EFFECTS OF ROOT DEFORMATION AND LIGHT AVAILABILITY ON GROWTH AND BIOMASS ALLOCATION OF *Senna multijuga* seedlings (Rich) H. S. Irwin & Barneby

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ABSTRACT – The effects of root deformation caused by errors in the pricking-out process in forest nurseries are still unknown for tropical tree seedlings. We analyzed the effects of light availability and root deformation on growth and biomass allocation in seedlings of *Senna multijuga*, a pioneer tropical tree commonly used in forest restoration programs. Our hypotheses were: (a) as a typical light-demanding species, the seedlings of *S. multijuga* may have their growth compromised by low light availability; (b) root deformation impairs growth rates and induces changes in biomass allocation; and (c) the effects of low light availability on growth and biomass allocation are increased by root deformation. Seedlings with and without root deformation were cultivated for 43 days under three levels of total daily photosynthetically active radiation (PAR) (28, 12, and 1 mol photons m⁻² day⁻¹). Seedlings of *S. multijuga* had their growth rates severely affected by values of PAR at about 1 mol photons m⁻² day⁻¹, but root deformation did not affect the relative growth rates of the whole plant. Instead, root deformation caused a decrease in the relative growth rate of roots in all light availability and root deformation of biomass to leaves are more pronounced at low light availability. Root deformations may lead to the production of seedlings with a low competitiveness capacity regardless of light conditions.

Keywords: Brazilian Atlantic Rainforest; Pricking-out; Seedling quality.

EFEITOS DA DEFORMAÇÃO RADICULAR E DA DISPONIBILIDADE DE LUZ NO CRESCIMENTO E NA DISTRIBUIÇÃO DE BIOMASSA DE MUDAS DE Senna multijuga (Rich) H. S. Irwin & Barneby

RESUMO – Os efeitos e consequências da deformação da raiz causada por erros no processo de repicagem em viveiros florestais ainda são desconhecidos para as mudas de árvores tropicais. Analisamos os impactos da disponibilidade de luz e de deformação das raízes no crescimento e alocação de biomassa em mudas de **Senna multijuga**, uma árvore tropical pioneira comumente utilizada em programas de restauração florestal. Nossas hipóteses foram: (a) como espécie típica exigente de luz, as plântulas de **S. multijuga** podem ter seu crescimento comprometido em baixa disponibilidade de luz, (b) a deformação da raiz induz prejuízos nas taxas de crescimento e mudanças na alocação de biomassa, e (c) os efeitos da baixa disponibilidade de luz



Revista Árvore 2020;44:e4418 http://dx.doi.org/10.1590/1806-908820200000018 no crescimento e alocação de biomassa são potencializados pela deformação da raiz. As plântulas, com e sem deformação das raízes, foram cultivadas durante 43 dias sob três níveis de radiação fotossinteticamente ativa total (PAR) (28, 12 e 1 mol fótons m² dia⁻¹). Mudas de **S. multijuga** tiveram suas taxas de crescimento severamente afetadas nos valores de PAR em cerca de 1 mol fótons m² dia⁻¹, mas a deformação das raízes não afetou as taxas de crescimento relativo da planta inteira. Em vez disso, a deformação da raiz causou uma diminuição na taxa de crescimento relativo das raízes em todas as disponibilidades de luz. As mudanças no crescimento das raízes afetaram a alocação de biomassa para as raízes. Os efeitos da disponibilidade de luz e da deformação da raiz na alocação de biomassa para folhas são mais pronunciados a baixos valores de disponibilidade de luz. As deformações das raízes poderiam levar à produção de mudas com baixa capacidade de competitividade independentemente das condições de luz.

Palavras-Chave: Mata Atlântica; Repicagem; Qualidade de mudas.

