoderes circumflexus* Jacquelin du Val, 1857 is a junior synonymy of *Acanthocinus rusticus* Klug, 1829 (= *Psapharochrus rusticus* (Klug, 1829)). However, both the conditions demanded by ICZN (1999: Article 23.9.1.1 and 23.9.1.2) to preserve a commonly used name are met, *Acanthoderus circumflexus* Jacquelin Du Val, 1857 is hereby valid and a *nomen protectum* (ICZN,

1999: Article 23.9.2), and *Acanthocinus rusticus* Klug, 1829, is hereby a *nomen oblitum*. A list of works giving evidence of the validity of *Acanthoderus circumflexus* by use of its name as a valid taxon is given in the ICZN reference above.

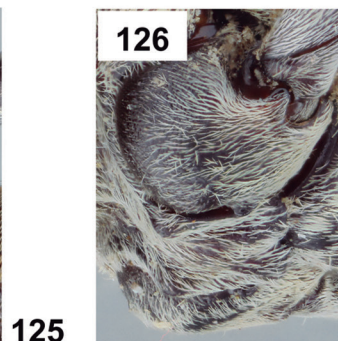
Material examined: MEXICO, *Jalisco* (new state record): MX200, 21 km N Melaque, 1 male, 06.VII.2006, Skillman & Hildebrand col. (FWSC). *Oaxaca*: MX190, 11 km SE Zanatepec, 1 male, 20.X.2001, Skillman & Davidson col. (FWSC); Finca San Carlos (30 km E Palomares), 1 female,



Figures 110-120. (110-114) *Aegomorphus circumflexus*, male from Cuba. (110) Dorsal habitus. (111) Ventral habitus. (112) Lateral habitus. (113) Head, frontal view. (114) Lower eye lobe. (115-120) *Aegomorphus irumus*. (115) Dorsal habitus, female. (116) Ventral habitus, female. (117) Lateral habitus, female. (118) Head, frontal view, female. (119) Lower eye lobe, female. (120) Dorsal habitus, male.

29.V.1959, G. Halftter col. (MZSP). *Michoacán* (new state record): Tuxpan, 1 male, VIII.1960, G. Halftter col. (MZSP). *Morelos* (new state record): Cuernavaca, 1 female, no date and collector indicated (MZSP). *Distrito Federal*

(new state record): 1 male, 23.V.1956, G. Halftter col. (MZSP). *Veracruz*: Playa de San Vicente, 1 male, II.1910, C.C. Halfmann col. (MZSP). *Puebla* (new state record): Villa Juarez (La Ceiba), 1 male, X.IV.1953, L. Vesguez col.



Figures 121-129. (121-125) *Aegomorphus consentaneus*, male from Bolivia. (121) Dorsal habitus. (122) Ventral habitus. (123) Lateral habitus. (124) Head, frontal view. (125) Lower eye lobe. (126-127) *Aegomorphus satellinus*, mesoventral process, male from Bolivia. (128-129) *Aegomorphus jaspideus*, mesoventral process, male from Bolivia.

(MZSP). *Quintana Roo*: 1 female, IV.1967, no collector indicated (MZSP); Palenque, 1 male, 07-09.V.1969, Bright & Campbell col. (MZSP). GUATEMALA, *Escluinta*: El Salto, 1 male, 1934, F.A. Bianchi col. (MZSP). BELIZE (new country record): *Toledo*: Punta Gorda, 1 male, no date and collector indicated (MZSP). EL SALVADOR (new country record): *San Salvador*: San Salvador, 1 female, XI.1960, collector illegible (MZSP). COSTA RICA, *Guanacaste*: Canas, 1 male, 1 female, 10.V.1991, D.B. Thomas col. (ACMT). *Cartago*: Turrialba, 1 female, 13.III.1970, no collector indicated (MZSP). *Limón*: near Cahuita town (09.6827°N, 82.8105°W), 1 male, VI.2015, A. Kozlov col. (MZSP). PANAMA, *Panama*: Barro Colorado Island, 1 male, 05-08.VIII.1965, B. Malkin & S. Rand col. (MZSP); 1 female, XI.1965, H. Britski (MZSP). CUBA, *Corralillo* [there are places with this name in more than one province], 3 males, 9 females, VI.1951, Zayas col. (MZSP); 1 male, VI.1958, Zayas col. (MZSP). *Sierra del Rosario* [this place extends into two provinces: Pinar del Rio, and Artemisa], 1 female, 07.VII.1933, A. Bierig col. (MZSP); 1 male, 15.X.1933, A. Bierig col. (MZSP). *Portugalete* [there are places with this name in more than one province], 1 female, 17.II.1929, A. Bierig col. (MZSP). *La Habana*: Habana, 1 female, V.1927, A. Bierig col. (MZSP); 1 male, VI.1928, A. Bierig col. (MZSP); 1 female, IV.1932, A. Bierig col. (MZSP). *Artemisa*: Aspiro (Sierra del Rosario), 1 male, 06.V.1934, A. Bierig col. (MZSP); 3 males, 1 female, 29.VI.1934, A. Bierig col. (MZSP). COLOMBIA, *Bolívar*: Zambrano (Hacienda Monterey, 09°45'N, 74°49'W, 10 m), 1 female, 09.IX.1993, F. Fernandez col. (MZSP). VENEZUELA, *Distrito Capital*: Caracas, 1 female, 15.VII.1959, Bordon col. (MZSP); 1 female, 10.VII.1960, Bordon col. (MZSP).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019; new records): Mexico (San Luis Potosí, Veracruz, Oaxaca, Chiapas, Yucatán, Tamaulipas, Quintana Roo, Jalisco, Michoacán, Morelos, Puebla, Distrito Federal), Guatemala, Honduras, El Salvador, Belize, Nicaragua, Costa Rica, Panama, Cuba, Puerto Rico, Cayman Islands, Curaçao, Colombia, Venezuela.

***Aegomorphus irumus* (Galileo & Martins, 2011),
comb. nov.
(Figs. 115-120)**

Psapharochrus irumus Galileo & Martins, 2011: 178;
Wappes & Arias, 2016: 8 (holotype); Monné, 2019: 296
(cat.).

Description: Female (Figs. 115-119): Integument mostly black; mouthparts reddish-brown, except maxillary palpomeres II-IV, and labial palpomeres II-III black with reddish-brown apex; anteclypeus, labrum, protarsomeres III-V, meso- and metatarsomeres I and III-V reddish-brown; antennomere IV with basal and medial dark reddish-brown rings; remaining antennomeres dark reddish-brown on basal half; apex of meso- and metatibiae yellowish-brown.

Head: Frons coarsely, sparsely punctate; with yellowish-brown pubescence partially obscuring integument, white pubescence centrally, between lower eye lobes and antennal tubercles; with a few long, erect dark setae (apex yellowish) close to lower eye lobes. Area between antennal tubercles and upper eye lobes with central diamond-shaped area, coarsely, sparsely punctate, with sparse yellowish-white pubescence with yellowish-brown pubescence interspersed, sides of posterior area with denser yellowish-brown pubescence; sides extending toward prothorax with dense yellowish-brown pubescent band, narrow nearly glabrous area close to eyes. Central area of vertex close to prothorax with one large semielliptical area with brown pubescence on each side of median groove. Area behind eyes with yellowish-brown pubescence close to eyes (this area widened toward ventral surface), with white pubescence interspersed, glabrous toward prothorax. Genae about as long as lower eye lobe; with sparse light yellowish-brown pubescence, sparser toward apex. Postclypeus centrally and laterally glabrous, with bristly yellowish-brown pubescence on remaining surface, with long setae of same color interspersed, and a few long, erect dark setae. Posterior $\frac{3}{4}$ of labrum coplanar with anteclypeus, inclined at anterior quarter; with short yellowish-white pubescence close to anteclypeus, gradually longer toward apex of coplanar area; anterior margin with fringe of nearly golden setae. Gulamentum smooth, wide posterior area glabrous, depressed, anteriorly with short yellowish-brown pubescence not obscuring integument. Distance between upper eye lobes 0.58 times length of scape; in frontal view, distance between lower eye lobes equal to length of scape. Antennae 1.6 times elytral length, reaching elytral apex at posterior quarter of antennomere VIII. Scape clavate, not sulcate dorsally; dark-brown pubescent with white pubescence interspersed, white pubescence more abundant ventrally, somewhat yellowish-brown on base of ventral surface; with long, erect, sparse yellowish setae ventrally. Pedicel dark-brown pubescent with white pubescence interspersed, especially ventrally; ventral surface with long, sparse, erect yellowish setae. Antennomeres III-IV with basal and almost central white pubescent rings (denser on IV); remaining surface with dark-brown pubescence; ventral surface with long, sparse, erect yellowish setae. Remaining antennomeres with white pubescence on light area, dark-brown pubescence on dark area; with long, sparse, erect dark setae ventrally (yellowish on anterior area of V). Antennal formula (ratio) based on length of antennomere III (only one female measured): scape = 0.72; pedicel = 0.22; IV = 0.80; V = 0.56; VI = 0.48; VII = 0.46; VIII = 0.38; IX = 0.36; X = 0.28; XI = 0.26.

