

Note

**ANTIBIOTIC RESISTENCE OF *Aeromonas hydrophila*
ISOLATED FROM *Piaractus mesopotamicus* (Holmberg, 1887)
AND *Oreochromis niloticus* (Linnaeus, 1758)**

Andréa Belém-Costa¹; José Eurico Possebon Cyrino^{2*}

¹UFAM - Centro de Apoio Multidisciplinar - Divisão de Biotecnologia, Av. Rodrigo Otávio, 3000 - 69077-900 - Manaus, AM - Brasil.

²USP/ESALQ - Depto. de Zootecnia - Setor de Piscicultura, C.P. 09 - 13418-900 - Piracicaba, SP - Brasil.

*Corresponding author <jepcyrin@esalq.usp.br>

ABSTRACT: One of the most important problems involving treatments with antibiotics against *Aeromonas hydrophila* isolated from fishes is that antibiotic resistance develops readily. The antimicrobial activity of chemotherapeuticants in isolates from pacu *Piaractus mesopotamicus* (Holmberg, 1887) and tilapia *Oreochromis niloticus* (Linnaeus, 1758) was tested by the Kirby-Bauer disk method, over Mueller-Hinton surface agar previously inoculated with 100 µL of bacterial suspensions. After regular incubation, isolates from tilapia and pacu were uniformly resistant to amoxicillin, ampicillin, lincomycin, novobiocin, oxacillin, penicillin, and trimetoprim+sulfametoazole. The *A. hydrophila* type strain presented resistance to the same antimicrobial substances and also against rifampicin; the bacterial isolate from pacu were the only strain resistant to tetracycline. Isolates from both pacu and tilapia had intermediate reaction with erytromycin. The use of drugs in commercial fish farms in Brazil can favor the development of resistant bacterial strains in native fish species as already observed for exotic species, commercially produced for longer time.

Key words: bacterial isolates, bacterial strains, antimicrobial substances, chemotherapeuticants, drugs

RESISTÊNCIA DE *Aeromonas hydrophila* ISOLADA DE *Piaractus mesopotamicus* (Holmberg, 1887) E *Oreochromis niloticus* (Linnaeus, 1758) A ANTIBIÓTICOS

RESUMO: Um dos maiores problemas envolvendo o tratamento com antibióticos contra *Aeromonas hydrophila* isolada de peixes confinados é a rápida resistência ao antibiótico desenvolvida pela bactéria. A atividade antimicrobiana de quimioterapêuticos em isolados a partir de pacu *Piaractus mesopotamicus* (Holmberg, 1887) e tilápia *Oreochromis niloticus* (Linnaeus, 1758) foi verificada pelo método de difusão de antibiótico em discos de Kirby-Bauer, sobre uma superfície de Agar Mueller-Hinton previamente inoculada com 100 µL de suspensão bacteriana. Após o período de incubação, os isolados de tilápia e pacu foram uniformemente resistentes a amoxicilina, ampicilina, lincomicina, novobiocina, oxacilina, penicilina e trimetoprim+sulfametoazol. A cepa tipo para *A. hydrophila* apresentou resistência às mesmas substâncias antimicrobianas e também contra a rifampicina; o isolado bacteriano de pacu foi a única linhagem resistente a tetraciclina. Isolados de pacu e tilápia apresentaram resistência intermediária frente a eritromicina. O uso de medicamentos em pisciculturas comerciais no Brasil pode favorecer o desenvolvimento de linhagens bacterianas resistentes em espécies de peixes nativos, como já observado em espécies exóticas comercialmente produzidas há mais tempo.

Palavras-chave: isolados bacterianos, cepas bacterianas, antimicrobianas, quimioterapêuticos, drogas

INTRODUCTION

Intensification of fish farming in Brazil has increased the number of disease outbreaks in intensive production systems. Most etiological agents were not yet identified and their morbid processes still not studied. The *Aeromonas* complex are important fish patho-

genic bacteria, causing septicemic infections and associated economic losses in fish culture worldwide (Holliman, 1993).

Multiple antibiotic resistance (MAR) has been registered for *Aeromonas hydrophila* isolated from freshwater fish farms in association with a variety of drugs, commonly used as feed additives (Aoki et al.,

1971; Pettibone et al., 1996; Son et al., 1997; Vivekanandhan et al., 2002). The main problem involving the use of antibiotics against *Aeromonas* infections is the development of resistance by these bacteria (Aoki & Egusa, 1971; Mitchell & Plumb, 1980), generally related to the presence of plasmids (Chang & Bolton, 1987; Ansary et al., 1992).

