

Biology of *Triatoma carcavallo* Jurberg, Rocha & Lent, 1998 under laboratory conditions

**Margareth Cardozo-de-Almeida^{[1],[2]}, Simone Caldas Teves Neves^[1],
Carlos Eduardo de Almeida^[3], Nathanielly Rocha Casado de Lima^[1],
Maria Luiza Ribeiro de Oliveira^{[1], [2]}, Jacenir Reis dos Santos-Mallet^[1]
and Teresa Cristina Monte Gonçalves^[1]**

[1]. Laboratório de Transmissores de Leishmanioses, Setor de Entomologia Médica e Forense, Fundação Oswaldo Cruz, Rio de Janeiro, RJ. [2]. Programa de Pós-Graduação em Biologia Animal, Universidade Federal Rural do Rio de Janeiro, Seropédica, RJ. [3]. Departamento de Ciências Biológicas, Faculdade de Ciências Farmacêuticas, Universidade Estadual Paulista, Araraquara, SP.

ABSTRACT

Introduction: *Triatoma carcavallo* is a wild species that is found in sympatry with *Triatoma rubrovaria* and *Triatoma circummaculata*, which are vectors of *Trypanosoma cruzi* currently found in rural areas of Rio Grande do Sul, Brazil. **Methods:** Fertility was assessed and to determine the incubation period, the eggs were observed until hatching. The first meal was offered to 1st stage nymphs. The intermolt period was also determined. The number of blood meals was quantified at each nymphal stage and the resistance to fasting as the period between ecdysis and death. Mortality was assessed and longevity was determined by recording the time that elapsed from molting to the adult stage and until death. The developmental cycle was assessed by recording the length in days of each stage from molting to adult hood. **Results:** The average incubation period was 22.7 days. The average first meal occurred 3.1 days after hatching. The 5th stage nymph to adult intermolt period was the longest at 193.4 days. The average number of feedings during nymphal development was 13.4. The resistance to fasting assay indicated that the 3rd, 4th and 5th stage nymphs presented higher resistance than did adults. The highest mortality rate was observed in the 3rd stage nymphs (22.2%). The average length of adult survival was 25.6 weeks, and the average total life cycle lasted 503.4 days. **Conclusions:** This study is the first report on the biology of *T. carcavallo* that fed on mice. The presented findings expand the bionomic knowledge of these species.

Keywords: Triatominae. *Triatoma carcavallo*. Biological behavior.

INTRODUCTION

The Brazilian State of Rio Grande do Sul harbors eleven species of triatomines, including introduced species with synanthropic habits. These insects are associated with bird nests, mammals and rocky habitats. They are considered autochthonous¹ and are dispersed in a discontinuous pattern throughout the state, with some species restricted to the central-southern region and others in the northwest and northeast regions. Among the wild species of triatomines, *Triatoma rubrovaria* (Blanchard, 1843) is the most important in epidemiological terms due to its wide geographic distribution and capacity to transmit *Trypanosoma cruzi* (Chagas, 1909), followed by *Triatoma circummaculata* (Stål, 1859) and *Triatoma carcavallo* (Jurberg, Rocha & Lent, 1998). These

species are sympatric, inhabiting rocky environments and exhibiting feeding habits that are remarkably eclectic, with irregular intervals observed in their biological cycles². Field data have shown that *T. rubrovaria* is frequently found in domiciliary and peridomiciliary areas³. Only *T. carcavallo* has been observed in Canguçu, Dom Feliciano, Pinheiro Machado and São Jerônimo⁴ (Source: IPB-LACEN/RS), appearing inside domiciles throughout Rio Grande do Sul^{5,6}.

Morphological observations have led *T. carcavallo* to be included in the same species complex (*infestans* complex) and subcomplex (*rubrovaria* subcomplex) as *T. rubrovaria*⁷.

Characterizing the biology of wild species is important while evaluating the efficiency of these insects as vectors of *T. cruzi*, as these species consistently invade environments that are subject to human modification. As a result, knowledge of the biological characteristics of these insects is essential for designing control strategies, mainly in relation to secondary vectors that have the potential to become established in human habitats⁸.

The life cycles of triatomines vary according to the species, environmental conditions and the availability of suitable sources of blood^{9,10}.

Resistance to fasting could be of great importance and might directly affect control campaigns targeting these vectors¹¹. Tolerance to long periods of fasting helps these

Address to: Dr^ª Jacenir Reis dos Santos Mallet. Lab. Transmissores de Leishmanioses/Setor Entomologia Médica e Forense/FIOCRUZ. Av. Brasil 4365, 21045-900 Rio de Janeiro, RJ, Brasil.