1.INTRODUCTION

Forest degradation in the tropics has caused the loss of habitats for flora and fauna and has been promoted through anthropic activities, such as logging, hunting, and deforestation for agricultural expansion (Calegari et al., 2010; Giam, 2017). In this context, forest restoration represents an important way to mitigate the environmental problems caused by forest degradation (Lamb, 2012; Brancalion and Chazdon, 2017). One of the most important steps in the establishment of forest stands is to ensure quality seedling production at the nursery stage because this directly affects seedling establishment and growth in the field (Binotto et al., 2010; Grossnickle, 2016). Due to the initial fragility of tree seedlings, the seedling production process requires appropriate management strategies in the nursery to obtain a better uniformity of height and a well-formed root system because these qualities can promote satisfactory survival and growth in the field, even under adverse conditions (Gomes et al., 2002; Thomas et al., 2008).

Plants depend on their roots for soil anchorage and the absorption of water and nutrients (Lynch, 1995; Amoroso et al., 2010). Roots also need to be capable of acquiring resources efficiently and penetrating the soil, even under conditions of compaction and mechanical resistance (Bizet et al., 2016). Root deformations can be induced at three stages of seedling production in the nursery, that is, in the stages of pricking-out, growth, and harvesting. The deformation of roots in the prickingout process occurs when the hole made in the substrate to accommodate the roots is smaller than the root's total length. Although the use of containers with inadequate size and form and the time spent in the nursery can promote deformations in the root system, the pricking out process is crucial because errors during this process are

Revista Árvore 2020;44:e4418

irreversible (Carneiro, 1995; Jaenicke, 1999; Wendling et al., 2002; Gregorio et al., 2010; Wilkinson et al., 2014; Duboc, 2015). Root deformations reduce the absorption and translocation of water and nutrients, as deformation points can offer resistance to hydraulic conductivity and indirectly cause nutritional imbalance, compromising the establishment and competitiveness of field tree seedlings (Reis et al., 1991). Low competitiveness capacity may lead to the impairment of plant growth and mortality (Ortega et al., 2006), resulting in higher costs for the maintenance of forest stands with the replacement of new seedlings (Freitas et al., 2005). Root deformation caused at the moment of seedling pricking out is a problem in small- and medium-sized nurseries in the tropics. Although several nursery manuals draw attention to problems arising from root deformation at the time of pricking out (Carneiro, 1995; Jaenicke, 1999; Gregorio et al., 2010; Wilkinson et al., 2014), the topic has never been the focus of intensive experimental studies or the subject of experiments on the interactive effects of root deformation and environmental stress.

The opening and closing of gaps in the developing forest favor the formation of light gradients, making creating ideal conditions for the occurrence of a great diversity of plant species (Valladares et al., 2012). Plants of the early and intermediate stages of forest succession are typically light-demanding. Well-adapted to deal with high light intensities, they have a great capacity to survive and grow in gaps and forest edges (Bazzaz and Picket, 1980; Whitmore, 1989). Typically, pioneer tree species have mechanisms that enable them to tolerate excess light but exhibit limited ability to establish and grow under low light availability (Kitajima and Fenner, 2000). Pioneer tree species grow rapidly and are suitable for planting in projects to restore degraded forests (Montagnini, 2005).



Senna multijuga belongs to the Leguminosae family (Carvalho, 2004). In Brazil, this species is distributed in almost all regions of the country, especially in the Brazilian Atlantic rainforests (Lorenzi, 2002). It is classified as a pioneer species, demanding light and being indifferent to the physical conditions of the soil (Lorenzi, 2002). This species has high growth rates and is recommended for planting in mixed reforestation schemes for degraded areas (Lorenzi, 2002; Carvalho, 2004; Doust et al., 2006). The fact that *S. multijuga* has a short life cycle compared to other forest species, plus the ability to colonize poor soils rapidly and disperse its seeds by autochory, makes it suitable for use in forest restoration programs (Lacerda et al., 2004).

In this study, we analyzed the effects of light availability and root deformation on growth and biomass allocation in *S. multijuga seedlings*. Our hypotheses were as follows: (a) as a typical light-demanding species, the seedlings of *S. multijuga* may have their growth compromised by low light availability; (b) root deformation impairs growth rates and induces changes in biomass allocation; and (c) the effects of low light availability on growth and biomass allocation are increased by root deformation.

