Major Article

Identification and characterization of methicillin-resistant Staphylococcus spp. isolated from surfaces near patients in an intensive care unit of a hospital in southeastern Brazil

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Abstract

Introduction: Contaminated hospital environments contribute to the transmission of microorganisms associated with healthcare. Contaminated surfaces handled by patients or healthcare professionals are a source of microorganism transmission by hand. Methicillin-resistant Staphylococcus bacteria are among the main agents responsible for increasing healthcare-associated infections in Brazil and worldwide. Methods: The objective of this study was to screen and characterize methicillin-resistant Staphylococcus spp. on surfaces near patients in an intensive care unit. Microbiological samples, collected from ten beds in an intensive care unit with five sampling sites, were inoculated into a methicillin-resistant Staphylococcus aureus chromogenic medium. MALDI-TOF and PCR analyses were used to identify the bacteria. Antimicrobial susceptibility was determined using the disk diffusion test. The presence of the mecA gene was investigated using PCR. Results: We observed that 44 out of the 50 sampling sites presented grown isolates in the methicillin-resistant Staphylococcus aureus medium. The incidence of isolated microorganisms on the right side rail, left side rail, tables, infusion pump keypad, and cardiac monitor were 18.8 %, 36.7 %, 10.9 %, 2.4 %, and 31 %, respectively. The 42 isolates included in this study were identified as coagulase-negative Staphylococcus. All of these microorganisms were multidrug-resistant and mecA gene-positive. Conclusions: This study identified the presence of methicillin-resistant coagulase-negative Staphylococcus on the beds of an intensive care unit, providing evidence for the necessity of assertive actions to decrease the risk of healthcare-associated infections at the site.

Keywords: Coagulase-negative Staphylococcus. Equipment contamination. Methicillin resistance.

INTRODUCTION

Healthcare-associated infections (HAIs) are infections that patients acquire while receiving healthcare1. HAIs include cross-infection among patients infected or colonized with pathogenic bacteria, which can be transmitted from patient to patient directly or indirectly through vomit, devices, healthcare professionals, companions, and the environment around the patients2. HAIs have an impact on morbidity, mortality, and length of patient stay in nosocomial units, leading to increased mean costs of hospitalization3. The environment surrounding the patients is an important source of HAI microorganisms. Surfaces such as side rails, medical equipment, support tables, and curtains, among others, can become reservoirs of harmful microorganisms that can be transmitted to patients either directly, when the patient comes into contact with the medium, or indirectly, by contamination of the hands and gloves of health professionals4. Surface cleaning and disinfection are extremely important to reduce the occurrence of microbial contamination and consequently the risk of HAIs. This cleaning process is highly complex and multidimensional, involving the physical friction of the surfaces to remove organic
and inorganic materials, and then using a disinfectant solution, as well as monitoring strategies to ensure proper hygiene procedures. Bacteria of the genus Staphylococcus are the main cause of HAIs; their high prevalence on the skin of patients facilitates infection after a medical procedure breaks the barrier, or may be associated with decreased immunity. Coagulase-negative Staphylococcus (CoNS) strains are the main etiological agents of a series of infectious processes acquired in a hospital environment. In 2018, CoNS were the main causative agents of catheter-associated bloodstream infection in ICUs in Brazil. The occurrence of these CoNS-related HAIs has several predisposing factors, including the immune and health status of the patient and the environment close to the patient, which can be a source of both origin and transmission. Another characteristic that makes CoNS-related HAIs worrisome is their increased resistance to beta-lactam antibiotics, especially methicillin.

Resistance mechanisms against this class of antibiotics, developed by bacteria of the genus Staphylococcus spp., include two specificities that are important for their prevalence: the production of the beta-lactamase enzyme encoded by the blaZ gene, and the modified effector mechanism of beta-lactam antibiotics with the modification of the PBP binding protein into PBP2A protein, which has a low affinity for penicillin-binding, encoded by the mecA gene and its homologs, including mecALGA25. As the main human skin colonizer with a high survival capacity in inert, inhospitable environments with low substrate levels, CoNS often colonize the environment near the patients. They can be differentiated from S. aureus at the species level through PCR identification of the femA gene (an essential factor for methicillin resistance). This gene encodes a 48–50 kDa protein recognized as a specific factor and is contained in the chromosomes of these pathogens. Thus, the objective of this study was to screen and characterize methicillin-resistant Staphylococcus spp. on surfaces near patients in the ICU of a hospital in southeastern Brazil.