Phone: 55 21 2562-1352/1232

e-mail: jacenir@ioc.fiocruz.br

Received 27 February 2014

Accepted 18 June 2014

insects survive difficult periods when food shortages occur. During these periods, they hide in wall cracks and escape residual insecticides¹², after which they are able to recolonize the household.

The aim of this study was to examine the bionomic features of *Triatoma carcavallo* under laboratory conditions, including incubation time, fertility, first meal of the 1st stage nymphs, intermolt period, number of blood meals, resistance to starvation, mortality, longevity and developmental cycle to better understand its potential capacity as a vector and to use this knowledge to help monitor this species during control activities.

METHODS

Specimens of *T. carcavallo* were collected in the State of Rio Grande do Sul, Brazil, in Encruzilhada do Sul City (30°32'38"S; 52°31'19"W), from natural ecotopes and peri-household locations. The insects were maintained at 26°C under 70% RH (relative humidity) in the Laboratory of *Transmissores de Leishmanioses*, Department of *Entomologia Médica e Forense*, Oswaldo Cruz Institute, *Fundação Oswaldo Cruz*.

Fertility

Sixteen females were individually separated into Borrel tubes closed with nylon mesh and observed daily, and the color of their eggs and whether they hatched was recorded.

Incubation period

Sixty eggs from eight females were separated into Borrel tubes closed with nylon mesh (according to the laying date) and then observed daily until they hatched.

First meal

Thirty eggs were individually housed in Borrel tubes. After hatching, a blood meal from a Swiss mouse was offered to each nymph within 10 minutes.

Intermolt period

Fifty nymphs from each stage, as well as 15 males and 15 females, were separated and placed in individual plastic containers (5.5cm in diameter × 10.5cm in height) containing individuals from the same day of hatching or ecdysis. Filter paper was used to cover the inner surface of the container. The specimens were observed daily from ecdysis until the next stage. The intermolt period was noted as the number of days between two consecutive ecdysis events, and the period of nymphal development was recorded as the number of days between the hatching date and ecdysis into the imago stage.

Number of blood meals according to the nymphal stage

The number of blood meals taken during each nymphal stage was quantified. A blood meal was provided once a week until repletion.

Resistance to fasting

The period between ecdysis and the death of the insect was recorded. The insects fed on mice (*Mus musculus*) weekly for a period of 60-80 minutes. After ecdysis, the specimens were separated into the nymphs of each phase (n = 50) and adults (n = 30) and stored in Borrel tubes with other individuals from the same day of ecdysis. The specimens were observed daily over the period from ecdysis to death.

Mortality

In total, 177 1st stage nymphs were separated, and the mortality was calculated according to the molting of each nymphal stage up to adulthood.

Longevity

The time elapsed from the molting of each individual to the adult stage until its death was recorded.

Developmental cycle

Sixty eggs laid by females of *T. carcavallo* obtained from the colony maintained in the laboratory were collected by randomly selection and individually placed in plastic containers that contained filter paper to provide the developing insects with access to a food source. Each container was numbered and examined daily until the egg hatched. The nymphs were observed daily and fed on mice weekly until repletion, and the time required to pass through the various developmental stages and reach adulthood was recorded in days. The development times (total and by stage) were expressed using average values.

RESULTS

Fertility

The first oviposition was observed on day 6. The mean number of eggs/week was 30. The greatest number of eggs/week was observed during the summer. It is important to note that the eggs that did not hatch were also fertile, as the coloring was the same for all eggs. The eggs were initially light pink and then became red, which intensified as the process of embryogenesis progressed. The eggs exhibited an ellipsoid shape and adhered to the substrate. The oviposition of these females began after the first meal.

Incubation period

The mean incubation period of the specimens was 22.7 days, with a minimum of 21 and a maximum of 26 days recorded. Among the 60 eggs observed, only 38.3% hatched.

First meal

The 1st stage nymphs obtained their first meal after a mean of 24 days. The first meal occurred during the other nymphal stages after a mean of 3.1 days, with a recorded minimum value of one day and a maximum of five days, although the majority occurred after two or four days.

Intermolting period

Among the 50 nymphs observed, eight died before reaching adulthood (three 1st stage nymphs, four 2nd stage nymphs and one 3rd stage nymph). **Table 1** shows that the shortest intermolting period was observed between the 1st/2nd stages (18.5 days) and the longest between the 5th stage/adulthood (193.4 days). The 5th stage nymphs required several blood meals to molt, which occurred after more than one year.