Thorax: Lateral conical tubercle of prothorax large, slightly inclined upward, with apex acute. Pronotum with large, elevated tubercle on each side, with top of tubercle truncate and somewhat bifid; a smaller, somewhat conical central tubercle at posterior half; coarsely, moderately abundantly punctate between tubercles, and between anterior margin and lateral tubercles, coarser

along posterior sulcus; punctures sparser, shallower on lateral tubercles of prothorax; tubercles glabrous on top; central area with yellowish-brown pubescence not obscuring integument, except narrow, longitudinal pale yellow-pubescent band anteriorly and posteriorly, and white pubescence on sides of posterior quarter; area between side of lateral tubercles and on lateral tubercles of prothorax with brown, yellowish-brown, and whitish pubescence intermixed. Sides of prothorax coarsely, moderately abundantly punctate; with white pubescence not obscuring integument, more yellowish anteriorly and posteriorly. Prosternum centrally with white pubescence not obscuring integument, more yellowish laterally. Prosternal process with narrowest area about as wide as half the width of procoxal cavity; with white pubescence not obscuring integument. Ventral surface of meso- and metathorax with pale-yellow pubescence laterally, white centrally (sparser on center of mesoventrite). Mesoventral process slightly narrower than width of mesocoxal cavity; lateral margins elevated, especially anteriorly. Scutellum with dark brown pubescence, except narrow, central, longitudinal yellowish pubescent band (sometimes absent). **Elytra:** Humeri very slightly projected forward; with centrobasal crest between humeri and scutellum elevated, covered with small tubercles; with slightly distinct carina from apex of centrobasal crest to posterior third, and another between the crest and humeri; coarsely, sparsely punctate; apex obliquely truncate, with outer angle projected; irregular areas with white pubescence (not obscuring punctures) surrounded by yellowish-brown pubescence, except dense, white U-shaped pubescent macula centrally on anterior third (connecting across both elytra), and three large, somewhat transverse, irregular areas with dark-brown pubescence (one on anterior third; one about middle; another on posterior third). **Legs:** Femora with dense white pubescence on peduncle, sparser on club. Tibiae with three white pubescent rings, one near base, one about middle, another at apex; remaining surface with brown pubescence not obscuring integument, with long, sparse, erect yellowish setae interspersed, especially in meso- and metatibiae.

Abdomen: Ventrites with white pubescence not obscuring integument; apex of ventrite V narrow, truncate.

Variation: Elytral pubescence is noticeably variable as in several species of *Aegomorphus*. Extremes could easily be confused with different species. In some specimens (including the holotype), the sides of anterior half have dense white pubescence forming large band distinctly attaining center of dorsal area, in others this band is restricted to inclined area, and in some it is nearly absent. The same occurs in the fascia along the sides in posterior half.

Dimensions (mm), male (2)/female (2): Total length, 10.55-10.70/10.60-10.95; prothoracic length, 2.05-2.10/1.90-2.00; anterior prothoracic width, 2.65-2.80/2.60-2.80; posterior prothoracic width, 2.55-2.65/2.35-2.70; maximum prothoracic width,

3.40-3.70/3.35-3.65; humeral width, 3.90-4.10/3.90-4.30; elytral length, 6.85-6.90/6.85-7.25.

Material examined: BOLIVIA, *Santa Cruz*: 4-6 km SSE Buena Vista (F&F Hotel), 1 male, 22-31.X.2002, Wappes col. (ACMT); 1 male, 26-27.X.2014, Wappes col. (ACMT); 4-5 km N Achira, road to Amboro, 1 female, 12-13.X.2000, Wappes & Dozier col. (MZSP); Huaico (14°40'S, 63°24'W; 430 m), 1 female, 21.XI.2013, Skillman & Wappes col. (FWSC).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Bolivia (*Santa Cruz*).

Remarks: The coarsely granulated eyes and the pronotum lacking a central depression places this species in *Aegomorphus*.

***Aegomorphus consentaneus* (Thomson, 1865),
comb. nov.
(Figs. 121-125)**

Psapharochrus consentaneus Thomson, 1865: 544, 1878: 15 (type); Monné, 2001: 47 (cat. hosts); Monné, 2005: 2034 (cat.); Monné, 2019: 293 (cat.).

Acanthoderes consentanea; Gemminger, 1873: 3145 (cat.); Lameere, 1884: 95 (distr.); Melzer, 1932: 223; Schottfeldt, 1944: 111 (hosts); Blackwelder, 1946: 610 (checklist); Zajciw, 1966: 9; Silva *et al.*, 1968: 397 (hosts); Zajciw, 1969a: 198 (distr.), 1970b: 187.

Acanthoderes (Psapharochrus) consentanea; Aurivillius, 1923: 386 (cat.); Gilmour, 1965: 615 (cat.); Monné & Giesbert, 1994: 230 (checklist); Monné, 1994: 60 (cat.).