METHODS

Microorganism sampling and collection sites

The samples included in this study were composed of microorganisms isolated from hospital ICU bed surfaces, not involving patients (or their data) or any approach to health professionals. The sample collection was authorized by the hospital where the research was conducted.

The study was conducted in a general tertiary-level hospital located in the southeast region of Brazil. It has 397 beds and 3 ICUs, of which the general ICU, containing 10 beds, was chosen for sample collection. The rationale for choosing the general ICU was based on the fact that this environment receives patients from different hospital sectors and with different clinical conditions, increasing the variability of their microbiota, with a greater chance of analyzing the real epidemiological profile of the institution. Each of the 10 ICU beds included 5 sampling sites: right side rail (RS), left side rail (LS), bedside table (BT), infusion pump keypad (PK), and cardiac monitor (CM) (Figure 1). These points were chosen based on the frequency with which they are manipulated/touched, acting as potential contamination sources for healthcare professionals and patients. Surface sampling was performed around ICU bed surfaces before any cleaning procedure was performed. The sampling procedure was performed with patients inside the room. The samples were collected on October 24, 2019, using swabs immersed in saline solution for sampling a 5 cm² area. Subsequently, the swabs were depleted on Petri dishes containing a methicillin-resistant Staphylococcus aureus (MRSA) chromogenic growth medium (Probac do Brasil, Brazil). The plates were incubated at 37°C for 24 h. After the incubation period, colony-forming units (CFUs) were counted and registered on a form.

Microorganism identification

Sampling sites where no growth could be observed were excluded from the study. A total of 42 colonies from different sampling sites were analyzed using the Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry (MALDI-TOF MS) proteomic identification approach in the AQUACEN laboratory of the Veterinary School of the Federal University of Minas Gerais (UFMG), according to the methodology previously described by Assis et al., using a Microflex TM MALDI-TOF MS device (Bruker Daltonics, USA). Furthermore, the genomic PCR method was used to identify S. aureus at the species level through the detection of the femA marker gene, as described by Xavier et al.

Genomic analyses

The bacteria were selected on plates containing MRSA medium, transferred to Brain Heart Infusion (BHI) broth (Laborclin, Brazil), inoculated onto mannitol salt agar (Laborclin, Brazil) plates, and incubated at 37°C for 24 h. The growing colonies were analyzed using the DNA extraction procedure described by Gu et al. The integrity and quantification of the extracted DNA were verified by 1.0 % (w/v) agarose gel electrophoresis. This material was used for PCR. All primers used in this study were synthesized by GenOne Biotechnologies, Brazil.

PCR tests were used for femA gene detection, a species-specific marker for S. aureus, and mecA gene detection, a specific marker for
methicillin resistance, as described by Xavier et al. The reactions were performed using a mix containing 2× Go Taq Green Master Mix® (Promega Corporation, USA), 10 μM of each primer, and 50 ng of DNA, with a final reaction volume of 50 μL. PCR was performed in a Veriti Thermal Cycler (Applied Biosystems, USA). The PCR thermal conditions were described by Xavier et al. The amplicons were visualized on 1.5 % (w/v) agarose gel stained with ethidium bromide and photo-documented. S. aureus ATCC 43300, S. aureus ATCC 29213, and S. aureus 25923 strains were used as controls for PCR.

**Antimicrobial sensitivity profile**

The susceptibility to beta-lactam antibiotics was determined by the disk diffusion test according to the Clinical & Laboratory Standards Institute (CLSI) guidelines, using the following antimicrobials (Laborclin): oxacillin, 1 μg (OXA); erythromycin, 15 μg (ERI); cefoxitin, 30 μg (CFO); cefazolin, 30 μg (CFZ); cefuroxime, 30 μg (CRX); ciprofloxacin, 5 μg (CIP); sulfatrim, 25 μg (SUT); amikacin, 30 μg (AMI); clindamycin, 2 μg (CLI); cephalothin, 30 μg (CFL); and gentamicin, 10 μg (GEN).