2.MATERIALS AND METHODS

Seeds of S. multijuga were collected by the Instituto Floresta Viva (IFV) from trees located in the district of Serra Grande, Uruçuca, Bahia, Brazil (14046'59"S, 39002'39"W). The experiment was installed in the nursery of the Universidade Estadual de Santa Cruz (UESC), Ilhéus, BA, Brazil (14045'15"S, 39o13'59"W). The UESC's nursery measures 20 \times 15 m and is covered with a shade cloth that allows the passage of approximately 60% of light radiation at full sun. Seeds were planted in plastic boxes containing washed sand. One month after germination, seedlings were pricked out to plastic bags with a capacity of 1.5 L. The soil used had been collected from a small area of forest located on the UESC campus. At the moment of pricking out, the roots of 50% of the seedings were deliberately deformed. The deformation of roots was carried out by bending the root towards the stem and forcing the fold to 180° (Figure 1). Also, the hole for root accommodation was approximately half the total length of the main root. After 21 days of acclimation, the plastic bags were placed inside PVC tubes of 100 mm diameter and 250 mm height and transferred to three different light availability conditions.





Figure 1 – Seedling of S. multijuga with root deformation. The arrow indicates where the deformation occurs. Source: Authors.
Figura 1 – Mudas de S. multijuga com deformação radicular. A seta indica onde ocorre a deformação. Fonte: Autores.

Seedlings without (R1) and with (R2) root deformation were distributed in three conditions of light availability, repeated three times each. Light conditions were obtained by constructing shaded houses with 1 m wide \times 1 m long \times 0.80 m high, covered with different layers of shade cloths. The shade houses were then arranged randomly on three wooden and metal benches with 1 m height and 4.0 x 1.5 m area, so as to have a replica of each light environment per bench. Throughout the experiment, the photosynthetically active radiation (PAR) was measured using S-LIA-M003 light sensors coupled to Hobo Micro Station Data Loggers (Onset Computer, Massachusetts, USA). The data loggers were programmed to perform readings at 1-second intervals and to store the mean PAR values every 10 min. During the experiment, the average daily total PAR was 28, 12, and 1 mol photons m⁻² day⁻¹, respectively, for the three light environments. Air temperature and relative humidity were monitored inside the nursery using a

Hobo Pro V2 Data Logger sensor (Onset Computer, Massachusetts, USA). The average daytime temperature was 27.5 °C and the average nighttime temperature was 23.8 °C, and the maximum and minimum temperatures were 31.6 °C and 23.1 °C, respectively. The average daily vapor pressure (DPV) deficit was 0.81 kPa.

At the beginning of the experiment, when plants were transferred to the light environment, (RDM,), stems (SDM₁), leaves (LDM₁) and total dry mass (TDM) were evaluated for 10 seedlings with root deformation and 10 seedlings without root deformation. Dry masses of roots, stems, and leaves were obtained by drying plant parts in an air-circulating oven at 50 °C until a constant dry mass was obtained. At 43 days after the beginning of the experiment, measures of plant height (H) and diameter (D) were measured. The plants were removed from the substrate, and the root volume (RV) was measured by the displaced water method using a 500 mL measuring beaker. The plants were then separated into roots, stems, and leaves for the determination of RDM₂, SDM₂, LDM₂, and TDM₂. From these values, the following variables were calculated:

root mass ratio $_{RMR_2} = \frac{RDM_1}{TDM_2}$, stem mass ratio $_{SMR_2} = \frac{SDM_2}{TDM_2}$, and leaf mass ratio $_{LMR_2} = \frac{LDM_1}{TDM_2}$, relative growth rate based on total plant dry mass $_{RGR_{i}} = \frac{\ln TDM_{i} - \ln TDM_{i}}{43}$, relative growth rate based on leaf mass, $_{RGR_{i}} = \frac{\ln LDM_{2} - \ln LDM_{i}}{43}$, relative growth rate based on real mass, $R_{GR_2} = \frac{43}{16SDM_2 - \ln SDM_1}$, and relative growth rate based on root dry mass $_{RGR_3} = \frac{\ln RDM_2 - \ln SDM_1}{43}$, where indices 1 and 2 correspond to the evaluations performed at the end and at the beginning of the experiment, respectively, and 43 days was the duration of the experiment. Based on data of H, D, RDM, and RV, we calculated the ratios of D/H, D/RV, and H/RV.