Number of blood meals

The minimum number of blood meals observed in females and males were zero and one meal, respectively, whereas the maximum number (42 meals) was recorded in the 5th stage nymphs. The mean number of feedings throughout all of nymphal development was 13.4 meals (**Figure 1**).

Resistance to fasting

Observations of all of the nymphal and adult (male and female) stages indicated that the 3rd, 4th and 5th stage nymphs presented greater resistance than did the adults, whereas the males displayed similar resistance to the 1st and 2nd stage nymphs and were less resistant overall than were the females (**Table 2**). The total observation period was 8 months.

Mortality

From the initial 177 *Triatoma carcavallo* nymphs, only 95 reached the adult stage (global mortality rate of 57.2%). The mortality rate for each stage varied from 4.0 to 22.2%. The highest mortality rate was recorded in the 3rd (22.2%) stage nymphs, followed by the 2nd (14%) stage nymphs (**Table 3**).

Longevity

The mean adult survival was 25.6 weeks (n = 42). Males presented a longer life span, with a mean survival of 25.6 weeks, whereas females presented a life span of 23.1 weeks.

TABLE 1 - Intermolting period of *Triatoma carcavallo*.

Stage	N	Duration in days			SD
		minimum	maximum	mean	
1 st	50	15	24	18.52	2.34
2 nd	47	25	95	62.77	18.45
3 rd	43	44	138	86.93	19.00
4 th	42	55	189	119.05	36.38
5 th	42	59	295	193.43	70.33

N: number of nymphs; SD: standard deviation.

TABLE 2 - Resistance to fasting in *Triatoma carcavallo* (days).

Stage	N	Minimum	Maximum	Mean	SD
1 st	50	15	59	37	15.52
2 nd	50	10	116	50	29.72
3 rd	50	3	128	65.5	37.54
4 th	50	58	210	134	54.25
5 th	50	7	185	146	43.08
♀	30	21	81	61	18.30
♂	30	4	81	42.5	18.24

N: number of nymphs; SD: standard deviation; ♀: female ; ♂: male.

Table 4 shows developmental cycle: The total *T. carcavallo* life cycle lasted 503.4 days (72 weeks). Differences were observed for the 5th stage nymphs, whose development lasted a mean of 193.43 days (27 weeks).

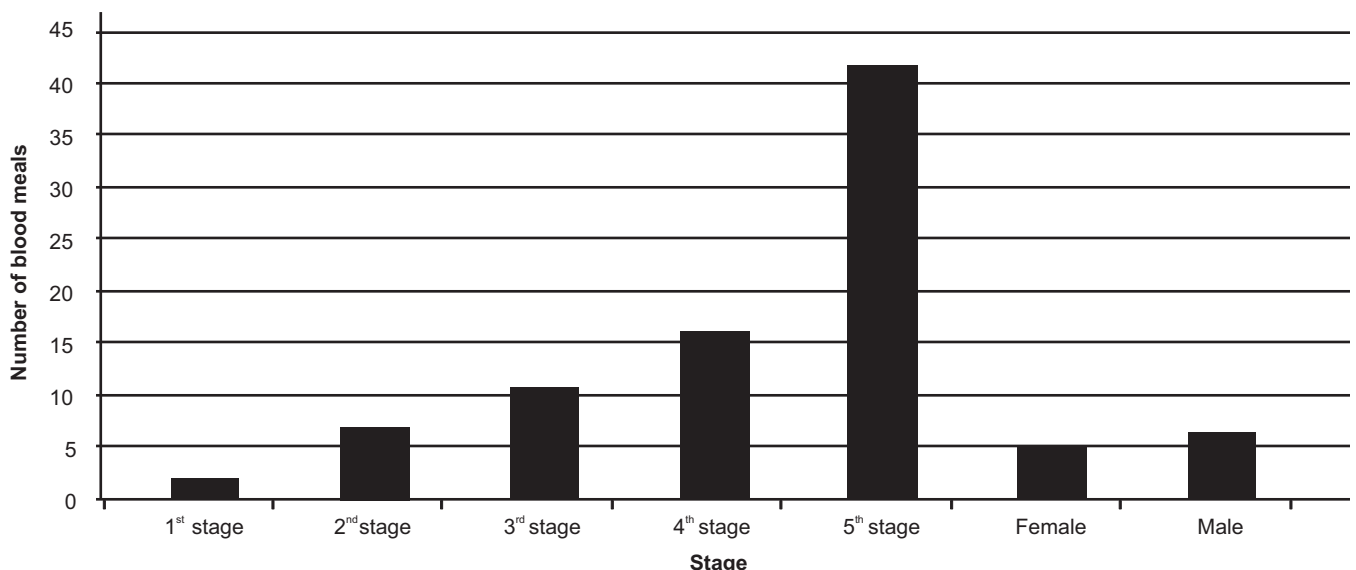


FIGURE 1 - The number of blood meals according to the stage of *Triatoma carcavallo*.