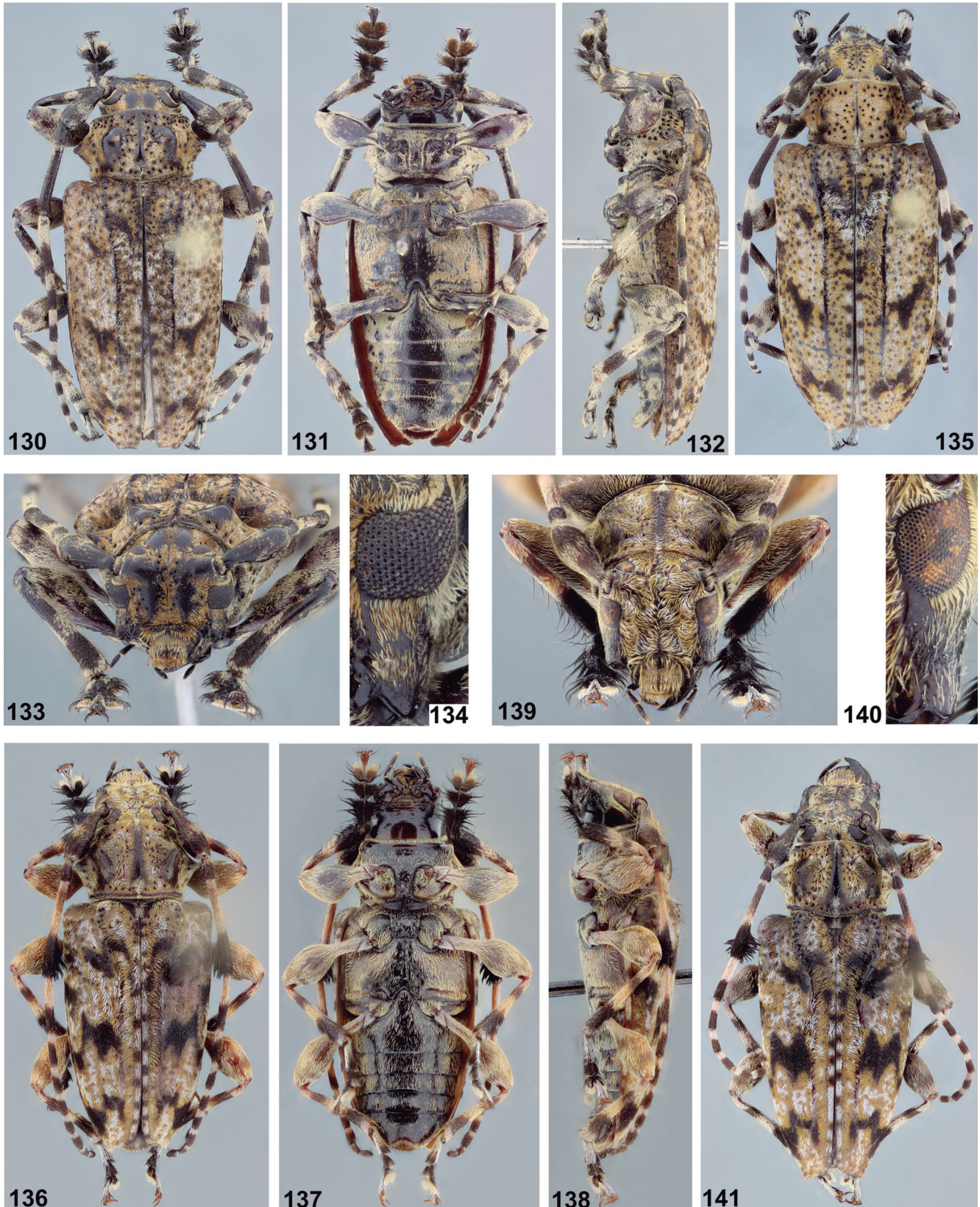
Psapharochrus consentaneus was described by Thomson based on a single specimen from Brazil without a specific locality given. Lameere (1884) reported the species from "Env. De Rio-Janeiro.", referring to a place near to Rio de Janeiro city (Rio de Janeiro, Brazil) and more recently Zajciw (1969a) recorded the species to two Brazilian states (Minas Gerais and Rio de Janeiro). The MZSP collection has a large number of Cerambycidae from southern Brazil, but all specimens of *P. consentaneus* are only from Northern Brazil and Bolivia. Accordingly, it is possible that the specimens examined by Lameere (1884) and Zajciw (1969a) are not really *P. consentaneus*.

The eyes coarsely granulate, and the pronotum lacking a central depression place this species in *Aegomorphus*. One of the primary features allowing recognition of the species is the sparse white setae on the large dark pubescent area along the sides of the posterior half of the elytra. Only one other species of *Aegomorphus* has this pattern of pubescence: *Aegomorphus pseudosatellinus* (Tavakilian & Néouze, 2013).

Material examined: BRASIL, *Amazonas* (new state record): Benjamin Constant (Rio Javari), 1 female, XI.1961, formerly Dirings collection (MZSP); 1 female, X.1962, formerly Dirings collection (MZSP); 1 male, X.1963, formerly Dirings collection (MZSP). *Pará* (new state record): 1 male, 1 female,

no date and collector indicated (MZSP); Santarém, 1 female, VIII.1920, Hagmann col. (MZSP); (Fazenda Taperinha), 1 male, 01-11.II.1968, no collector indicated (MZSP); Tucuui (Rio Tocantins), 1 male, 03.VII.1984, W. Overall

(MZSP). BOLIVIA (new country record): Cochabamba: Chaparé (400 m), 3 females, XI.1957, formerly Dirings collection (MZSP); Villa Tunari, 1 male, XI.1953, Prosen col. (MZSP). Santa Cruz: 4-6 km SSE Buena Vista (F&F Hotel),



Figures 130-141. (130-135) *Aegomorphus robustus* sp. nov. (130) Dorsal habitus, holotype male. (131) Ventral habitus, holotype male. (132) Lateral habitus, holotype male. (133) Head, frontal view, holotype male. (134) Lower eye lobe, holotype male. (135) Dorsal habitus, paratype female. (136-141) *Plagiosarus literatus*. (136) Dorsal habitus, male. (137) Ventral habitus, male. (138) Lateral habitus, male. (139) Head, frontal view, male. (140) Lower eye lobe, male. (141) Dorsal habitus, female.

1 male, 02-12.II.2000, J.E. Wappes col. (ACMT); 2 male, 1 female, 23-25.X.2000, R. Morris col. (RFMC); Reserva Natural Potrerillo del Guenda (Sake farm, 17°40.26'S, 63°27.43'W, 400 m), 1 male, 06-09.X.2006, Wappes, Nearn & Eya col.

(ACMT); Potrerillo del Guenda (400 m), 1 female, 06-08.XII.2011, Morris & Wappes col. (RFMC); (Reserva Natural, 40 km NW Santa Cruz, 17°40'S, 63°27'W, 370 m), 1 female, 30.VII-03.X.2007, Wappes & Morris col. (ACMT).



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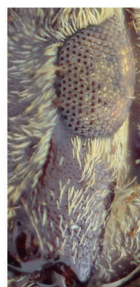
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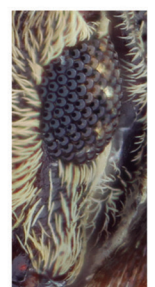
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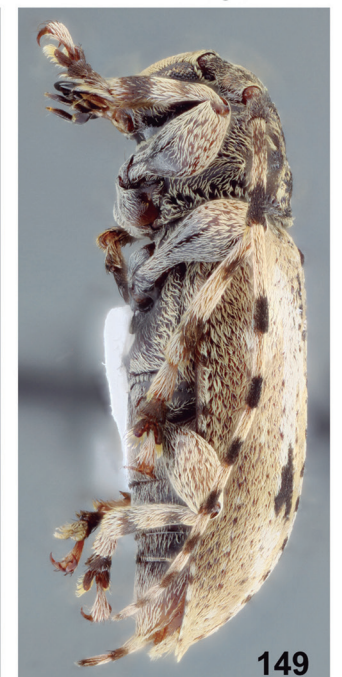
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Figures 142-151. (142-146) *Eupromerella quadrituberculata*, male. (142) Dorsal habitus. (143) Ventral habitus. (144) Lateral habitus. (145) Head, frontal view. (146) Lower eye lobe. (147-151) *Eupromerella boliviana* holotype female. (147) Dorsal habitus. (148) Ventral habitus. (149) Lateral habitus. (150) Head, frontal view. (151) Lower eye lobe.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Brazil (Amazonas, Pará, Minas Gerais, Rio de Janeiro), Bolivia.

***Aegomorphus satellinus* (Erichson, 1847), comb. nov.**
(Figs. 126-127)

Acanthoderes satellinus Erichson, 1847: 143; White, 1855: 357.

Acanthoderes satellina; Gemminger, 1873: 3147 (cat.); Blackwelder, 1946: 611 (checklist).

Psapharochrus satellinus; Lacordaire, 1872: 751; Monné, 2005: 212 (cat.); Wappes et al., 2006: 33 (distr.); Monné et al., 2012: 33 (distr.); Wappes et al., 2013: 10 (distr.); Monné, 2019: 305 (cat.).

Acanthoderes (Psapharochrus) satellina; Aurivillius, 1923: 388 (cat.); Gilmour, 1965: 614 (cat.); Monné & Giesbert, 1994: 231 (checklist); Monné, 1994: 68 (cat.).