The experiment was conducted in a randomized block design, in a split-plot scheme, with and without root deformation and three light availabilities (28, 12 and 1 mol photons m⁻² day⁻¹) with four to five plants per experimental unit. The data were submitted to analysis of variance (ANOVA) followed by Tukey's test at a 5% significance level. All assumptions about the residuals were tested, and when there were violations, the Box-Cox transformation was used. Statistical analyses were carried out using R software (R Development Core Team, 2019) with the Tinn-R interface (Tinn-R Team, 2018).

3.RESULTS

For the variables H, D, RGR, RGR_s, RGR_p, SMR_y, RMR₂, D/H, D/RV, and H/RV, there were no significant

(R) deformation day ⁻¹ , L12 - 12	1 for young plants of S. mol photons m ⁻² day ⁻¹ , a	<i>multijuga</i> , without (R1) and L1 - 1 mol photons m	and with $(R2)$ root def ² day ⁻¹), for 43 days. Fo	ormation and under threat the variables presented	e light environments (L' l, there were no significa	28-28 mol pho nt interactions	tons m ⁻² oetween
root detormatio Tabela 1 – Valores médi deformação de m² dia ⁻¹ , L12 - deformação da:	n and ught. os e erros padrão das 1 huz (L) e raiz (R) para pi 12 mol fótons m² dia ¹ s raízes e huz.	variáveis de crescimentc lantas jovens de S. multi e LI - I mol fótons m² ^c	v e de alocação de bior uga , sem (R1) e com (R lia ¹), por 43 dias. Para	nassa e níveis de signij 2) deformação radiculai as variáveis apresenta	ficância de ANOVA ao c r e sob três ambientes de das, não houve interaçõ	comparar os ej luz (L28-28 m ss significativas	eitos de al fótons entre a
	Light (L)			Root (R)		ANO	VA
Variable	L28	L12	LI	RI	R2	L	×
H (cm)	$37.4 \pm 1.0 a$	40.31 ± 0.1 a	$25.20 \pm 1.0 \text{ b}$	35.1 ± 0.82 a	33.5 ± 0.82 a	* * *	su
D (mm)	$4.84 \pm 0.1 \mathrm{a}$	$4.73 \pm 0.1 \text{ b}$	$2.24 \pm 0.1 c$	$3.88 \pm 0.07 a$	$3.82 \pm 0.07 a$	* * *	su
RGR (mg g ⁻¹ day ⁻¹)	73.22 ± 1.2 a	68.25 ± 1.2 a	$28.64 \pm 1.2 \text{ b}$	$57.1 \pm 1.10 \text{ a}$	56.3 ± 1.11 a	* *	su
RGR _s (mg g ⁻¹ day ⁻¹)	76.30 ± 0.4 a	74.80 ± 0.3 a	$36.00 \pm 0.3 \text{ b}$	$58.60 \pm 0.31\mathbf{b}$	66.20 ± 0.33 a	* *	* *
RGR _R (mg g ⁻¹ day ⁻¹)	$74.20 \pm 0.2 \text{ a}$	$66.00\pm0.2~\mathrm{b}$	$16.20 \pm 0.1 \text{ c}$	56.60 ± 0.11 a	$45.90\pm0.16~\mathrm{b}$	* * *	* *
SMR ₂ (g g ⁻¹)	$0.19 \pm 0.01 \text{ a}$	0.23 ± 0.02 a	0.28 ± 0.04 a	0.20 ± 0.03 a	$0.27 \pm 0.06 a$	ns	su
RMR ₂ (g g ⁻¹)	$0.31 \pm 0.01 \text{ a}$	$0.24\pm0.01~\mathrm{b}$	$0.18 \pm 0.01 \text{ c}$	0.26 ± 0.01 a	$0.24 \pm 0.01 \text{ a}$	* *	su
D/H	0.13 ± 0.002 a	$0.11 \pm 0.002 \text{ b}$	$0.9 \pm 0.002 c$	$0.11 \pm 0.002 a$	$0.11 \pm 0.002 \text{ a}$	* *	ns
D/RV (mm mL ⁻¹)	$0.33 \pm 0.1 c$	$0.64 \pm 0.1 b$	$2.04\pm0.16a$	$1.13 \pm 0.08 a$	$0.08\pm0.08~\mathrm{b}$	* * *	* *
H/RV (cm mL ⁻¹)	$2.50\pm1.8~c$	$5.73 \pm 1.2 \text{ b}$	22.30 ± 1.8 a	11.53 ± 0.96 a	$8.82 \pm 0.96 \ b$	* *	* *