TABLE 3 - Mortality of *Triatoma carcavallo*.

Stage	N	n	Mortality (%)	Cumulative mortality (%)
1 st	177	157	11.3	11.3
2 nd	157	135	14.0	25.3
3 rd	135	105	22.2	47.5
4 th	105	99	5.7	53.2
5 th	99	95	4.0	57.2

N: number of nymphs; n: number of nymphs molting into the following stage.

TABLE 4 - Duration of the developmental cycle of *Triatoma carcavallo*.

Stage	N	Mean (days)	Quantiles 25% - 75%
Egg	60	22.7	21 - 26
1 st	50	18.52	15 - 24
2 nd	47	62.77	25 - 95
3 rd	43	86.93	44 - 138
4 th	42	119.05	55 - 189
5 th	42	193.43	59 - 295
Total	42	503.4	219 - 757

N: number of nymphs.

DISCUSSION

Several authors have highlighted the importance of studying the bionomic features of triatomines under laboratory conditions to increase the biological knowledge of these species, improve breeding conditions for the development of laboratory colonies, and provide recommendations to support control measures¹³⁻¹⁶.

Under laboratory conditions, which generally involve more stable abiotic factors, vital cycles are generally shorter¹⁷. Nevertheless, certain species may present a longer life cycle. For example, the diapause of 5th stage nymphs has been recorded in several wild species¹⁸ and was observed in this study, as fifth-stage nymphs of the examined species require several blood meals to permit molting, which can lengthen this phase up to a year. Similar findings have been reported for *T. rubrovaria*, *T. carcavallo* and *T. circummaculata*¹⁹.

The first study on the biological cycle of *T. carcavallo* under laboratory conditions involved allowing the insects to feed on pigeons, blaberids and lizards²⁰. Other authors have studied the biological cycles of *T. rubrovaria* and *T. circummaculata* fed the hemolymph of Blattodea (cockroaches) to assess nymphal development²¹. These studies suggested that this type of food may be naturally exploited by these triatomine species and most likely represents a means of surviving under natural

conditions. The dependence on hemolymph is obvious in *T. circummaculata*, as this species does not develop into a 1st stage nymph in the laboratory if this food source is not available^{1,5}. Certain species of triatomines display rather eclectic eating habits under natural conditions²².

Triatoma rubrovaria develops well under laboratory conditions at temperatures between 20°C and 30°C and a relative humidity maintained at approximately 80%. The various life cycle times were recorded for the two sexes. Males survived for an average of 115.37 days, whereas females survived for an average of 99.6 days^{16,23}. The influence of temperature (25°C and 30°C) on *T. rubrovaria* was studied, and faster development was observed at 30°C¹⁷. In the present study, temperatures equal to or higher than 28°C influenced the development of *T. carcavallo*, resulting in a shorter life cycle compared to that of *T. rubrovaria*.

In studies examining the life cycle of *T. infestans*²⁴ and the resistance to fasting in *Rhodnius prolixus*²⁵, the authors suggested that the blood meal represents an essential condition for oviposition in general and fertility in particular. In a study of *T. rubrovaria*, it is not known whether the low rate of hatching was due to these factors because the experimental specimens were fed once a week for an hour, without establishing the amount of blood ingested by each individual²⁶. In the present study, the feeding was standardized, with *T. carcavallo* being fed mice once a week until repletion. Therefore, we cannot conclude whether the low rate of hatching recorded was due to food or other factors, such as temperature and humidity, present under the laboratory conditions.

In our experiments, not all of the eggs hatched, indicating that fertility was compromised or that small variations in temperature or humidity (which are known to influence development directly) or some other unknown cause limited hatching.