**RESULTS**

This study demonstrated environmental surface contamination through the collection of microbiological samples from 50 surfaces near patients in an ICU. We observed that 44 out of the 50 samples grew on MRSA medium-containing plates. Although they grew on MRSA medium, two samples did not grow in mannitol salt agar and were therefore excluded.

Beds 4, 5, 6, 7, 8, and 9 presented contamination levels higher than 300 CFUs (Figure 2). The overall incidence of microorganisms in samples collected from RS, LS, BT, PK, and CM were 18.8 %, 36.7 %, 10.9 %, 2.4 %, and 31 %, respectively. This result revealed the dissemination of phenotypically methicillin-resistant strains on the ten sampling beds.

Proteomic genus identification confirmed that all isolates were Staphylococcus bacteria; however, species identification was only possible for 45.22 % (19/42) of the isolates. The species isolated from the surface samples of this study were all CoNS. The microorganisms identified were S. haemolyticus (11), S. epidermidis (4), S. capitis (2), and S. cohnii (1) (Table 1).

Due to the low species identification index obtained by MALDI-TOF, PCR was used to trace the femA gene, a species-specific marker for S. aureus, in all isolates. Control S. aureus strains were used to standardize the PCR for positive femA gene identification (Figure 3A). None of the isolates were femA gene-positive (Table 1), confirming the species identified using the MALDI-TOF and excluding the possibility that the strains could be identified as S. aureus.

Although the isolated strains were not S. aureus, they all grew on MRSA medium-containing plates, and their resistance to methicillin was tested at the genomic level by PCR screening for the mecA gene (Figure 2). Control S. aureus strains were used to standardize the PCR to screen for mecA gene-positive samples (Figure 3A). Among all the isolates, 90.47 % (38/42) were mecA gene-positive, confirming the phenotypic growth result in MRSA medium, even if they were not S. aureus strains. Figure 3B shows the mecA gene PCR results of representative isolates.

The antimicrobial susceptibility test showed the presence of extensively drug-resistant strains (Table 1). Among the 42 isolates, 100 % (42/42) were resistant to oxacillin, 98 % (41/42) to erythromycin, 100 % (42/42) to cefoxitin, 60 % (25/42) to cefazolin, 81 % (34/42) to ceftriaxone, 50 % (21/42) to ciprofloxacin, 74 % (31/42) to trimethoprim-sulfamethoxazole, 57 % (24/42) to amikacin, 98 % (41/42) to clindamycin, and 86 % (36/42) to gentamicin (Table 1). After the microorganisms were separated by species as identified by MALDI-TOF, the resistance to the tested antibiotics was Staphylococcus spp. (microorganisms classified only
The presence of bacterial contamination on inanimate surfaces in a hospital environment has attracted the interest of healthcare professionals in order to relate these microorganisms to HAIs. The presence of bacterial contamination on inanimate surfaces in a hospital environment has attracted the interest of healthcare professionals in order to relate these microorganisms to HAIs. The presence of bacterial contamination on inanimate surfaces in a hospital environment has attracted the interest of healthcare professionals in order to relate these microorganisms to HAIs. The presence of bacterial contamination on inanimate surfaces in a hospital environment has attracted the interest of healthcare professionals in order to relate these microorganisms to HAIs. The presence of bacterial contamination on inanimate surfaces in a hospital environment has attracted the interest of healthcare professionals in order to relate these microorganisms to HAIs.

### DISCUSSION

Gram-positive bacteria have been researched as they are more prevalent in these environments, especially in ICUs. Most of the studies on this subject show environmental contamination by *Staphylococcus aureus*.

CoNS represent a group of microorganisms that are the main definitive or transitory colonizers of human skin and mucous membranes. However, as they live in balance with the human skin and mucous membranes, they play a significant role in the transmission of infections. The presence of bacterial contamination on inanimate surfaces in a hospital environment has attracted the interest of healthcare professionals in order to relate these microorganisms to HAIs.
FIGURE 3: Result of PCR analysis to detect femA and mecA genes among Staphylococcus isolated from different surfaces of an ICU bed. Panel A. Standardization of the PCR for detection of femA and mecA genes in standard strains of Staphylococcus aureus. Line 1: Mid-Range DNA Ladder molecular weight marker (Cellco Biotechnology); Lines 2 to 4: Amplification of the femA gene in standard strains of S. aureus ATCC 43300, S. aureus ATCC 29230, and S. aureus 25923. Lines 5 and 6: negative and positive controls, respectively, for the PCR of the mecA gene in strains of S. aureus ATCC 29230 and S. aureus ATCC 43300. Panel B. Result of PCR analysis for detection of the mecA gene among the isolated Staphylococcus. Line 1: Mid-Range DNA Ladder molecular weight marker (Cellco Biotechnology); Lines 2 to 12: Amplification of the mecA gene in isolates 32 to 42.