This work aims to identify antibiotic-resistant *Aeromonas* strains isolated from pacu *Piaractus mesopotamicus* and Nile tilapia *Oreochromis niloticus*, in intensive farming systems in South East Brazil.

MATERIAL AND METHODS

The antimicrobial activity of *A. hydrophila* isolates from pacu (LQUSP22) and tilapia (LQUSP15), collected during disease outbreaks in fish farms in Piracicaba (22°41'15"S; 47°41'15"W) and Jaú (22°18'45"S; 48°33'45"W), State of São Paulo, Brazil, and of the *A. hydrophila* type strain (ATCC7966), from the American Type Culture Collection, were compared by growth inhibition by the disk-agar diffusion method on Mueller-Hinton Agar (Merck®), as recommended by Stukus (1997). Both isolates were identified through biochemical methods and preserved in liquid nitrogen until use.

The following commercial disks antimicrobial substances used were: amoxicillin (25 µg); ampicillin (10 µg); chloramphenicol (30 µg); erythromycin (15 µg); gentamicin (10 µg); kanamycin (30 µg); lincomycin (2 µg); nitrofurantoin (300 µg); norfloxacin (10 µg);

novobiocin (30 µg); oxacillin (1 µg); penicillin G (10 U = 6 µg); rifampicin (5 µg); streptomycin (10 µg); sulbactam (10 µg) + ampicillin (10 µg); tetracycline (30 µg); and trimethoprim (1.25 µg) + sulphamethoxazole (23.75 µg). All tested antibiotics are ordinarily used in human medicine (Rhodes et al., 2000; MacMillan, 2001; Holmström et al., 2003). The commercial disks carrying small volumes of highly concentrated drugs provide a method of rapid diagnostic in which antibiotics serve as selective agents (Holt & Krieg, 1994).

Disks were spreaded over the surface of the agar, previously inoculated with 100 µL of suspensions of the test organisms prepared in Yeast Extract broth at 28°C for 24h (Costa et al., 1998). After overnight incubation at 28°C, the zone of inhibition was measured (mm), and calculations of zone diameter breakpoints, corresponding to limits of NCCLS - National Committee for Clinical Laboratory Standards (Cooper, 1955; Kronvall, 1982), were done.

RESULTS AND DISCUSSION

The isolates from tilapia and pacu were all resistant to amoxicillin, ampicillin, lincomycin, novobiocin, oxacillin, penicillin, and trimetoprim + sulfamethoxazole. The *A. hydrophila* type strain, presented resistance to the same antimicrobial substances, and also against rifampicin; the bacterial isolates from pacu were also resistant to tetracycline. The bacterial isolate from pacu and the *A. hydrophila* type strain had intermediate reaction with erythromycin (Table 1).

Table 1 - Susceptibility/Resistance profiles of bacteria isolated from fish and *A. hydrophila* (ATCC7966) type strain used in the present study. R-resistant; S-susceptible; I-intermediate.

ANTIBIOTICS	LQUSP22 (PACU)	LQUSP15 (TILAPIA)	ATCC7966
amoxicillin, 25 µg	R	R	R
ampicillin, 10 µg	R	R	R
chloramphenicol, 30 µg	S	S	S
erythromycin, 15 µg	I	S	I
gentamicin, 10 µg	S	S	S
kanamycin, 30 µg	S	S	S
lincomycin, 2 µg	R	R	R
nitrofurantoin, 300 µg	S	S	S
norfloxacin, 10 µg	S	S	S
novobiocin, 30 µg	R	R	R
oxacillin, 1 µg	R	R	R
penicillin G, 6 µg	R	R	R
rifampicin, 5 µg	S	S	R
streptomycin, 10 µg	S	S	S
sulbactam + ampicillin, 10 µg + 10 µg	R	R	R
tetracycline, 30 µg	R	S	S
trimethoprim + sulphamethoxazole, 1.25 µg + 23.75 µg	S	S	S

LQUSP = Luiz de Queiroz, Universidade de São Paulo.

The motile *Aeromonas* group is responsible for severe fish diseases, such as hemorrhagic septicemia (Bullock et al., 1971; Egusa, 1978; Schäperclaus et al., 1992) and ulcerative disease of fish (Karunasagar et al., 1995). The worldwide distribution of this group is a consequence of its great adaptability to different aquatic environments (Mateos et al., 1993). Infections by *Aeromonas* had been observed in various freshwater fish species and eventually in marine fish, reptile, bovine, and man (Bullock et al., 1971; Khordori & Fainstein, 1988). MAR has been recorded for several *A. hydrophila* strains around the world (Pettibone et al., 1996; Son et al., 1997). However, cumulative effects of chemotherapeuticants on the environment and on the bacterial ecology of farmed fish have received little attention (DePaola et al., 1995).