The hatching incidence varies among species of the *Triatoma* genus and even within the same species. In *T. carcavallo*, the obtained rate of 38.3% is considered low compared to those of *Triatoma flavida*, in which 93% of eggs have been reported to hatch²⁷; *T. pseudomaculata*, with a rate was 88.3%²⁸; and *T. rubrovaria*²⁹, *Triatoma dimidiata*³⁰, *T. infestans*³¹, *T. maculata*³², *Triatoma rubrofasciata*³³ and *Triatoma pallidipennis*³⁴, in which rates range from 60 to 80%. In the *Rhodnius* genus, temperature was observed to influence not only the incubation period but also the incidence of hatching, as shown for *R. robustus*³⁵.

The average incubation period reported for *T. rubrovaria* is 24.84 days²⁶, whereas the incubation period recorded for *T. carcavallo* in the present study was 22.7 days.

In this study, the search for the first meals of the 1st stage nymphs of *T. carcavallo* showed a higher frequency on days 2 and 4, similar to results reported for *T. pseudomaculata*²⁸ but different from that of *Triatoma vitticeps*, which shows a preference for days 3, 6 and 10³⁶.

It is important to manage newly fed triatomines carefully because improper handling is a major cause of mortality during their development^{17,37}.

The *T. carcavallo* life cycle requires an average of 13.4 blood meals, which is higher than the values reported for *T. rubrovaria* (11.1)³⁸,

Triatoma vitticeps (8)³⁶ and *Triatoma brasiliensis* (11)³⁹ but lower than the value reported for *T. pseudomaculata* (14.7)²⁸. However, differences in the experimental conditions used by the various authors must be considered. We also must consider that these species live in distinct habitats and exhibit different feeding habits. Females of *T. pseudomaculata* make up for the lower amount of blood ingested by seeking out food sources quickly and consuming a larger number of blood meals²⁸. The 5th stage nymphs of *T. carvalhoi* require several feedings to perform ecdysis.

The average intermolt period of the 2nd stage nymphs of *T. rubrovaria* was longer than that of the 3rd stage nymphs³⁸. *Triatoma brasiliensis*, under standard temperature conditions, tends to show a crescent-shaped scale of intermolt periods from the 1st stage to the 5th stage nymphs^{15,17}, which corroborates our findings.

One of the barriers to successfully combating the vectors of Chagas disease (using insecticides with residual action) is the capacity of these insects to resist fasting⁴⁰. Nymphs tolerate longer fasting periods than do adults¹⁸, confirming that our findings indicate a compensation mechanism for the higher dispersion capacity of adults²⁵.

The ability to resist fasting increased in *T. carvalhoi* from the 1st to the 5th nymphal stage. During the adult phase, lower resistance was observed in males compared to females, corroborating data obtained at a temperature of 30°C for *T. rubrovaria*¹⁷ and *T. sordida*⁴¹. These results contradict results reported for *T. vitticeps*⁴⁰ and *T. rubrovaria*⁴², in which males display a higher resistance to fasting.

In the present study the mortality rate was higher in the 3rd stage nymphs of *T. carvalhoi* (22.2%), corroborating the rate observed in *T. rubrovaria*⁸. When nymphs of *T. infestans* were fed at longer intervals, their mortality rate increased, indicating the influence of feeding on nymphal mortality^{24,43}.

In this study, the higher longevity was observed in *T. carvalhoi* males, as was also observed in *T. rubrofasciata*³³ and *T. rubrovaria*^{8,17,26,29}. The longevity of *T. carvalhoi* females was found to be higher when they were fed pigeons, suggesting a likely influence of feeding¹⁹.

The developmental cycle of the fifth stage nymphs of *T. carvalhoi* fed with pigeons lasted an average of 259.67 days¹⁹, in contrast to the findings of the present study, in which this species was fed mice and showed an average developmental cycle of 193.43 days. The total life cycle of *T. rubrovaria* has been reported to be approximately 300 days²⁹, whereas a much longer duration was observed in *T. carvalhoi* in the present study (503.4 days).

Our results suggest that biological traits are important criteria for determining the relationships between the *Triatoma carvalhoi*, *Triatoma circummaculata* and *Triatoma rubrovaria* species in the presence of the same food source and climatization and based on specimens collected in domiciles in State of Rio Grande do Sul, Brazil.

The efficacy of vector control campaigns has been impaired by the resistance to fasting. This resistance allows these animals to remain in their shelters, free from insecticides and sometimes

even from their residual effects, thereby increasing the possibility of later recolonization by the remaining individuals^{12,14,42,44-47}.

The capacity for transmission observed in *T. carvalhoi* is as high as that of *T. rubrovaria*, whereas *T. circummaculata* is infected less often¹, most likely because it feeds on mammals less frequently. These data together with the results of this study demonstrate the need for constant entomological surveillance of *T. carvalhoi*.