The general appearance of *Aegomorphus satellinus* is similar to that of *Aegomorphus jaspideus* (Germar, 1823) making it difficult to separate them simply from the dorsal view. Fortunately, the mesoventral process (Figs. 126-127) differs in the two species with *A. satellinus* having a somewhat centrally flattened process with a small tubercle on each side of anterior margin. In *A. jaspideus*, the mesoventral process (Figs. 128-129) is centrally tumid and lacks the tubercles.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Peru, Bolivia (Cochabamba, Santa Cruz), Brazil (Mato Grosso, Goiás).

***Aegomorphus robustus* sp. nov.**
(Figs. 130-135)

Description: Male (Figs. 130-134): Integument mostly black; mouthparts dark reddish-brown, except palpomeres black with apex yellowish; anteclypeus semi-translucent; apex of labrum light reddish-brown.

Head: Frons coarsely, sparsely punctate; with orangish-brown pubescence, partially obscuring integument (worn away in the holotype and many of the paratypes), with white pubescence interspersed and a few long, erect brown setae close to eyes. Area between antennal tubercles coarsely, sparsely punctate; with orange-brown pubescence partially obscuring integument, with yellowish-white pubescence interspersed, central area with yellowish-white pubescence. Area between upper eye lobes with orange pubescence (much of it missing in the holotype), with white pubescence interspersed. Central area of vertex close to prothorax with a large semielliptical area with short brown pubescence on each side of median groove (lost in the holotype), and orangish-brown pubescence along median groove. Area behind upper eye lobes with orange-white pubescence, with distinct yellowish-white pubescence close to ver-

tex and dense, narrow yellowish-white pubescent band close to eye, widened toward inferior side, remaining surface glabrous. Genae almost 1.5 times length of lower eye lobe; with yellowish-white pubescence close to eye toward posterior area, with orangish-brown pubescence and white pubescence interspersed toward anterior area, glabrous apex. Postclypeus coarsely, sparsely punctate on wide central area, smooth laterally; with bristly orangish-brown pubescence on wide central area (nearly glabrous centrally), glabrous laterally; with long, sparse, erect dark setae on wide central area. Posterior $\frac{3}{4}$ of labrum coplanar with anteclypeus, inclined at anterior quarter; yellowish-white pubescent, with long, erect dark setae interspersed on coplanar area, anterior margin with fringe of nearly golden pubescence. Gulamentum with transverse, slightly distinct striae on wide posterior area, depressed on narrow anterior area; wide posterior area glabrous, with yellowish-white pubescence on depressed area. Distance between upper eye lobes 0.56 times length of scape; in frontal view, distance between lower eye lobes 0.95 times length of scape. Antennae 1.35 times elytral length, reaching elytral apex at middle of antennomere XI. Scape clavate, slightly sulcate in basal third dorsally; with yellowish-white pubescence (partially lost in the holotype), with orangish-brown pubescence interspersed dorsally. Pedicel with basal and distal pubescent rings, distally distinctly narrower, pubescence mostly pale-yellow dorsally, whiter ventrally. Antennomere III with pale-yellow pubescence on basal quarter of dorsal surface, pubescence whiter on basal half of ventral surface; remaining surface with brown pubescence, with a few white setae interspersed; with a few short, erect dark setae ventrally; remaining antennomeres pale-yellow pubescent on basal half, brown on distal surface; with a few short, erect dark setae on ventral surface of antennomeres IV-X (sparser toward X). Antennal formula (ratio) based on length of antennomere III (only one male measured): scape = 0.77; pedicel = 0.20; IV = 0.72; V = 0.52; VI = 0.44; VII = 0.38; VIII = 0.33; IX = 0.29; X = 0.24; XI = 0.25.

Thorax: Lateral tubercle of prothorax large, conical, slightly inclined upward, with apex blunt. Pronotum with large, elevated tubercle on each side, gradually more elevated anteriorly, becoming nearly conical with apex rounded; with carina-shaped central tubercle, from anterior margin to posterior sulcus, widened posteriorly; coarsely, sparsely punctate around tubercles and anteriorly and posteriorly; with orangish-brown and yellowish-white pubescence intermixed centrally, top of tubercles glabrous, and orangish-brown pubescence with yellowish-white pubescence interspersed (yellowish-white pubescence denser on sides of posterior area and sides of anterior area of lateral tubercles); with a few long, erect dark setae laterally on posterior area. Sides of prothorax coarsely punctate; with yellowish-white pubescence; with orangish-brown pubescence interspersed on area close to pronotum. Ventral surface of thorax with pale-yellow pubescence in some areas partially obscuring integument, distinctly sparser on cen-

tral area of mesoventrite. Prosternal process 0.7 times as wide as procoxal cavity; lateral margins slightly sinuous. Mesoventral process about as wide as mesocoxal cavity; longitudinally tumid centrally (more elevated toward anterior area), without tubercle on sides of anterior area. Scutellum with orangish-brown pubescence laterally (partially lost in the holotype), yellowish-white centrally. **Elytra:** Humeri slightly projected forward; with slightly elevated centrobasal crest between humeri and scutellum, covered with small tubercles, and distinct carina from apex of centrobasal crest to near apex; with another carina between humeri and the former carina; both carina fused distally; with small, sparse tubercles on basal quarter; coarsely, sparsely punctate; apex truncate, concave centrally; with yellowish-white and orangish-brown pubescence irregularly mixed, except: white V-shaped band centrally on anterior third (encompassing both elytra); irregular area with brown pubescence between centrobasal crests (pubescence lost in the holotype and most paratypes); oblique brown pubescent band on sides of anterior third dorsally; oblique, large, irregular brown pubescent band dorsally after middle; oblique, moderately large, irregular brown pubescent band dorsally near apex; with pubescent band along suture, from V-shaped pubescent band to apex, with rounded brown pubescent spots interspersed. **Legs:** Femora with pale-yellow pubescence on some areas partially obscuring integument, forming two dense, longitudinal bands ventrally on base of profemora. Tibiae with three pale-yellow pubescent rings, one basally, one centrally, another at apex; remaining surface with brownish pubescence not obscuring integument. Sides of dorsal surface of protarsomere I and tarsomere V with pale-yellow pubescence; remaining surface of protarsomere I and protarsomeres II-IV with dark pubescence. Meso- and metatarsi with pale-yellow pubescence dorsally.

Abdomen: Ventrites with dense pale-yellow pubescence laterally, sparser centrally, except glabrous, subrounded macula on each side, near apex in I-IV, near base in V; apex of ventrite V moderately wide, truncate, centrally emarginate.

Female (Fig. 135): Differs from male by the shorter antennae (1.2 times elytral length, reaching about distal ninth of elytra), and apex of ventrite V narrower.

Variation: Area between upper eye lobes coarsely, sparsely punctate; pubescence on dorsal surface of scape mostly orangish-brown; elytral apex not centrally concave, with outer angle slightly, triangularly projected.

Dimensions (mm), holotype/paratypemales/paratype females: Total length, 17.80/14.95-18.75/16.75-20.20; prothoracic length, 3.30/2.80-3.40/2.95-3.20; anterior prothoracic width, 4.60/3.80-4.75/4.25-5.00; posterior prothoracic width, 4.70/3.95-4.80/4.40-5.10; maximum prothoracic width, 6.10/5.30-6.30/5.70-6.25; humeral width, 7.25/6.20-7.50/7.05-8.05; elytral length, 12.70/10.30-12.95/12.10-14.05.

Type material: Holotype male from BOLIVIA, *Santa Cruz:* Andrés Ibáñez (Jardin Botanico), 02-03.XII.2010, J.L. Castro R. Flores col. (FSCA), formerly ACMT). Paratypes – BOLIVIA, *Santa Cruz:* same data as holotype, 2 male, 2 females (MNKM; 1 male, 1 female, ACMT); 4 males, 2 females, same data as holotype except, 26.27.XI.2010, J.L. Castro M. Perez col. (MNKM; 1 male, ACMT; 1 male, 1 female, MZSP); 20 km N Camiri (Road to Eyti; 1250 m; 6-8 km E Hwy 9; 19°52'S, 63°29'W), 1 female, 05-10.XII.2012, Wappes, Bonaso & Skillman col. (ACMT); (6-8 km E Hwy 9; 1250 m; 19°52'S, 63°29'W), 1 female, 26.XI.2013, Wappes & Skillman col. (ACMT); 4 km N Bermejo (Refugio los Volcanes; 1045-1350 m; 18°06'S, 63°36'W), 1 male, 17-24.X.2014, Wappes & Morris col. (ACMT).