Table 1 – Mean values and standard errors of growth and biomass allocation variables and significance levels of ANOVA while comparing the effects of light (L) and root (R) deformation for young plants of *S. multijuga*, without (R1) and with (R2) root deformation and under three light environments (L28-28 mol photons m^2 day¹, L12 - 12 mol photons m^2 day¹, and L1 - 1 mol photons m^2 day¹, for 43 days. For the variables presented, there were no significant interactions between

effects of the interactions between light and root (p>0.05) (Table1). Seedlings of S. multijuga showed severely affected PAR values at about 1 mol photons m⁻² day⁻¹. With the exception of SMR₂, significant differences between light environments were found for all variables analyzed. For H, RGR, and RGR_s, non-significant differences were found between L28 and L12. The mean values of D, RGR_R, RMR₂, and D/H were significantly higher in L28 than in L12 and L1, whereas the mean values of D/RV and H/RV were significantly higher in L1 than in L28 and L12. The mean values of RGR_R were 2.6, 2.1 and 4.6 times higher in L28 than in L1. The mean values of D/RV and H/RV were respectively 6.2 and 8.9 times higher in L1 than in L28.

For RGR_L and LMR₂, significant interactions were verified between root condition and light environment (p < 0.05). The lowest averages were presented in the condition of lower light availability (L1). Plants without deformation presented 2.0 times higher values for RGR_L in L28 compared to L1 (Table 2), whereas plants with deformation were 3.0 times higher in L28 than in L1. On the other hand, the mean values of LMR₂ in plants without root deformation (R1) were significantly higher than in plants with root deformation (R2) in L1. In this lower light environment (L1), the mean values of LMR₂ in R1 were 1.3 times higher than in R2 (Table 2).

For biomass allocation, R1 and R2 had less root biomass in L1 (Figure 2). Nevertheless, the biomass allocation to the stems was much higher in R2 than in the other factorial combinations (roots x light availability). The highest biomass allocation to the leaves was observed in R1 in L1, while the highest biomass allocation to the roots was observed in L28, independent of root deformation. Plants without root deformation (R1) had approximately 70%, 72%, and 80% of biomass allocated to leaves and stems in L28, L12, and L1, respectively. In comparison, plants with root deformation (R2) had about 70%, 79%, and 90 % of biomass allocated to leaves and stems in L28, L12, and L1, respectively. The lowest biomass allocated to the roots was observed for R2 in L1.

4.DISCUSSION

The significant differences in H, RGR, and RGR_s of seedlings in L28 and L12 in comparison with seedlings in L1 indicate that S. multijuga is a typical light-demanding and shade-intolerant species, in accordance with its ecological classification (Lorenzi, 2002). Significant differences in RMR, in plants grown in L28 in comparison with L12 and L1 are related to the characteristics of acclimatization to low light availability. Plants grown under low light availability tend to allocate more biomass to stems and leaves than to the roots (Montgomery, 2004; Valladares et al., 2012). This tendency can be more easily observed in fastgrowing pioneer species, reinforcing the classification of S. multijuga as a light-demanding species. Increases in the biomass allocation to leaves in relation to stems and roots contribute to the maintenance of a positive carbon balance in counterparts with low photosynthetic capacity under low light availability (Givnish, 1988). In contrast, plants grown in environments with higher light intensity tend to have higher transpiration rates, and there is usually an increase in carbon allocation to the roots to increase the efficiency of water and nutrient absorption (Poorter, 1990; Zuidema et al., 1999; Poorter et al., 2012). Due to the high levels of irradiance, the air temperature rises, which can cause water restriction in the soil. As a strategy to escape water stress, plants