ACKNOWLEDGMENTS

We thank Dra. Célia Lammerhist and MSc. Cleonara Bedin for support during the field collection of triatomines.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

FINANCIAL SUPPORT

Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ) and Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES).

REFERENCES

1. Ruas-Neto AL, Corseuil E. Hábitos, Distribuição Geográfica e Potencialidade dos triatomíneos rupestres como vetores da doença de Chagas no Rio Grande do Sul, Brasil (Hemiptera: Reduviidae: Triatominae). *Entomol y Vect* 2002; 9: 231-249.
2. Cardozo-de-Almeida MAR. Estudos morfológicos, morfométricos e ultraestruturais em cinco espécies do gênero *Triatoma* Laporte, 1832 (Hemiptera: Reduviidae: Triatominae) incluindo a biologia de *Triatoma carvalhoi*. [Dissertation]. [Rio de Janeiro]: Universidade Federal Rural do Rio de Janeiro 2007; 102 p.
3. Costa J, Lorenzo M. Biology, diversity and strategies for the monitoring and control of triatomines - Chagas disease vectors. *Mem Inst Oswaldo Cruz* 2009; 104 (supl I): 46-51.
4. Bedin C, Mello F, Wilhelms TS, Torres MA, Estima C, Ferreira CF, et al. Vigilância Ambiental: Doença de Chagas no Rio Grande do Sul. *Bol Epidemiológico* 2009; 11:8.
5. Martins LP, Castanho RE, Casanova C, Caravelas DT, Frias GT, Ruas-Neto AL, et al. Triatomíneos rupestres infectados por Trypanosomatidae, coletados em Quaraí, Rio Grande do Sul, 2003. *Rev Soc Bras Med Trop* 2006; 39:198-202.
6. Jurberg J, Rocha DS, Lorosa ES, Vinhaes MC, Lent H. Uma nova espécie de *Triatoma* do Estado do Rio Grande do Sul, Brasil (Hemiptera, Reduviidae, Triatominae). *Entomol y Vect* 1998; 5:295-310.
7. Almeida CE, Marcet PL, Gumiel M, Takiya DM, Cardozo-de-Almeida MAR, Pacheco RS, et al. Phylogenetic and phenotypic relationships among *Triatoma carvalhoi* (Hemiptera: Reduviidae: Triatominae) and related species collected in domiciles in Rio Grande do Sul State, Brazil. *J Vect Ecology* 2009; 34:164-173.
8. Oscherov EB, Bar ME, Damborsky MP, Milano AMF. Estadísticos poblacionales de *Triatoma rubrovaria* en condiciones de laboratorio. *Rev Saude Publica* 2005; 39:211-216.
9. Schofield CJ. Population dynamics and control of *Triatoma infestans*. *Ann Soc Belge Med Trop* 1985; 65: 149-164.