Remarks: *Aegomorphus robustus* sp. nov. differs from other species of the genus with a similar appearance (e.g., *A. circumflexus*) by the mesoventral process distinctly longitudinally tumid centrally, and by lacking anterolateral tubercles (similar species have the mesoventral process flattened or somewhat depressed centrally, and with distinct anterolateral tubercles).

Etymology: This species name “robustus” simply references its robust appearance.

Taxonomic notes in *Eupromerella* Fisher, 1938; *Miriochrus* Galileo & Martins, 2012; and *Plagiosaurus* Bates, 1880.

***Plagiosaurus* Bates, 1880**

***Plagiosarus literatus* Bates, 1885
(Figs. 136-141)**

Plagiosarus literatus Bates, 1885: 382; Aurivillius, 1923: 383 (cat.); Blackwelder, 1946: 610 (checklist); Gilmour, 1965: 611 (cat.); Chemsak *et al.*, 1992: 132 (cat.); Monné, 1994: 51 (cat.); Monné & Giesbert, 1994: 237 (checklist); Monné, 2005: 198 (cat.); Hovore, 2006: 376 (distr.); Monné, 2019: 287 (cat.).

Description: Male (Figs. 136-140): Integument mostly black; mouthparts dark reddish-brown except palpomeres black with yellow apex; posterior area of gula-mentum dark reddish-brown; anteclypeus, labrum, most of tarsomeres V reddish-brown; basal half of antennomeres III-IV orangish-brown; basal third of antennomeres V-VIII reddish-brown; femora mostly brown with irregular reddish-brown areas; protibiae with four rings, from base to apex their color is: reddish-brown, dark-brown, reddish-brown, and then black; meso- and metatibiae dark-brown brown with three reddish-brown rings, one basally, one centrally (widest), another at apex (narrowest).

Head: Frons finely, sparsely punctate; with white, pale-yellow and yellowish-brown pubescence intermixed; with a few long, erect dark setae close to eyes. Vertex with punctures as on frons, nearly absent toward

prothoracic margin; pubescence as on frons between antennal tubercles and upper eye lobes, forming semi-circular yellowish-brown macula on each side of median groove close to prothorax. Area behind eyes finely punctate (punctures finer than on frons), denser close to eye, sparser toward prothorax; area behind upper eye lobes with yellowish-brown pubescence; area behind lower eye lobes with pale-yellow pubescence close to eye, with white setae interspersed, wide area close to prothorax glabrous. Genae 1.3 times longer than lower eye lobe; with dense pale-yellow pubescence close to eye, area close to frons glabrous, with sparse pale-yellow pubescence toward glabrous apex, with white setae interspersed. Postclypeus centrally and laterally glabrous, with bristly pale-yellow and yellowish-brown pubescence on remaining surface; with one long dark seta on each side of pubescent area. Posterior $\frac{2}{3}$ of labrum coplanar with anteclypeus, inclined at anterior fifth; with pale-yellow pubescence on coplanar area not obscuring integument, inclined area nearly glabrous, distal margin with golden pubescent fringe; with long dark setae directed forward at interface of coplanar and inclined area. Gula mentum smooth, wide posterior area glabrous, depressed, with short yellowish-white setae not obscuring integument anteriorly. Distance between upper eye lobes 0.51 times length of scape; in frontal view, distance between lower eye lobes 0.87 times length of scape. Antennae 1.55 times elytral length, reaching elytral apex at basal third of antennomere IX. Scape clavate, slightly sulcate dorsally at basal third; with dark reddish-brown and brown pubescence intermixed, central pubescent ring with dense, yellowish-brown on outer side, yellowish-white, sparse on remaining surface (sparser dorsally), and narrow pubescent area at apex. Basal ring of pedicel with pale-yellow pubescence, dark-brown pubescence on remaining surface. Antennomere III with yellowish-white pubescence from base to after middle, not obscuring integument, with black pubescence on remaining surface, forming dense tuft of long setae ventrally and on inferior surface of sides; with a few long, erect brownish setae ventrally on light area. Antennomeres IV-X with basal white pubescent ring, and black pubescence on remaining surface; antennomeres IX-XI with erect black setae on distal half of ventral surface. Antennal formula (ratio) based on length of antennomere III (only one female measured): scape = 0.62; pedicel = 0.24; IV = 0.54; V = 0.38; VI = 0.30; VII = 0.25; VIII = 0.22; IX = 0.21; X = 0.12; XI = 0.18.

Thorax: Lateral tubercles of prothorax large, conical, slightly inclined upward, with apex acute. Pronotum with large, elevated tubercle each side of central area, inclined sideways, with apex blunt; central area with elongate tubercle, from anterior margin to posterior sulcus, carina-shaped from anterior margin to near middle, gradually widened from this point to posterior sulcus; coarsely, sparsely punctate between tubercles, anteriorly and posteriorly; central area with yellowish-brown pubescence, except short yellowish-white pubescent band on central tubercle close to anterior margin, and another more irregular one close to posterior margin, six white pubescent

irregular macula, one each side of anterior quarter, one each side of posterior third, and one each side of posterior quarter; remaining anterior surface of central tubercle glabrous, and posterior area with brown pubescence not obscuring integument; apex of lateral tubercles glabrous; with dark-brown pubescent band close to outer side of lateral tubercles, and remaining dorsal surface of lateral tubercles on prothorax with yellowish-brown, pale-yellow and white pubescence intermixed; with a few long, erect dark setae laterally in posterior area. Sides of prothorax coarsely, sparsely punctate; with yellowish-brown pubescence and white pubescence interspersed. Prosternum with moderately dense yellowish-brown pubescence laterally, gradually whiter, distinctly sparser centrally, anterior area glabrous. Prosternal process slightly wider than half width of procoxal cavity; with sparse white pubescence. Ventral surface of mesothorax with dense abundant yellowish-brown pubescence laterally, white, notably sparse on central area of mesoventrite, white and sparse on mesoventral process. Mesoventral process slightly wider than width of mesocoxal cavity; with small tubercle each side of anterior area. Ventral surface of metathorax with abundant yellowish-brown pubescence laterally, whiter, sparser centrally. Scutellum with yellowish-white pubescence centrally, dark brown laterally. **Elytra:** Humeri very slightly projected forward; centrobasal crest short, elevated, covered with small tubercles; with slightly distinct carina from apex of centrobasal crest to posterior quarter, and another between the first and the humeri; apex truncate, with outer angle triangularly projected; coarsely, sparsely punctate on basal quarter, gradually finer, sparser toward apex; with abundant yellowish-brown pubescence with irregular white maculae interspersed, and three black pubescent dorsal areas, one at anterior third, one about middle, another at posterior quarter; with white pubescent band along suture, with small black pubescent maculae interspersed; with long, erect, sparse dark setae on basal quarter, especially on centrobasal crest. **Legs:** Femora with abundant yellowish-brown and pale-yellow pubescence intermixed, not obscuring integument. Protibiae with pale-yellow pubescence on anterior $\frac{2}{3}$ not obscuring integument, black on distal third; with sparse, long, erect black setae interspersed. Meso- and metatibiae with pale-yellow pubescence on light areas (partially white on distal light ring), dark brown on dark areas. Protarsomeres I-IV with black pubescence, white pubescence interspersed on sides of I; protarsomere V mostly with white pubescence. Meso- and metatarsomeres I and most of V with white pubescence; tarsomeres II and IV with black pubescence; tarsomeres III, partially with black pubescence and white pubescence interspersed.