The extensive use of antibiotics and other chemotherapeuticants in fish farms, either to prevent or cure fish diseases, both as feed additives or dissolved directly into the water, has resulted in increasing antibiotic resistance among pathogenic bacteria. The bacterial isolate from pacu was resistant to eight out of the seventeen antibiotics tested, and the isolated from tilapia presented resistance against seven antimicrobial substances. The reference strain showed resistance to seven of eight antimicrobial substances to which the isolate of the Brazilian-native pacu was also resistant, and also to rifampicin.

The isolate from pacu was the only strain that presented resistance against tetracycline. Apparently, the most common antibiotic, indiscriminately used by Brazilian fish farmers, have already given rise to resistant bacterial strains affecting native fish species which have been produced commercially for no longer than a decade. Therefore, controlled, in-depth studies are needed to determine the effect of antimicrobial therapy on the microbial ecology of cultured fishes in Brazil.

According to Adams et al. (1998) and Rhodes et al. (2000), the aquaculture is related to the presence of bacterial resistant strains in a manner that interactive relationships between two different compartments, like hospitals and fish farms, can be though off. Based on the plasmid distribution into the *Aeromonas* group, these places behave in a reciprocal way. It is therefore important to investigate the use of antibiotics in aquaculture, the associated resistance to other relevant bacteria, and broaden the research to other geographic regions and other fish pathogens.

Because of the negative economic impact of disease outbreaks, and the fact that infections by *A. hydrophila* are most significant in warm water fisheries and aquaculture, it would be very important to study

the virulence of *A. hydrophila*, their resistant plasmid transference, and hability to adhere and infect other Brazilian-native fish. Understanding the role of different motile *Aeromonas* species in tropical fish disease will provide information leading to adequate prophylactic practices, and development of new drugs to be used in the Brazilian aquaculture.

ACKNOWLEDGEMENT

To Fundação de Amparo à Pesquisa do Estado de São Paulo (Sao Paulo State Research Foundation - FAPESP) for the grant-in-aid (99/12892-4) and the scholarship to Dr. A.B. Costa (99/00414-0).

REFERENCES

- ADAMS, C.A.; AUSTIN, B.; MEADEN, P.G.; McINTOSH, D. Molecular characterization of plasmid-mediated oxytetracycline resistance in *Aeromonas salmonicida*. *Applied and Environmental Microbiology*, v.64, p.4194-4201, 1998.
- ANSARY, A.; HANEEF, R.M.; TORRES, J.L.; YADAV, M. Plasmids and antibiotic resistance in *A. hydrophila*. *Journal of Fish Biology*, v.15, p.191-196, 1992.
- AOKI, T.; EGUSA, S. Drug sensitivity of *Aeromonas liquefaciens* isolated from freshwater fishes. *Bulletin of the Japanese Society for Scientific Fisheries*, v.37, p.176-185, 1971.
- AOKI, T.; EGUSA, S.; OGATA, Y.; WATANABE, T. Detection of resistance factors in fish pathogen *Aeromonas liquefaciens*. *Journal of General Microbiology*, v.65, p.343-349, 1971.
- BULLOCK, G.L.; CONROY, D.A.; SNIESKO, S.F. Septicemic diseases caused by motile aeromonads and pseudomonads. In: SNIESKO, S.F.; AXELROD, H.R. (Ed.) *Diseases of fishes*. Neptune: T.F.H. Publ., 1971. v.2A, p.21-41: Bacterial diseases of fishes.
- CHANG, B.J.; BOLTON, S.M. Adherence haemagglutination and cell surface characteristics of motile aeromonads virulent for fish. *Journal of Fish Diseases*, v.13, p.255-268, 1987.
- COOPER, K.E. Theory of antibiotic inhibition zones in agar media. *Nature*, v.176, p.510-511, 1955.
- COSTA, A.B.; KANAI, K.; YOSHIKOSHI, K. Serological characterization of atypical strains of *Edwardsiella tarda* isolated from SeaBream. *Fish Pathology*, v.33, p.265-274, 1998.
- DEPAOLA, A.; PEELER, J.T.; RODRICK, G.E. Effect of oxytetracycline-medicated feed on antibiotic resistance of Gram-negative bacteria in catfish ponds. *Applied and Environmental Microbiology*, v.61, p.2335-2340, 1995.
- EGUSA, S. *Infectious diseases of fish*. Tokyo: Kouseisha Kouseikaku, 1978. 554p.
- HOLLIMAN, A. The veterinary approach to trout. In: BROWN, L. (Ed.). *Aquaculture for veterinarians*: fish husbandry and medicine. Oxford: Pergamon Press, 1993. cap.14, p.223-247.
- HOLMSTRÖM, K.; GRÄSLUND, S.; WAHLSTRÖM, A.; POUNGSHOMPOO, S.; BENGTSSON, B.; KAUTSKY, N. Antibiotic use in shrimp farming and implications for environmental impacts and human health. *International Journal of Food Science and Technology*, v.38, p.255, 2003.
- HOLT, J.G.; KRIEG, N.R. Enrichment and isolation. In: GERHARDT, P.; MURRAY, R.G.E.; WOOD, W.A.; KRIEG, N.R. *Methods for general and molecular bacteriology*. Washington: American Society for Microbiology, 1994. cap.8, p.179-215.
- KARUNASAGAR, I.; SUGUMAR, G.; KARUNASAGAR, I. Virulence characters of *Aeromonas* sp. isolated from EUS-affected fish. In: SHARIF, M.; ARTHUR J.R.; SUBASINGHE, R.P. (Ed.). *Diseases in Asian aquaculture II*. Manila: Asian Fisheries Society, Fish Health Section, 1995. p.307-314.