TABLE 2: Antimicrobial resistance profile tested for each species identified by MALDI-TOF.

<table>
<thead>
<tr>
<th>Species</th>
<th>Quantity</th>
<th>OXA</th>
<th>ERI</th>
<th>CFO</th>
<th>CFZ</th>
<th>CRX</th>
<th>CIP</th>
<th>SUT</th>
<th>AMI</th>
<th>CLI</th>
<th>CFL</th>
<th>GEN</th>
<th>Total Resistant</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staphylococcus spp</td>
<td>23 (55%)</td>
<td>23</td>
<td>23</td>
<td>23</td>
<td>10</td>
<td>17</td>
<td>7</td>
<td>16</td>
<td>15</td>
<td>5</td>
<td>21</td>
<td></td>
<td>182</td>
</tr>
<tr>
<td>Staphylococcus haemolyticus</td>
<td>11 (26.2%)</td>
<td>11</td>
<td>10</td>
<td>11</td>
<td>10</td>
<td>11</td>
<td>9</td>
<td>10</td>
<td>3</td>
<td>11</td>
<td>9</td>
<td>10</td>
<td>105</td>
</tr>
<tr>
<td>Staphylococcus epidermidis</td>
<td>4 (9.5%)</td>
<td>4</td>
<td>4</td>
<td>4</td>
<td>1</td>
<td>3</td>
<td>3</td>
<td>2</td>
<td>4</td>
<td>4</td>
<td>0</td>
<td>3</td>
<td>32</td>
</tr>
<tr>
<td>Staphylococcus captis</td>
<td>2 (4.8%)</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>18</td>
</tr>
<tr>
<td>Staphylococcus hominis</td>
<td>1 (2.4%)</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>7</td>
</tr>
<tr>
<td>Staphylococcus cohnii</td>
<td>1 (2.4%)</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>8</td>
</tr>
<tr>
<td>Total resistant</td>
<td>42</td>
<td>41</td>
<td>42</td>
<td>25</td>
<td>34</td>
<td>21</td>
<td>31</td>
<td>24</td>
<td>41</td>
<td>15</td>
<td>36</td>
<td></td>
<td>352</td>
</tr>
</tbody>
</table>

OXA: Oxacillin; ERI: Erythromycin; CFO: Cefoxitin; CFZ: Cefazolin; CRX: Cefuroxime; CIP: Ciprofloxacin; SUT: Sulfitram; AMI: Amikacin; CLI: Clindamycin; CFL: Cephalothin; GEN: Gentamicin.

microbiota, for some time the CoNS were considered simple contaminants of biological samples and often underestimated, unlike S. aureus. CoNS are considered pathogens of great relevance to hospital environments, both for their ability to cause infections and to develop antibiotic resistance. They have been considered especially important in ICUs due to their high transmissibility by the hands of healthcare professionals or by indirect contact with contaminated equipment and surfaces. Species relevant to humans include S. epidermidis, S. capitis, S. warneri, S. haemolyticus, S. hominis, S. saccharolyticus, S. caprae, S. pasteuri, S. saprophyticus, S. xylosus, S. cohnii, S. simulans, S. auricularis, S. lugdunensis, and S. schleiferi. These microorganisms are considered opportunistic agents, and S. epidermidis is the most associated with HAI.

