

Phylogenetic and pathotypic characterization of newcastle disease virus in Tibetan chickens, China¹

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Chickens are considered to be potential reservoirs of Newcastle disease virus (NDV). In this study, six Newcastle disease virus strains were isolated and characterized in Tibetan chickens. The HN gene was sequenced, and phylogenetic relationship to reference strains was studied. The phylogenetic analysis demonstrated that these six isolated strains were closely related to NDV isolates of the reference strains GQ245823, KT002186, KU527561, KJ563939, AY225110, EU305607, KM056357, Y18898, GQ245832, AF077761 and lasota strain. Among them, EU305607, KJ563939 and KM056357 were isolated from India, while lasota strain came from attenuated vaccine widely used in China. Then, mean death time (MDT) and intracerebral pathogenicity index (ICPI) were used to estimate the pathogenicity of the isolates. Pathogenicity experiment showed HNH1 and HN17 to be virulent. Our results indicated that genetically diverse viruses circulate in Tibetan chickens, and based upon the phlogeographic analysis, we estimated the origin of ancestral viruses of the isolates and its sister strains located in India and China (lasota strain). It indicates the importance of continuous surveillance to enhance current understanding of the genetic evolution of the NDV strains.

INDEX TERMS: Newcastle disease virus (NDV), Tibetan chickens, phylogenetic analysis, pathogenicity.

INTRODUCTION

Newcastle disease (ND) is recognized as one of the highly contagious and lethal infectious disease of poultry industry globally (Munir et al. 2015), mainly characterized by diarrhea, dyspnea, neurological symptoms, cloaca hemorrhage, focal glandular gastric bleeding or ulcers, intestinal mucosal bleeding and necrosis of the pancreas or the spleen (Zhang et al. 2011). ND is caused by virulent strains of the ND virus (NDV), which belong to the genus Avulavirus, sub-family Paramyxoviridae, family Paramyxovirinae (Ge et al. 2015), with six transcriptional units encoding nucleocap-

sid protein (NP), phosphoprotein (P), matrix protein (M), fusion protein (F), haemagglutinin-neuraminidase protein (HN) and polymerase protein (L) (Mayo 2002a, 2002b).

For many years, NDV has caused large-scale outbreaks in many countries around the world, especially in China (Qin et al. 2008, Zhang et al. 2010, Kang et al. 2016), and the infection types and the virulent strength vary with obvious regionality. In spite of extensive vaccination with the live attenuated vaccines, massive outbreaks are reported in China, and there is a long way to go to ultimately eradicate NDV (Bogoyavlenskiy et al. 2009, Kumar 2015). Previously, the phylogenetic and pathotypic characterization of NDV has been reported in most breeds of chickens in China (Cao et al. 2001, Qin et al. 2008, Zhang et al. 2010); however, none of them has been reported in Tibetan chickens.

The Tibetan chicken, aboriginal chicken breed, has a very wide distribution at altitudes of 2,200 to 4,100 m in the Qinghai-Tibet Plateau, with a history of domestication more than 1,000 years at high altitude, breeds successfully and has good resistance to hypobaric hypoxia and cold grazing conditions (Zhang et al. 2005, Zhang et al. 2006).

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To the best of our knowledge, no information is available on the genetic characterization and pathogenicity of NDV in Tibetan chickens from the Tibetan Plateau of China. In this study, we investigated the presence of NDV in Tibetan chickens and selected six viruses to evaluate the genetic characterization and pathogenicity of NDV derived from Tibetan chickens.

MATERIALS AND METHODS

The study site. The present study was carried out in Nyingchi Prefecture, in southeastern Tibet, China. As described by researcher (Zhang et al. 2014a), this area is geographically isolated from Tibet and Sichuan Provinces by Himalayas, and shares borders with Yunnan and Qinghai Provinces. Nepal in the southwest, and India and Myanmar in the south. The average elevation of the surveyed area is 3100 m above sea level.

Sample collection, NDV isolation and identification. A total of six NDVs were isolated from the organs (brain, heart, liver, spleen, lungs) of dead or killed moribund Tibetan chickens between 2015 and 2016. The virus isolation was carried out by using the guidelines and standards provided by the World Organization for Animal Health (Anonymous 2006). Briefly, homogenized and clarified organ suspensions were made in viral transportation media (VTM) containing penicillin (1000 U/mL) and streptomycin (1000 U/mL), at 4°C overnight, and the organ suspensions were plaque-purified three times by inoculating into the allantoic cavity of 9 to 11 day old specific pathogen free (SPF, Beijing Meril Vital Laboratory Technology Co., Ltd) chicken embryos (five eggs per sample) according to the standard methods of the virus isolation, and the chicken embryos were incubated at 37°C (Kumar & Kumar 2015). After 3 days, allantoic fluid was harvested and after three series of blind passages, the NDV in the harvested allantoic fluid was tested using haemagglutination inhibition (HI) and haemagglutination (HA) tests. Inactivated NDV antigen strain (lasota strain) was used as a positive control. The virus stocks grown in allantoic fluids were stored at -70°C until subsequent use and further analysis.

Pathogenicity analysis. The pathogenic potential for the isolated viruses was evaluated using standard assay methods to determine the mean death time (MDT) using 9-days-old SPF embryonated chicken eggs and intracerebral pathogenicity index (ICPI) test using 1-day-old SPF chicks (Anonymous, 2006, Kim et al. 2012).

Molecular detection and characterization. Total RNA of the isolated strains was extracted using the TIANamp Virus DNA/RNA Kit (TianGen, China) according to the instructions of manufacture. A RT-PCR amplification approach was used to amplify HN and F genes. Based on the published F and HN gene sequences in the GenBank database, we designed two pairs of primers (Table 1).