10. Carbajal de la Fuente AL, Cunha V, Rocha NL, Lopes CM, Noireau F. Comparative biology of the two sister species of Triatominae (Hemiptera: Reduviidae). *Rev Soc Bras Med Trop* 2010; 43:15-18.
11. Moreira CJC, Spata MCD. Dynamics of evolution and resistance to starvation of *Triatoma vitticeps* (Stål, 1859) (Reduviidae: Triatominae), submitted to two different regimens of food deprivation. *Mem Inst Oswaldo Cruz* 2002; 97:1049-1055.
12. Cabello DR. Resistance to Starvation of *Rhodnius neivai* Lent, 1953 (Hemiptera: Reduviidae: Triatominae) under Experimental Conditions. *Mem Inst Oswaldo Cruz* 2001; 96:587-591.
13. Rocha DS, Galvão C, Jurberg J. Biologia do *Rhodnius pictipes* Stål, 1872 em condições de laboratório (Hemiptera, Reduviidae, Triatominae). *em Inst Oswaldo Cruz* 1994; 89:265-270.
14. Galvão C, Jurberg J, Lent H. Resistência ao jejum de *Triatoma nitida* Usinger, 1939 em laboratório (Hemiptera: Reduviidae). *Mem Inst Oswaldo Cruz* 1996; 91:639-640.
15. Costa J, Marchon-Silva V. Período de intermuda e resistência ao jejum de diferentes populações de *Triatoma brasiliensis* Neiva, 1911 (Hemiptera, Reduviidae, Triatominae). *Entomol y Vect* 1998; 5:23-34.
16. Rodrigues VLCC, Ferraz Filho AN, Silva EOR. *Triatoma rubrovaria* (Blanchard, 1843): tábuas das ninfas, duração das formas e oviposição das fêmeas. *Rev Soc Bras Med Trop* 2005; 38:251-254.
17. Silva IG. Influência da temperatura na biologia de 18 espécies de triatomíneos (Hemiptera: Reduviidae) e no xenodiagnóstico. [Thesis]. [Curitiba]: Universidade Federal do Paraná 1985; 169 p.
18. Lent H, Wygodzinsky P. Revision of the Triatominae (Hemiptera, Reduviidae) and their significance as vectors of Chagas' disease. *Bull American Mus Nat Hist* 1979; 163:125-520.
19. Ruas-Neto AL. Aspectos morfológicos, distribuição geográfica, hábitos e importância vetorial de *Triatoma carcavallo* Jurberg, Rocha & Lent, 1998, *Triatoma circummaculata* (Stål, 1859) e *Triatoma rubrovaria* (Blanchard, 1843), triatomíneos rupestres do Rio Grande do Sul (Hemiptera: Reduviidae: Triatominae). [Thesis]. [Porto Alegre]: Pontifícia Universidade Católica do Rio Grande do Sul 2002; 108p.
20. Ruas-Neto AL, Corseuil E, Cavalleri A. Development of rupestrian triatomines (Hemiptera: Reduviidae: Triatominae) following hemolymphagy on blaberids (Blattodea: Blaberidae) in Rio Grande do Sul State, Brazil. *Entomol y Vect* 2001; 8:205-216.
21. Lorosa ES, Jurberg J, Souza ALA, Vinhaes MC, Nunes IM. Hemolinfa de Dictyoptera na manutenção do ciclo biológico silvestre de *Triatoma rubrovaria* (Blanchard, 1843) e *Triatoma circummaculata* (Stål, 1859) (Hemiptera, Reduviidae, Triatominae). *Entomol y Vect* 2000; 7:287-296.
22. Lorosa ES, Nunes IM, Vinhaes MC, Esteves-de-Andrade R, Jurberg J. Preferência alimentar de algumas espécies de Triatomíneos capturados no estado do Rio Grande do Sul, Brasil, com auxílio da técnica de precipitina e grau de infectividade. *Entomol y Vect* 2000; 7:211-225.
23. Di Primio R. Sobre o *Triatoma rubrovaria* (Blanchard, 1843) no Rio Grande do Sul. *Rev Med Rio Grande do Sul* 1953; p. 402-408.
24. Perlowagora-Szumlewicz A. Estudos sobre a biologia do *Triatoma infestans*, o principal vetor da doença de Chagas no Brasil (importância de algumas de suas características biológicas no planejamento de esquemas de combate a esse vetor). *Rev Bras Malariol D Trop* 1969; 21:117-159.
25. Feliciangeli D, Rabinovich J, Fernandez E. Resistencia al ayuno em triatomíneos (Hemiptera, Reduviidae) Venezolanos. I. *Rhodnius prolixus* Stål. *Rev Inst Trop São Paulo* 1980; 22:53-61.
26. Argüello NV, Mischis CC, Civitta G, Bonino EE. Ciclo biológico de *Triatoma rubrovaria* (Blanchard, 1843) (Reduviidae, Triatominae) em laboratório. *Rev Bras Zool* 1988; 5:245-251.
27. Cabello DR, Lizano E. Biology of *Triatoma flavida* Neiva, 1911 (Hemiptera: Reduviidae) under Laboratory Conditions. *Mem Inst Oswaldo Cruz* 2001; 96:879-881.