Abdomen: Ventrites I-IV with abundant yellowish-brown pubescence laterally, gradually whiter, sparser centrally; ventrite V with sparse yellowish-brown pubescence, slightly whiter centrally; ventrite V with apex concave.

Dimensions (mm), male (1)/female (2): Total length, 8.90/9.70-12.10; prothoracic length, 1.70/1.75-2.10; ante-

rior prothoracic width, 2.10/2.30-2.80; posterior prothoracic width, 2.20/2.40-2.90; maximum prothoracic width, 2.85/3.25-3.80; humeral width, 3.40/3.70-4.50; elytral length, 5.65/6.40-7.70.

Material examined: MEXICO, Veracruz: Los Tuxtlas (UNAM), 1 male, 05-06.V.1994, J.E. Wappes col. (ACMT); 15 km W Sontecomapan, 2 females, 10-13.IV.1993, J.E. Wappes col. (ACMT; MZSP).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Mexico (Veracruz), Guatemala.

Remarks: *Plagiosarus literatus* was originally described based on a single female from Guatemala with Monné & Giesbert (1994) subsequently listing the species from Mexico (Veracruz). Recently, the species has only been mentioned in catalogs and checklists and until now the male had never been described.

Eupromerella Fisher, 1938

Eupromerella inaequalis (Galileo & Martins, 2011), comb. nov.

Psapharochrus inaequalis Galileo & Martins, 2011: 177; Wappes & Arias, 2016: 8 (holotype); Monné, 2019: 296 (cat.).

Galileo & Martins (2011) described *Psapharochrus inaequalis* based on a single male from Bolivia. According to Martins & Santos-Silva (2013): "The main feature that allows the separation of *Eupromerella* from *Psapharochrus* is the shorter body when the proportion between the head and prothorax is compared with the elytra. No other reliable character can be used to distinguish these genera. The main problem is that there are currently several different forms placed in *Psapharochrus*. However, this does not prevent the transfer of species from *Psapharochrus* to *Eupromerella* and does not classify it as a doubtful genus."

Psapharochrus inaequalis is nearly identical to *Eupromerella orbifera* (Aurivillius, 1908). The latter species was originally described in *Psapharochrus* and transferred to *Eupromerella* Fisher, 1938 by Monné & Giesbert (1994). In reality, *E. orbifera* is a problematic species that could be included in either *Psapharochrus* (now *Aegomorphus*) or *Eupromerella*. Also, it is probable that *P. inaequalis* may only be a variation of *E. orbifera*, a species that also occurs in Bolivia. For now, it will be kept as a different species because the apex of the protibiae is entirely dark (only apex in the holotype and specimens examined), and its elytral pubescence slightly sparser (but with same pattern). It will be necessary to examine the holotype before a synonymy could be considered.

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Bolivia (Santa Cruz).

Eupromerella quadrituberculata (Zajciw, 1964), comb. nov. (Figs. 142-146)

Acanthoderes quadrituberculata Zajciw, 1964: 160, 1969a: 198 (distr.), 1974: 67 (distr.); Julio *et al.*, 2000: 31 (holotype).

Acanthoderes (Psapharochrus) quadrituberculata; Zajciw, 1969b: 608; Monné, 1994: 67 (cat.); Monné & Giesbert, 1994: 231 (checklist).

Psapharochrus quadrituberculatus; Monné, 2005: 212 (cat.); Monné & Monné, 2016: 57 (holotype); Monné, 2019: 304 (cat.).

The eyes of *Acanthoderes quadrituberculata* (Fig. 146) are not distinctly finely granulated hence its inclusion in this genus is incorrect. Furthermore, its general shape indicates that it is much better placed in *Eupromerella* than in *Aegomorphus* (see comments in *Eupromerella inaequalis*).

Unfortunately, both the holotype and all paratypes were destroyed during the 2018 fire in the MNRJ.

Material examined: BRAZIL, Espírito Santo: Parque Sooretama, 2 males, 27.XI.1967, F. Oliveira col. (MZSP).

Known geographical distribution (Monné, 2019; Tavakilian & Chevillotte, 2019): Brazil (Espírito Santo).

Eupromerella boliviana sp. nov. (Figs. 147-151)

Description: Female: Integument mostly black; mouthparts dark reddish-brown, except palpi black with reddish-brown apex of last palpomeres; antennomere III brown on basal $\frac{2}{3}$, black on distal third (slightly reddish-brown on apical area); remaining antennomeres yellowish-brown on basal half/third, black on remaining surface; anteclypeus and labrum mostly testaceous.

Head: Frons finely, densely punctate; with yellowish-brown and pale-yellow pubescence intermixed, nearly obscuring integument, with yellowish-white setae interspersed, large yellowish-white pubescent macula between eyes and antennal tubercles centrally. Vertex finely, abundantly punctate; area between antennal tubercles with yellowish-white pubescence not obscuring integument centrally, with yellowish-white setae interspersed, and dense yellowish-white pubescence laterally; area between upper eye lobes and beginning of area behind upper eye lobes with yellowish-white pubescence not obscuring integument, with yellowish-brown setae interspersed, dense yellowish-brown pubescence on remaining surface, except brown pubescent macula between the two pubescent areas; area close to prothorax nearly glabrous; remaining surface area behind upper eye lobes close to eye with yellowish-brown and brown pubescence not obscuring integument, glabrous toward prothorax. Area behind lower eye lobes with sparse yellowish-white pubescence close to superior area of

eye, denser toward ventral surface, distinctly sparser toward prothorax. Genae about as long as lower eye lobe; with yellowish-brown pubescence, denser close to eye, sparser on remaining surface. Postclypeus with bristly yellowish-brown pubescence on wide central area not obscuring integument, central area nearly glabrous laterally glabrous. Posterior $\frac{3}{4}$ of labrum coplanar with anteclypeus, inclined on anterior quarter; with abundant yellowish-white pubescence on coplanar area, becoming yellowish toward anterior margin. Gulamentum glabrous except narrow anterior area with yellowish-white pubescence not obscuring integument. Distance between upper eye lobes 0.73 times length of scape; in frontal view, distance between lower eye lobes 1.17 times length of scape. Antennae 1.6 times elytral length, reaching elytral apex at basal third of antennomere IX. Scape nearly straight at outer surface, strongly, gradually widened from base to middle of inner surface, then slightly narrowed toward apex; with yellowish-brown and brown pubescence mixed dorsally, not obscuring integument, except dense, narrow yellowish pubescent macula on apex; with yellowish-white pubescence not obscuring integument on remaining surface. Pedicel brown pubescent dorsally, with yellowish-brown pubescence interspersed, especially basally; remaining surface with yellowish-white pubescence not obscuring integument. Light area of antennomeres with yellowish-white pubescence (sparser on some areas of III), and dark brown pubescence on dark area; ventral surface of antennomeres III-IV with short, sparse, erect yellowish-brown setae; apex of antennomeres V-X with short, erect, brownish setae at apex. Antennal formula (ratio) based on length of antennomere III: scape = 0.78; pedicel = 0.24; IV = 0.87; V = 0.58; VI = 0.51; VII = 0.42; VIII = 0.37; IX = 0.36; X = 0.31; XI = 0.33.