Table 2 – Mean values and standard errors of relative growth rate based on leaf dry mass (RGR₁) and leaf mass ratio (LMR₂) of *S. multijuga* seedlings, without (R1) and with (R2) root deformation and under three light levels (L28- 28 mol photons m² day⁻¹, L12 - 12 mol photons m² day⁻¹, and L1 - 1 mol photons m⁻² day⁻¹), at the end of 43 days.
Tablea 2 – Valores médios e erros padrão da taxa de crescimento relativo com base na massa seca das folhas (RGR₁) e da razão da massa

Tabela 2 – Valores médios e erros padrão da taxa de crescimento relativo com base na massa seca das folhas (RGR₁) e da razão da massa foliar (LMR₂) de mudas de S. multijuga, sem (R1) e com (R2) deformação das raízes e sob três níveis de luz (L28-28 mol fótons m² dia⁻¹, L12 - 12 mol fótons m² dia⁻¹, e L1 - 1 mol fótons m² dia⁻¹), ao final de 43 dias.

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Variable	Root	L28	L12	L1
RGR	R1	70.20 ± 0.2 Aa	69.70± 0.2 Aa	0.51 ± 0.02 Ba
2	R2	63.10 ± 0.2 Aa	67.70 ± 0.2 Aa	$0.48\pm\!0.03~\mathrm{Cb}$
LMR ₂	R1	$34.40\pm0.2\;\mathrm{Ba}$	23.10 ± 0.2 Ba	0.64 ± 0.03 Aa
	R2	0.51 ± 0.02 Ca	$0.57\pm0.03\mathrm{Aa}$	$0.52\pm0.03~\mathrm{Bb}$
M C 11			1 1 (10 05) C 11 1 1	1 1 4 11 14

Means followed by the same letter are not significantly different from each other by the Tukey test (p < 0.05). Capital letters represent comparisons between light environments within root conditions and lowercase letters represent comparisons between root conditions for each light environment. Médias seguidas da mesma letra não significativamente diferentes entre si pelo teste de Tukey (p < 0.05). Letras maiúsculas representam comparações entre si pelo teste de Tukey (p < 0.05). Letras maiúsculas representam comparações entre su pelo teste de Tukey (p < 0.05). Letras maiúsculas representam comparações entre su pelo teste de Tukey (p < 0.05). Letras maiúsculas representam comparações entre su pelo teste de Tukey (p < 0.05).

ambientes de luz dentro das condições de raiz e letras minúsculas representam comparações entre as condições raiz para cada ambiente de luz.





- Figure 2 Biomass allocation for leaves, stems and roots in seedlings of *S. multijuga*, without (R1) and with (R2) root deformation and subjected to three light levels (L28- 28 mol photons m⁻² day⁻¹, L12 -12 mol photons m⁻² day⁻¹, and L1-1 mol photons m⁻² day⁻¹) during 43 days.
- Figura 2 Álocação de biomassa para folhas, caules e raízes em mudas de S. multijuga, sem (R1) e com (R2) deformação da raiz e submetidos a três níveis de luz (L28-28 mol fótons m² dia⁻¹, L12 -12 mol fótons m² dia⁻¹, e L1-1 mol fótons m² dia⁻¹) durante 43 dias.

invest in root growth, seeking the deepest and most humid layers in the soil (Scalon et al., 2011). Plants of *S. multijuga* occur naturally in several regions of Brazil, including the dry forests of the northeastern Brazilian interior, which are characterized by pronounced climatic seasonality, with well-defined dry and rainy seasons (Carvalho, 2004). Therefore, the increase in the allocation of biomass to the roots in environments with higher light availability is a strategy to maintain the water balance because, at high solar radiation and high temperatures, plants tend to transpire more (Markesteijn and Poorter, 2009).