28. Gonçalves TCM, Cunha V, Oliveira E, Jurberg J. Alguns aspectos da biologia de *Triatoma pseudomaculata* Côrrea & Espinola, 1964, em condições de laboratório (Hemiptera: Reduviidae: Triatominae). *Mem Inst Oswaldo Cruz* 1997; 92:275-280.
29. Damborsky MP, Bar ME, Gorla D. Life cycle and reproductive patterns of *Triatoma rubrovaria* (Blanchard, 1843) (Hemiptera: Reduviidae) under constant and fluctuating conditions of temperature and humidity. *Rev Soc Bras Med Trop* 2005; 38:433-437.
30. Zeledón R, Guardia VM, Zúñiga A, Swartzwelder JC. Biology and ethology of *Triatoma dimidiata* (Latreille, 1811). I. Life cycle, amount of blood ingested, resistance to starvation, and size of adults. *J Med Entomol* 1970; 7:313-319.
31. Rabinovich JE. Vital statistics of Triatominae (Hemiptera: Reduviidae) under laboratory conditions. I. *Triatoma infestans* Klug. *J Med Ent* 1972; 9:351-370.
32. Feliciangeli MD, Rabinovich J. Vital statistics of Triatominae (Hemiptera: Reduviidae) under laboratory conditions. II. *Triatoma maculata*. *J Med Entomol* 1985; 22:43-48.
33. Braga MV, Pinto ZT, Lima MM. Life cycle and reproductive patterns of *Triatoma rubrofasciata* (De Geer, 1773) (Hemiptera: Reduviidae), under laboratory conditions. *Mem Inst Oswaldo Cruz* 1998; 93:539-542.
34. Martínez-Ibarra JA, Kattchain-Duchateau G. Biology of *Triatoma pallidipennis* (Stål, 1872) (Hemiptera: Reduviidae: Triatominae) under laboratory conditions. *Mem Inst Oswaldo Cruz* 1999; 94:837-840.
35. Rocha DS, Jurberg J, Carcavallo RU, Presgrave OAF, Cunha V, Galvão C. Influência da temperatura e umidade no desenvolvimento ninfal de *Rhodnius robustus*. *Rev Saude Pub* 2001; 35:400-406.
36. Gonçalves TCM, Victorio VNM, Jurberg J, Cunha V. Biologia do *Triatoma vitticeps* (Stål, 1859) em condições de laboratório (Hemiptera, Reduviidae, Triatominae) I. Ciclo evolutivo. *Mem Inst Oswaldo Cruz* 1988; 83:519-523.
37. Gomes AB, Silva IG. Influência da temperatura na biologia de triatomíneos. XXI. *Triatoma jurbergi* Carcavallo, Galvão & Lent, 1998 (Hemiptera, Reduviidae). *Rev Patol Trop* 2000; 29:85-93.
38. Almeida CE, Folly-Ramos E, Agapito-Souza R, Magno-Esperança G, Pacheco RS, Costa J. *Triatoma rubrovaria* (Blanchard, 1843) (Hemiptera, Reduviidae, Triatominae). IV: bionomic aspects on the vector capacity of nymphs. *Mem Inst Oswaldo Cruz* 2000; 100:231-235.
39. Soares RP, Evangelista LG, Laranja LS, Diotaiuti L. Population dynamics and feeding behavior of *Triatoma brasiliensis* and *Triatoma pseudomaculata*, main vectors of Chagas disease in Northeastern Brazil. *Mem Inst Oswaldo Cruz* 2000; 95: 151-155.
40. Gonçalves TCM, Victorio VMN, Jurberg J, Cunha V. Biologia do *Triatoma vitticeps* (Stål, 1859) em condições de laboratório (Hemiptera, Reduviidae, Triatominae) I. Resistência ao jejum. *Mem Inst Oswaldo Cruz* 1989; 84:131-134.
41. Juarez E, Silva EPC. Comportamento do *Triatoma sordida* em condições de laboratório. *Rev Saude Publica* 1982; 16 (suppl):1-35.
42. Almeida CE, Francischetti CN, Pacheco RS, Costa J. *Triatoma rubrovaria* (Blanchard, 1843) (Hemiptera-Reduviidae-Triatominae). III: Patterns of feeding, defecation and resistance to starvation. *Mem Inst Oswaldo Cruz* 2003; 98:761-768.
43. Perlowagora-Szumlewicz A. Ciclo evolutivo do *Triatoma infestans* em condições de laboratório. *Rev Bras Malar* 1953; 5:35-47.
44. Dias JCP. Observações sobre o comportamento de triatomíneos brasileiros frente ao jejum, em laboratório. *Rev Bras Malariol D Trop* 1965; 17:55-63.
45. Costa MJ, Perondini ALP. Resistência do *Triatoma brasiliensis* ao jejum. *Rev Saude Publica São Paulo* 1973; 7:207-217.
46. Cortéz MGR, Gonçalves TCM. Resistance to starvation of *Triatoma rubrofasciata* (De Geer, 1773) under laboratory conditions (Hemiptera: Reduviidae: Triatominae). *Mem Inst Oswaldo Cruz* 1998; 93:549-554.
47. Cailleaux SRP, Cunha V, Verly S, Lamas Junior VD, Jurberg J. Resistência ao jejum de *Rhodnius stali* Lent, Jurberg & Galvão, 1993 (Hemiptera, Reduviidae, Triatominae) em condições de laboratório. *Rev Pan-Amaz Saude* 2011; 2:39-43.