Thorax: Lateral tubercle of prothorax large, conical, slightly curved upward, with apex nearly acute. Pronotum with large gibbosity on each side of central area, and slightly elevated central gibbosity on posterior half, becoming carina-shaped on anterior half; coarsely, densely punctate, except smooth posterior area of central gibbosity; with irregular tufts of light yellowish-brown pubescence (more pale yellow posteriorly), and moderately sparse brown pubescence between them, except glabrous posterior area of central gibbosity; with a few long, erect dark setae on sides of posterior area. Sides of prothorax coarsely, abundantly punctate; with light yellowish-brown pubescence throughout. Ventral surface of thorax with moderately dense pale-yellow pubescence laterally, gradually sparser, whiter centrally. Narrowest area of prosternal process slightly less than half width of procoxal cavity. Apex of mesoventral process about as wide as mesocoxal cavity. Scutellum with short brownish pubescence not obscuring integument, with yellowish-brown setae interspersed. **Elytra:** Coarsely, abundantly punctate on basal third, punctures gradually sparser on remaining surface toward apex; apex obliquely truncate; with dense light yellowish-brown pubescence nearly entirely obscuring integument, except: irregular, narrow, fragmented

V-shaped (across both elytra) brownish pubescent area on center of basal third; small, irregular white pubescent spot on base of each arm of V-shaped area; large, irregular white pubescent macula dorsally on basal half; irregular, nearly triangular brown pubescent macula dorsally after middle, laterally margined with narrow, white pubescence, especially on outside edge; irregular, white pubescent macula dorsally on posterior quarter close to latter dark macula; small, irregular white pubescent spot near apex; narrow white pubescent macula along suture and epipleural margin (nearly absent on basal quarter of sutural area), with brown pubescent areas interspersed.

Legs: Femora with yellowish-white pubescence on peduncle and basal area of club, remaining surface densely pale-yellow. Tibiae with yellowish-white pubescence, except two brown pubescent macula covering dorsal area and sides, one less conspicuous on basal third, another wider on posterior half (more conspicuous in protibiae). Tarsomeres I and V with yellowish-white pubescence dorsally (sparser on protarsomere I); tarsomeres II-IV with brown pubescence dorsally.

Abdomen: Ventrites with yellowish-white pubescence not obscuring integument, slightly yellower and denser on V; ventrite V centrally sulcate at basal third; apex of ventrite V truncate.

Dimensions (mm), holotype female/paratype female:

Total length, 6.30/6.25; prothoracic length, 1.25/1.20; anterior prothoracic width, 1.80/1.80; posterior prothoracic width, 1.80/1.80; maximum prothoracic width, 2.40/2.30; humeral width, 2.85/2.80; elytral length, 4.50/4.35.

Type material: Holotype female from BOLIVIA, *Santa Cruz*: Potrerillo del Guendá (Reserva Natural, Snake Farm; 400 m; 17°40'15"S, 63°27'26"W), 23-30.X.2013, Wappes & Kuckartz col. (FSCA, formerly ACMT). Paratype female from BOLIVIA, *Santa Cruz*: Andrés Ibáñez (Potrerillo del Guendá; 370 m; 17°40'S, 63°27'W), 23-27.X.2007, S.W. Lingafelter col. (SWLC).

Remarks: *Eupromerella boliviana* sp. nov. is similar to *E. plaumanni* (Fuchs, 1959) but females differ as follows: size smaller; lower eye lobes proportionally smaller; femoral peduncle shorter; elytral pubescent pattern with posterior dark macula not arch-shaped; and metaventrite not punctate laterally. Females of *E. plaumanni* are larger, lower eye lobes are proportionally larger, femoral peduncle are longer, posterior dark macula of the elytra is arch-shaped, and the metaventrite is punctate laterally. The new species differs from *E. picturata* Martins, Galileo & Limeira-de-Oliveira, 2009 (females) by the smaller size, metaventrite not punctate laterally (punctate in *E. picturata*), the pubescence of the scape is sparser (denser in *E. picturata*), and posterior dark macula of the elytra not arch-shaped (arched in *E. picturata*). *Eupromerella boliviana* also differs from *E. propinqua* (Melzer, 1931), and *E. pseudopropinqua* (Fuchs, 1959), by the different pubescent pattern of the elytra (see photographs of the types of those two species at Bezark, 2019).

Etymology: This species is named “boliviana” after the country (Bolivia) where it is found.

***Miriochrus* Galileo & Martins, 2012**

Miriochrus Galileo & Martins, 2012: 66; Monné, 2019: 256 (cat.).

Miriochrus was described to include *M. minimus* Galileo & Martins, 2012 from Paraguay (see photograph of the holotype at Bezark, 2019). In the original description, the genus was compared to *Nesozineus* Linsley & Chemsak, 1996 (translated): “*Miriochrus* gen. nov. by the pronotum lacking tubercles, and by the small dimensions, resembles *Nesozineus* Linsley & Chemsak, 1966, but differs by the eye lobes nearly divided (one ommatidium in the narrowest area between the lobes), scape piriform, short elytra with 1.5 times the humeral width, and presence of white seta inside of the elytral punctures. In *Nesozineus*, the eye lobes are separated between them by three or four rows of ommatidia, the scape is subcylindrical, the elytra are longer, with more than 1.5 times humeral width, and the elytral punctures have no white seta.” Actually, *Miriochrus* is much more similar to *Eupromerella* Fisher, 1938, and primarily differs by the presence of white seta inside its elytral punctures, which are absent in *Eupromerella*. This feature is also shared with the *Plistonax* Thomson, 1864, and *Melzerus* Monné, 2005. The species of *Plistonax* are slenderer, and the apex of the prothoracic tubercles is blunt (acute in *Miriochrus*); *Melzerus* has a distinct cavity in the ventral surface of the femora of males, which are absent in males of *Miriochrus*, and the apex of the prothoracic tubercles is rounded.

The following list of Acanthoderini genera and species reflects the taxonomic changes made based on the results of our studies. It also indicates the type of change made, such as: syn. nov., comb. nov., nomen protectum, nomen oblitum, sp. nov. and revalidation or resurrection, as appropriate.

***Aegomorphus* Haldeman, 1847. American species.**

= *Psapharochrus* Thomson, 1864 syn. nov.
Aegomorphus albosignus Chemsak & Noguera, 1995
Aegomorphus arietis (Bates, 1885) comb. nov.
Aegomorphus arizonicus Linsley & Chemsak, 1984
Aegomorphus auratus (García & Nascimento, 2019) comb. nov.
Aegomorphus atosignatus (Melzer, 1932) comb. nov.
Aegomorphus bezarki (Santos-Silva & Galileo, 2016) comb. nov.
Aegomorphus bicuspis (Germar, 1823) comb. nov.
Aegomorphus bimaculatus (Fuchs, 1958) comb. nov.
Aegomorphus binocularis (Martins, 1981) comb. nov.
Aegomorphus bivittus (White, 1855) comb. nov.
Aegomorphus borrei (Dugés, 1885) comb. nov.
Aegomorphus chamelae Chemsak & Giesbert, 1986
Aegomorphus brevicornis (Zajciw, 1964) comb. nov.
Aegomorphus brunnescens (Zajciw, 1963) comb. nov.
Aegomorphus carinicornis (Bates, 1880) comb. nov.
Aegomorphus cerdai (Tavakilian & Néouze, 2013) comb. nov.