This study analyzed 50 samples collected in the ICU of a general hospital. Each of the 10 beds had 5 collection sites chosen according to the frequency they are handled by the multidisciplinary team; therefore, they are considered potential sources of contamination. Of the 50 samples collected, 42 (84 %) showed bacterial growth due to the presence of microorganisms that commonly colonize hands, which spread CoNS on the equipment and surfaces near the patient. In this work, the sampled surfaces were not subjected to any disinfection process prior to sampling collection, since the objective was to track the presence or absence of Staphylococcus bacteria. Of the equipment and surfaces analyzed, LS and CM were the most contaminated. A study by Moraes et al. also reported high contamination rates in infusion pumps and, as in our work, it was not possible to determine the contamination source. Microorganisms could have been carried to the LS and RS by the patients or healthcare professionals’ handling. Studies concerning microorganism phylogenetic approaches could be useful in elucidating the origin of contamination sources. For this purpose,
patients’ and healthcare professional’s hands sampling should be investigated and their results should be compared with those of hospital ICU surfaces analyses.

MALDI-TOF MS analysis was performed in a second step for genus identification. Of the samples, 55 % (23/42) were identified as *Staphylococcus* spp. and the other 45 % (19/42) were identified as: 26.2 % (11/42) *Staphylococcus haemolyticus*, 9.5 % (4/42) *Staphylococcus epidermidis*, 4.8 % (2/42) *Staphylococcus capitis*, 2.4 % (1/42) *Staphylococcus hominis*, and 2.4 % (1/42) *Staphylococcus cohnii*. The results showed the diversity of CoNS species and similar results were obtained in a study by Bernardi and Pizzolitto, in which the main microorganisms identified were *S. epidermidis* and *S. haemolyticus*, different only by the fact that *S. epidermidis* had the highest prevalence in this study.

PCR analysis of the femA gene showed no presence of *Staphylococcus aureus* in any of the samples, confirming the above result, which did not identify the presence of *S. aureus*. The mecA gene was present in 95 % (40/42) of the samples. It was absent in only 3 *S. haemolyticus* samples and 1 *S. epidermidis* sample. Oxacillin resistance without the mecA gene may occur for other reasons, such as beta-lactamase enzyme production.

The phenotypic analysis of antimicrobial resistance included 11 antimicrobials from 5 classes: 5 beta-lactams (oxacillin, cefoxitin, ceftazidime, cefuroxime, and cephalothin), 2 aminoglycosides (amikacin and gentamicin), 1 macrolide (erythromycin), 1 quinolone (ciprofloxacin), 1 sulfonamide (sulfatrim), and 1 lincosamide (clindamycin). Resistance rates were 100 % (42/42) for oxacillin and cefoxitin, 98 % (41/42) for clindamycin and erythromycin, 86 % (36/42) for gentamicin, 81 % (36/42) for cefuroxime, 74 % (31/42) for sulfatrim, 60 % (25/42) for ceftazidime, 98 % (41/42) for clindamycin and erythromycin, 57 % (24/42) for amikacin, and 36 % (41/42) for cephapirin. According to Tavares, resistance of CoNS to antimicrobials has been increasing, and oxacillin resistance is often associated with resistance to macrolides, aminoglycosides, tetracyclines, mupirocin, and clotrimazole because of their similar biochemical mechanisms.

All samples showed resistance to 3 or more classes of antimicrobials, and 16 samples showed resistance to at least 10 of the 11 antimicrobials tested. This high resistance rate corroborates the literature regarding the high capacity of CoNS to develop antimicrobial resistance.

These results show that the equipment and surfaces analyzed are CoNS reservoirs and thus should be considered effective transmission and dissemination means of HAIs in the analyzed ICU. Our work was limited to investigating only the role of some ICU surfaces considered potential sources for transmission and dissemination of HAIs. It could be considered a study limitation that could have an impact on public healthcare. This makes it even more important for hospitals to strive for excellence when it comes to infection control in order to protect workers and patients as well as their family members. Hand hygiene combined with the correct decontamination procedure of surfaces and equipment is fundamental to control the spread of microorganisms and HAIs.

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**AUTHORS’ CONTRIBUTION**

LFF, ACA, MASX and AREOX designed the study. GHSC, HFSD and WSS collected the samples. GÁADS, TPPP and LFF performed the experiments. AREOX and LFF performed data analysis and wrote the manuscript. All authors edited, read, and approved the final manuscript.

**CONFLICTS OF INTEREST**

The authors declare that they have no conflicts of interest.

**REFERENCES**


### Erratum

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**Title:** Identification and characterization of methicillin-resistant *Staphylococcus* spp. isolated from surfaces near patients in an intensive care unit of a hospital in southeastern Brazil


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Anna Christina de Ameida

Should read:

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