For PCR, forward (5'-ATGGACCGCGCGTTAGC-3') and reverse (5'-CTAGCCAGACCTGGCTTCTCT-3') primers were used. Polymerase chain reaction was used and reverse transcription was done by Quant One Step RT-PCR Kit (TianGen Co., LTD, China). The PCR was performed with the following cycling protocol: one cycle at 95°C for 40 s, and 35 amplification cycles at 56°C for 50s, and 72°C for 1.2 min after an initial hot start at 95°C and ending with

72°C each for 10 min. The PCR products were separated on agarose gel (1 %) along with ethidium bromide (at the rate of 0.5µg/ml) in the 0.5×TBE buffer at 5 V/cm for 30 minutes by electrophoresis. The RT-PCR products were analyzed on a 1.5% agarose gel stained with ethidium bromide following electrophoresis and the products were purified using a TaKaRa MiniBEST Agarose Gel DNA Extraction Kit Ver.4.0 (Takara Biotechnology Co., Ltd, Dalian, China) according to manufacturer's instructions. The positive products were cloned into PGEM®-T Easy vector (You ni kang Co., Ltd, China) and the positive clones were identified by blue-white colony screening method. Further confirmation was carried out by PCR. Then the positive products were sequenced by a commercial company (Sangon Biosciences Co., Ltd, Wuhan, China).

Phylogenetic analysis. The nucleotide sequences editing of the HN gene of the six isolates in this study were compared with reference NDV sequences available at NCBI database using the maximum likelihood method with statistical analysis based on 1000 bootstrapping replicates. The alignments were conducted using Molecular Evolutionary Genetic Analysis (MEGA 6.0) and the DNASTAR (version 3.3.8; DNASTAR) software. Phylogenetic tree was constructed using the neighbor-joining method. The evolutionary distances were estimated using the Kimura two-parameter method (Tamura et al. 2013).

RESULTS AND DISCUSSION

The genetic characterization and pathogenicity of NDV has been reported in most parts of China; however, no such information is available in Tibetan chickens, China. In our study, six NDVs from Tibetan chickens were isolated on SPF embryonated chicken eggs in Tibet from 2013 to 2015, and the details of the NDV isolates are shown in Table 2. The identification and characterization was based on clinical symptoms, pathological observation and molecular investigations.

Table 2. The characteristics description of six Newcastle disease virus isolates from Tibetan chickens

NDV isolates	Year of isolation	Location	Pathotype	F-protein cleavage site ^a	MDT ^b	ICPI ^c
HNH1	2015	Bayi	Velogenic	RRQKRF	46	1.54
HN20	2016	Bayi	Lentogenic	GRQGRL	>120	0.2
HN10-T	2015	Bayi	Lentogenic	GRQGRL	>120	0.2
HN5-T	2015	Bujiu	Lentogenic	GRQGRL	>120	0
NHH3	2015	Mirui	Lentogenic	GRQGRL	>120	0
HN17	2016	Mirui	Velogenic	RRQKRF	43	1.66

^a Amino acid sequence from 112 to 117 of F protein. ^b Mean death time in 9-day-old SPF embryonated chicken eggs (hours) (velogen <60, mesogen 60-90, lentogen >90). ^c Intracerebral pathogenicity index in 1-day-old chickens (lentogen <0.7, mesogen 0.7-1.4, velogen 1.4-2.0).

As shown in Table 2, HNH1 and HN17 strains were classified as velogenic NDV, which were consistent with the typical virulent motif 112 RRQKRF 117 at the F cleavage sites; while HN20, HN10-T, HN5-T and NHH3 were lentogenic with 112 GRQGRL 117 at the F cleavage sites. These amino acids changes in the protein may be related to change of pathogenicity because all six isolates presented high homology between themselves and against attenuated vaccine lasota, including HNH1 and HN17 isolates, which were classified as velogenic pathotype. Furthermore, the lentogenic NDV strain was reported in Tibet (Cao et al. 2001),

Table 1. Primers used in the study

Fragment designation	Primer sequence (5'-3')	Expected size (bp)
F- forward	ATGGACCGCGCGTTAGC	1662
F- reverse	CTAGCCAGACCTGGCTTCTCT	
HN- forward	ATGGGCTCCAAACCTTCTA	1713
HN- reverse	TCTGTGGTGGCTCTCAT	

rate) and vaccine (wrong immunization approach; inadequate immunization dose; unreasonable vaccine dilutions; uneven dilutions, the interference of maternal antibody; poor quality vaccines). At present, lasota is widely and frequently used to prevent NDV infection in the poultry industry in China, and virulent NDV infections continue to occur in the well-vaccinated chicken flocks (Roy et al. 2004, Qin et al. 2008, Zhang et al. 2014a,b). Although this virus did not cause disease in chickens, the virus could effectively infect and replicate in these birds. Therefore, new vaccines to prevent the emerging infection in the chicken flocks should be developed as soon as possible, along with strict biosecurity measures to reduce the risk of Newcastle disease outbreaks. It suggests that irrational use of attenuated vaccines in Tibetan chickens may attribute to NDV infection (Palm et al. 2015). In addition, increase number of migratory birds to Tibet may be another possible explanation of ND in this region, as these birds play an important role in the transmission of NDV and act as main reservoirs of the virus.

In conclusion, this is the first report describing the occurrence and baseline genetic characteristics of NDV in high altitude region of China (Tibet). Our results revealed that Tibetan chickens carry NDVs of both velogenic and lentogenic types. Furthermore, genetic characterization showed clear evidence that irrational use of vaccines may be the key contributor to the occurrence of NDV in the study area. This report may help in the prevention and control strategies against NDV in Tibet, China.

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Conflict of interest.- None of the authors have any conflict of interest.

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