Aegomorphus chrysopus (Bates, 1861) comb. nov.
Aegomorphus circumflexus (Jacquelin DuVal, 1857) nomen protectum, comb. nov.
 = *Acanthocinus rusticus* Klug, 1829 nomen oblitum
 = *Psapharochrus histrio* Casey, 1913 syn. nov.
 = *Psapharochrus guatemalensis* Casey, 1913 syn. nov.
Aegomorphus clericus (Bates, 1880) comb. nov.
Aegomorphus comptus (Marinoni & Martins, 1978) comb. nov.
Aegomorphus coniferus (Zajciw, 1963) comb. nov.
Aegomorphus consentaneus (Thomson, 1865) comb. nov.
Aegomorphus contaminatus (Thomson, 1965) revalidat., comb. nov.
Aegomorphus corticarius (Tippmann, 1960) comb. nov.
Aegomorphus crocostigma (Bates, 1880) comb. nov.
Aegomorphus cylindricus (Bates, 1861) comb. nov.
Aegomorphus doctus (Bates, 1880) comb. nov.
Aegomorphus excellens (Zajciw, 1964) comb. nov.
Aegomorphus flavitarsis (Fuchs, 1962) comb. nov.
Aegomorphus galapagoensis galapagoensis (Linell, 1898) comb. nov.
Aegomorphus galapagoensis vonhageni (Mutchler, 1938) comb. nov.
Aegomorphus galapagoensis williamsi (Linsley & Chemsak, 1966) comb. nov.
Aegomorphus geminus (Galileo & Martins, 2012) comb. nov.
Aegomorphus gigas (Galileo & Martins, 2012) comb. nov.
Aegomorphus hebes (Bates, 1861) comb. nov.
Aegomorphus homonymus (Blackwelder, 1946) comb. nov.
Aegomorphus inquinatus (Bates, 1872) comb. nov.
Aegomorphus irumus (Galileo & Martins, 2011) comb. nov.
Aegomorphus itatiayensis (Melzer, 1935) comb. nov.
Aegomorphus jaspideus (Germar, 1823) comb. nov.
Aegomorphus junco (Fisher, 1938) comb. nov.
Aegomorphus laetificus (Bates, 1880) comb. nov.
Aegomorphus lanei (Marinoni & Martins, 1978) comb. nov.
Aegomorphus langeri (Martins, Santos-Silva & Galileo, 2015) comb. nov.
Aegomorphus lateralis (Bates, 1861) comb. nov.
Aegomorphus lengii (Wickham, 1914) comb. nov.
Aegomorphus leucodryas (Bates, 1880) comb. nov.
Aegomorphus longipennis (Zajciw, 1963) comb. nov.
Aegomorphus longispinis (Bates, 1861) comb. nov.
Aegomorphus longitarsis (Bates, 1880) comb. nov.
Aegomorphus lotor (White, 1855) comb. nov.
Aegomorphus luctuosus (Bates, 1880) comb. nov.
Aegomorphus maccartyi (Chemsak & Hovore, 2002) comb. nov.
Aegomorphus maculatissimus (Bates, 1861) comb. nov.
Aegomorphus magnus (Marinoni & Martins, 1978) comb. nov.
Aegomorphus meleagris (Bates, 1861) comb. nov.
Aegomorphus mexicanus Martins, Santos-Silva & Galileo, 2015
Aegomorphus modestus (Gyllenhal, 1817)
Aegomorphus morrisii (Uhler, 1855)
Aegomorphus mourei (Zajciw, 1964) comb. nov.
Aegomorphus nearnsi (Martins & Galileo, 2010) comb. nov.
Aegomorphus nigricans (Lameere, 1884) comb. nov.
Aegomorphus nigromaculatus (Fuchs, 1958) comb. nov.
Aegomorphus nigropunctatus (Tippmann, 1960) comb. nov.
Aegomorphus nigrovittatus (Zajciw, 1969) comb. nov.
Aegomorphus pantherinus (Tavakilian & Néouze, 2013) comb. nov.
Aegomorphus peninsularis (Horn, 1880)
Aegomorphus penrosei Chemsak & Hovore, 2002
Aegomorphus pereirai (Prosen & Lane, 1955) comb. nov.
Aegomorphus peritapnioides (Linsley, 1958) comb. nov.
Aegomorphus phasianus (Bates, 1861) comb. nov.
Aegomorphus pictus (Galileo & Martins, 2012) comb. nov.
Aegomorphus pigmentatus (Bates, 1861) comb. nov.

Aegomorphus pinima (Galileo & Martins, 2006) comb. nov.
Aegomorphus piperatus (Gahan, 1892) comb. nov.
Aegomorphus piraiuba (Martins & Galileo, 2003) comb. nov.
Aegomorphus polystictus (Bates, 1885) comb. nov.
Aegomorphus pseudosatellinus (Tavakilian & Néouze, 2013) comb. nov.
Aegomorphus pupillatus (Bates, 1880) comb. nov.
Aegomorphus purulensis (Bates, 1885) comb. nov.
Aegomorphus quadrigibbus (Say, 1831) comb. nov.
Aegomorphus ramirezi (Chemsak & Hovore, 2002) comb. nov.
Aegomorphus ridleyi (Waterhouse, 1890) comb. nov.
Aegomorphus rileyi (Tavakilian & Néouze, 2013) comb. nov.
Aegomorphus robustus sp. nov.
Aegomorphus rufitarsis (Kirsch, 1889) comb. nov.
Aegomorphus satellinus (Erichson, 1847) comb. nov.
Aegomorphus schmithi (Melzer, 1935) comb. nov.
Aegomorphus signatifrons (Zajciw, 1964) comb. nov.
Aegomorphus signatus (Gahan, 1892) comb. nov.
Aegomorphus socorroensis (Linsley, 1942) comb. nov.
Aegomorphus travassosi (Monné & Magno, 1992) comb. nov.
Aegomorphus umbratus (Bates, 1885) comb. nov.
Aegomorphus vetustus (Bates, 1880) comb. nov.
Aegomorphus wappesi (Galileo, Martins & Santos-Silva, 2015) comb. nov.
= *Psapharochrus jameswappesi* Tavakilian, 2018 syn. nov.

Eupromerella Fisher, 1938

Eupromerella boliviana sp. nov.
Eupromerella clavator (Fabricius, 1801)
Eupromerella fuscicollis (Bates, 1861)
Eupromerella gallardi Tavakilian & Néouze, 2013
Eupromerella griseofasciata (Fuchs, 1958)
Eupromerella inaequalis (Galileo & Martins, 2011) comb. nov.
Eupromerella leucogaea (Erichson, 1847)
Eupromerella maculata Martins, Galileo & Limeira-de-Oliveira, 2009
Eupromerella minima (Bates, 1861)
Eupromerella nigroapicalis (Aurivillius, 1916)
Eupromerella nigroocellata (Tippmann, 1960)
Eupromerella orbifera (Aurivillius, 1908)
Eupromerella picturata Martins, Galileo & Limeira-de-Oliveira, 2009
Eupromerella plaumanni (Fuchs, 1958)
Eupromerella propinqua (Melzer, 1931)
Eupromerella pseudopropinqua (Fuchs, 1958)
Eupromerella quadrituberculata (Zajciw, 1964) comb. nov.
Eupromerella semigrisea (Bates, 1861)
Eupromerella travassosi (Melzer, 1935)
Eupromerella versicolor (Melzer, 1935)

Scythropopsis Thomson, 1864 resurrection of the original status

Scythropopsis abstersa (Bates, 1880)
Scythropopsis albitarsis (Laporte, 1840)
Scythropopsis barrerae (Chemsak & Hovore, 2002) comb. nov.
Scythropopsis boucheri Tavakilian & Néouze, 2013
Scythropopsis cornuta (Bates, 1880) comb. nov.
Scythropopsis intricata sp. nov.
Scythropopsis lacrymans (Thomson, 1865) comb. nov.
Scythropopsis lugens (Thomson, 1865) comb. nov.
Scythropopsis melanostictica (White, 1855) comb. nov.
= *Pteridotelus contaminatus* Thomson, 1865 syn. nov.

Scythropopsis nigratarsis (White, 1855) comb. nov.
Scythropopsis pupillata (Bates, 1880)
Scythropopsis sallei (Thomson, 1865) comb. nov.
Scythropopsis wappesi (Chemsak & Hovore, 2002) comb. nov.

Symperasmus Thomson, 1864 resurrection of the original status

Symperasmus affinis (Thomson, 1865)
Symperasmus alboniger (Bates, 1861) comb. nov.
= *Acanthoderes (Psapharochrus) albomaculatus* Fuchs, 1963 syn. nov.
= *Acanthoderes griseomaculata* Zajciw, 1971 syn. nov.
Symperasmus thoracicus (White, 1855) comb. nov.

ACKNOWLEDGMENTS

Special thanks to the individuals who loaned specimens from their collections, or those in their care, for inclusion in this study. They include Kyle Schnepf and Paul Skelley (FSCA), Steven W. Lingafelter, Roy F. Morris and Fred W. Skillman. The second author is grateful to the "Fundação de Amparo à Pesquisa do Estado de São Paulo" (FAPESP) for a postdoctoral fellowship (process number 2017/17898-0).

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