- KHARDORI, N.; FAINSTEIN, V. *Aeromonas* and *Plesiomonas* as etiological agents. **Annual Review of Microbiology**, v.42, p.395-419, 1988.
- KRONVALL, G. Analysis of a single reference strain for determination of gentamicin regression line constants and inhibition zone diameter breakpoints in quality control of disk diffusion antibiotic susceptibility testing. **Journal of Clinical Microbiology**, v.16, p.784-793, 1982.
- MACMILLAN, J.R. Aquaculture and antibiotic resistance: A negligible public health risk? **World Aquaculture**, v.32, p.49-50, 2001.
- MATEOS, D.; ANGUITA, J.; NAHARRO, G.; PANIAGUA, C. Influence of growth temperature on the production of extracellular virulence factors and pathogenicity of environmental and human strains of *Aeromonas hydrophila*. **Journal of Applied Bacteriology**, v.74, p.111-118, 1993.
- MITCHELL, A.J.; PLUMB, J.E. Toxicity and efficacy of Furanace on channel catfish infected experimentally with *Aeromonas hydrophila*. **Journal of Fish Diseases**, v.3, p.93-100, 1980.
- PETTIBONE, G.W.; MEAR, J.P.; SAMPSELL, B.M. Incidence of antibiotic and metal resistance and plasmid carriage in *Aeromonas* isolated from brown bullhead (*Ictalurus nebulosus*). **Letters in Applied Microbiology**, v.23, p.234-240, 1996.
- RHODES, G.; HUYS, G.; SWINGS, J.; McGANN, P.; HINEY, M.; SMIYH, P.; PICKUP, R.W. Distribution of oxytetracycline resistance plasmids between Aeromonads in hospital and aquaculture environments: Implication of Tn1721 in dissemination of the tetracycline resistance determinant Tet A. **Applied and Environmental Microbiology**, v.66, p.3883-3890, 2000.
- SCHÄPERCLAUS, W.; KULOW, H.; SCHRECKEBACH, K. Infectious abdominal dropsy. In: SCHÄPERCLAUS, W. (Ed.). **Fish diseases**. Berlin: AkademieVerlag, 1992. v.1, p.401-458.
- SON, R.; RUSUL, G.; SAHILAH, A.M.; ZAINURI, A.; RAHA, A.R.; SALMAH, I. Antibiotic resistance and plasmid profile of *Aeromonas hydrophila* isolates from cultured fish, Telapia (*Telapia mossambica*). **Letters in Applied Microbiology**, v.24, p.479-482, 1997.
- STUKUS, P.E. Antimicrobial testing. The Kirby-Bauer method (filter-paper disk method). In: STUKUS, P.E. (Ed.). **Investigating microbiology: a laboratory manual for general microbiology**. Orlando: Harcourt Brace & Company, 1997. cap.44, p.243-247.
- VIVEKANANDHAN, G.; SAVITHAMANI, K.; HATHA, A.A.M.; LAKSHMANAPERUMALSAMY, P. Antibiotic resistance of *Aeromonas hydrophila* isolated from marketed fish and prawn of South India. **International Journal of Food Microbiology**, v.76, p.165-168, 2002.

Received March 08, 2005

Accepted March 31, 2006