The low mean RGR_R values probably triggered low RGR₁ values, as the lower biomass allocation to the roots decreases water and nutrient uptake, affecting photosynthesis and the availability of carbon for leaf growth. In this case, the decrease in net photosynthetic rates is a common response of light-demanding trees to low light availability. In such cases, the leaves cannot adjust to the PAR available and cannot reach their maximum photosynthetic capacity (Feijó et al., 2009). Also, root deformation can increase resistance to water flows and nutrients. This low capacity for transporting carbohydrates to non-photosynthetic tissues and increasing resistance to water and nutrient flows from roots to leaves can cause significant changes in RGR_p in plants with root deformation regardless of the light condition in which they were grown. At the same

Revista Árvore 2020;44:e4418

time, the reduction in light availability also caused the reduction of RGR_{R} regardless of root conditions. In addition to the phloem transport of carbohydrates from the leaves to the roots, the supply of minerals from the root to the leaves also affects carbon assimilation and partitioning, influencing the ratio of root to shoot biomass (Agren and Ingestad, 1987). In fact, root and shoot growth depend on a closely coordinated balance (Farrar and Jones, 2000), and environmental factors, such as availability of water, minerals, and light, can modify the root/shoot biomass ratio (McDonald et al., 1986). Thus, endogenous and exogenous factors, such as root deformation and low light availability, can drastically affect leaf and root growth, as observed in the experiment in seedlings with root deformation in L1.

Seedlings with root deformation are less able to compete for water and nutrient uptake and have less soil anchorage capacity. For example, in degraded tropical pastures, the death of tree seedlings due to competition for light, water, and nutrients with fast-growing grasses is a constant risk factor (Resende and Leles, 2017). In tropical regions, degraded pastures are often covered with fast-growing grasses, such as species of the genus Brachiaria spp. and Urochloa decumbens, which can reach up to 1.5 m in height and are difficult to control (Weidlich, 2011; Ferreira et al., 2016). Therefore, in the first months after planting, intensive mechanical and chemical weeding around seedlings are necessary to avoid and control the competition for light, water, and nutrients with native and exotic spontaneous plants (César et al., 2014; Santos et al., 2018). Weeding represents a significant part of maintenance costs in forest restoration projects, and the procedures used to control the competition of tree seedlings with fastgrowing plants are sometimes inefficient and costly (Brancalion et al., 2009). Thus, the poor quality of tree seedlings caused by errors in the pricking-out process can lead to increased mortality rates in the field, further increasing the costs of forest restoration.

Studies that investigate the impact of root deformation caused during the pricking-out process and its consequences for the establishment and growth of tree seedlings in the field are still in their early stages. In this study, root deformation did not affect the RGR of seedlings of the typical pioneer tropical tree *S. multijuga* 64 days after pricking out and subjected to different light conditions for 43 days. However, root deformation induced significant decreases in RGR_p, which may have



affected the LMR₂, D/RV, and H/RV. From a practical point of view, our results show that root deformation caused by errors during the pricking-out process potentially makes tree seedlings more susceptible to adverse factors in the field, which can lead to reduced survival rates and growth of these seedlings. Other studies exploring the adverse and interactive effects of root deformation and environmental stress are necessary. Such studies may help to elucidate the endogenous and exogenous factors affecting tree seedling mortality in forest restoration programs and may lead to improved technical procedures for tree seedling production in tropical forest nurseries.

5. CONCLUSIONS

The S. multijuga seedlings had their growth rates severely affected in the PAR values by about 1 mol of photons m⁻² day⁻¹, but root deformation did not affect the relative growth rates of the entire plant for 43 days. Instead, plants with root deformation showed a significant decrease in the relative growth rate based on the dry root mass. Changes in root growth reduced biomass allocation to the roots and increased biomass allocation to the stems and leaves. The interactive effects of low light availability and root deformation affected the allocation of leaf biomass at low light availability values. From a practical point of view, root deformations caused by pricking-out errors in forest nurseries can lead to the production of low-quality S. multijuga seedlings, which present less competitive capacity under a variety of light conditions.

6.ACKNOWLEDGMENTS

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